



US008865186B2

(12) **United States Patent**  
**Ichtchenko et al.**

(10) **Patent No.:** **US 8,865,186 B2**  
(45) **Date of Patent:** **\*Oct. 21, 2014**

(54) **GENETICALLY ENGINEERED CLOSTRIDIAL GENES, PROTEINS ENCODED BY THE ENGINEERED GENES, AND USES THEREOF**

(58) **Field of Classification Search**  
USPC ..... 435/69.1, 325, 320.1; 424/234.1  
See application file for complete search history.

(75) Inventors: **Konstantin Ichtchenko**, Brooklyn, NY (US); **Philip A. Band**, West Orange, NJ (US)

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(73) Assignee: **New York University**, New York, NY (US)

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(\* ) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

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This patent is subject to a terminal disclaimer.

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(21) Appl. No.: **13/252,177**

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(22) Filed: **Oct. 3, 2011**

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(65) **Prior Publication Data**

US 2012/0021002 A1 Jan. 26, 2012

*Primary Examiner* — Albert Navarro

*Assistant Examiner* — Ginny Portner

(74) *Attorney, Agent, or Firm* — LeClairRyan, a Professional Corporation

**Related U.S. Application Data**

(62) Division of application No. 12/762,909, filed on Apr. 19, 2010, now Pat. No. 8,044,188, which is a division of application No. 11/284,930, filed on Nov. 22, 2005, now Pat. No. 7,785,606.

(60) Provisional application No. 60/630,175, filed on Nov. 22, 2004.

(51) **Int. Cl.**

**A61K 38/00** (2006.01)

**C12N 9/52** (2006.01)

**C07K 14/33** (2006.01)

**A61K 39/08** (2006.01)

**A61K 39/00** (2006.01)

(52) **U.S. Cl.**

CPC ..... **C12N 9/52** (2013.01); **A16K 2039/6037** (2013.01); **A61K 39/00** (2013.01); **C12Y 304/24069** (2013.01); **C07K 14/33** (2013.01); **A61K 39/08** (2013.01)

USPC ..... **424/234.1**; **424/239.1**; **530/350**

(57) **ABSTRACT**

The present invention relates to an isolated Clostridial neurotoxin propeptide having a light chain region, a heavy chain region, where the light and heavy chain regions are linked by a disulfide bond, and an intermediate region connecting the light and heavy chain regions. An isolated nucleic acid molecule encoding a Clostridial neurotoxin propeptide is also disclosed. Also disclosed is an isolated, physiologically active Clostridial neurotoxin produced by cleaving a Clostridial neurotoxin propeptide, a vaccine or antidote thereof, and methods of immunizing against or treating for toxic effects of Clostridial neurotoxins. Methods of expressing recombinant physiologically active Clostridial neurotoxins are also disclosed. Also disclosed is a chimeric protein having a heavy chain region of a Clostridial neurotoxin and a protein with therapeutic functionality. A treatment method is also disclosed.

**29 Claims, 18 Drawing Sheets**



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FIG. 1A

Light Chain

BoNT A MEFVNKQFNFKDPVNGVDIAVYIKIPNA-GMQPVKAFKHNKIWVPERDTE-TNPEEGDLNPPPEAKQVVSYYDSTYLS  
 BoNT B MEVTTNNFNNDPFDNNNIMMEPPFARGTGRIYKAFKIDRIWVPERYTFGYKPEDFN-KSSGIFNRDVCEYDPPDYLH  
 BoNT C MEITNNFNYSDFDKNKILYLDLTLNLANEPKAFRITGNINWVPERDFSRKSNPNLWK--PPRVTSPK-SGYDPPYLS  
 BoNT D MTWPYKDFNYSDFVNDNDILYLRIEQNKLIITPVKAFMTQINWVPERFSSDTNPSLSK--PPRPTSKYQS--YDPSYLS  
 BoNT E MEK-INSFNNDPVDNDRITLYIKPG--GCQEFYKSNMKNINWVPERNVIGTTPQDFH--PPTSLKNGDSSYYDPPYLQ  
 BoNT F MEVVINSFNNDPVDNDDITLYMQIPYEEKSKYKAFEMRNWVWVPERNTIGTDPDFD--PPASLENGSSAYDPPYLT  
 BoNT G MEVNINKFNNDPFDNDDIMMEPPFNDPFGPTYYKAFRIDRIWVPERFTYGFQDQFNA-STGVFSKDYVEYDPTTYLK

BoNT A TDNEKDNVLRGVTKLFEIRIYSTDLGRMLLTSIVRGTPFFGSGSTIDTELKVIDINCINVI---QPDGSYRSEEL--NAYVI  
 BoNT B TNDKRNIFLQTMKLFNRITKSKPLGKLELMTINGTPYLGDRRVLEEFENTNIAASYVWVKLISNPFGEVERKKGTANLTIY  
 BoNT C TDSKDPFFREIKLFRKINSREIGELIYRLSTDPFGNNTPEINTEDFDYDFNSVDVTRQGNHWVYTGSNPSVLIY  
 BoNT D TDEQKDTFLKGIKLEKRIINERDIGKRLINYLTVGSEFGDSSTPEDTEFFRHTENIAYEKFNQSWKYVNIIPSVLIF  
 BoNT E SDEEKDRFLKIVTINRINNNLSGGILLEELSKANPYLGNNDNTPDNOFHIGDA-SAVEIKFNSGSD---ILLPRVIM  
 BoNT F TDAEKDRYLTATIKLFRKINSNPAGEVLLQETSYARPYLGNERTINEHPVIRTTSVNINISSTN---VKSSILNLVVL  
 BoNT G TDAEKDRFLKTMKLFNRINSKPSGQRLLDMIVDAPYLGNASFPDRFAANVANVSINKIYQPAEDQIKGLMNTLTIY

BoNT A GFSADTIQFEC-----KSFGEVNLNLRNGYGSTQYIRESPDFFGFEESLEYDTNPLLGAGKFAIDPAVTLAHLIHAHG  
 BoNT B GEGPVLNENET-----IDIGIQNHFAISREGFGGIMQMKCEPEYVSVENNVQENKASIFNRRGYFSDPAIIMLHHLIHYLH  
 BoNT C GERENTIDPETS---TFKLTNNTFAAQEGFGALSISISPRFMLTYSNATNDVGEGRFSKSEFCMDPIIIMLHHLNHAHE  
 BoNT D GELENTLDYITAS---LTLQGQSNPSEFEGFGLSLKVAPEFLTFSQVTSNQSAAVLGKSIFCMDPIVIALMHLHLSLH  
 BoNT E GAEFDL--FETNSNLSR--NNYMPSNHGFSGIAIVTESPEYSFRFNDNSM-----EPIQDPAITLMLHLIHSLE  
 BoNT F GAGPDIFENSYPVRKLDMSGGVYDPSNDGFGSINAVTESPEYETFDISGG---YNSSTESIADPAISLAHLIHALH  
 BoNT G GEGPVLSDNET-----DSMIMNGHSFISEGFGARMNIRECPSCLNVENNVQENKDTSIIFSRRAYFADPAITLMLHLIHYLH

BoNT A RLYGIAI-NPNRVFKVNTNAYYEMSGLEVSFEELRTFGGDAKFDLSLQENEFRLYYNKFKDIASTLNKAKSI--VGTTA  
 BoNT B GLYGNKV-DDLPVPEK-K-FMQSDAIQAEELYTFGGDPSITTPSTDKSYDQVLRQNFRIVDRLNKLVLCI-SDPNI  
 BoNT C NLYGHAIPNDQTISSVTSNIEYSQYNVKLEYAEIYAFGGPTIDLIPKSARKYFEKALDYVRSIARLNSITTANPSSFNK  
 BoNT D GLYGNIPSDKRIREQVSEGFSDGPNVQFEELYTFGGDVEITPQIERSQLREKALGHVKDIARLNNINKTIPSSWIS  
 BoNT E GLYGAKGITTKYITITQKQPLITNIRGTN-TEEFLETFGGDLNITTSQSNLYTNLADYKIKSLSKXQVS----NP  
 BoNT F GLYGARGVTKETIKVKQAPLMIAIK-PIRLEEFLETFGGDLNITTSAMKERTYNNLANVEKIASRLSRVNSAPF---EY  
 BoNT G GLYGNISNLPITPNTKE--FFMQHSDPVQAEELYTFGGDPSVLSPSTDNMNLYNKALQNFQDIANRLNIYSSAQGS--GI

BoNT A SLQYMNVEKEKYLSEDTSGKFSVDKLFKDKLYKMLTEITTEDNPFVKFKVLRKTYLNFDKA-FFKIIVPKVNYTID  
 BoNT B NINIVNKEKDKYKVEVDESEGRYSIDVSEFDKLYSLMFGFETENIAENYKIYTRASYFSDSLPPVYIKNLLDNEIYTIIE  
 BoNT C YLGEYKQKLRKYREVVESSEGYTVNRNKEVELYNELTQIFTEFNIAKLYNVQNRKIYLSNYTPTVTA-NILDDNVYDIQN  
 BoNT D NIDKYNKIFSEKYNEDKQNTGNFVVNIDKFNLSYDLTNVMSEVVYSSQYNVNRTHYFSRHYLPVFA-NILDDNYTIRD  
 BoNT E LNPYNDVFEAKYGLDKDASGIVSVNINKFNDFKRL-YSFTEFDLAKFKQVCRQTYIGQYK-FKLSNLLNDSIYHISE  
 BoNT F DINEYNDYEQWKYGLDKNADGSIVNENKENEIYKRL-YSFTEIDLANKFKVCRNTYFIKYGFLKVPNLLDDDIYVSE  
 BoNT G DLSLYKQIYKKNYDVEVDPNGKYSVDKDKFDKLYKALMFGFETENLAGEVYGIYTRYVSEYELPFIKTERLNDNTIYQNE

Heavy Chain

BoNT A GFNLRNTNLAANFNQNTETINNMNFTLKNFT---GLFFFYRL--LC--VVGIIITSKTRSLDKGYNKALNDLC--FNV  
 BoNT B GFNISKDMEKEYRGONKAIN---KQAYEISKHELAV-YKIQ-MC-KSV-----K--APGIC--FDV  
 BoNT C GFNIPKSNVNLFMGQNLRSNPAL-RVNVNPMYLY--L--FTK--FCHKAI--DGRSL--YNK--FLDCRELLV  
 BoNT D GFNLTNKGFNENSGQNTERNPAL-QLSSSESVVD--L--FTK--VC--L--LTK-----NSR--DDSTC--FVK  
 BoNT E GYNIN--NLYVNRGQNALNP---RIITPITGR--GLV--KIRFC--KNIVSVK-----GIR--KSTC--TEI  
 BoNT F GFNIGMLAV--NRRGQNLKLN---KIIDSPDK-GLVE--KIVKFC--KSVIPRK-----GTK--APPRIC--TRV  
 BoNT G GFNIASKNIKTEFNQNRVAVN---KRAYEISLHELVI-YRFA-MC-KPV--MYKN-----TGK--SEQC--FIV

FIG. 1B

BoNT A **NNWDLFF**SPSEDNFTN**DLNKGEET**TSDFNIEAAEEN**ISLDLI**QQYLT**FNF**DN**EPENISIN**ENSSDIIGQLELMEN**IERFP**  
 BoNT B **DNEDLFFI**ADKNS**SFSD**DLSKNER**EYNA**QSNYIEND**FPINEL**---**ILDTDLISKIE**-**TPSEN**TESLTDFNV-D**VYVY**EKQP  
 BoNT C **KNTDLPI**IGDISDVKT**DIFLRKD**NEE**EV**IYYPD**NVSV**DQV---**ILSKNT**SEEGQ**D**---**DLLY**PSIDSESEIL**EG**-**ENQV**  
 BoNT D **KNNR**LPIVADK**SISQ**EIPENKI**TDE**EVQNYSD**NFSL**DES---**ILDG**QVPIN**PEIV**---**DPL**LPNVNM**PEPL**EG**ERIV**  
 BoNT E **NNGEL**FFVASENS**SYND**DNIN**TPKE**IDD**VT**SNNNY**END**LQV---**ILNF**NSESAP**G**-**LSD**ERLNL**TIQ**ND-**AYI**EKYDS**NG**  
 BoNT F **NNREL**FFVASESS**YN**ENDIN**TPKE**IDD**VT**LN**NNY**RNN**LDE**V---**ILDY**NSE**TI**PQ-**IS**NO**TL**N**TLV**QDD-**SYV**PRYS**NG**  
 BoNT G **NNEDE**LFFI**ANKD**S**SF**SK**D**LAKA**ET**AYN**Q**NN**TI**EN**NF**SI**D**QL---**ILD**N**D**LSS**G**ID-**LP**N**EN**TE**PT**TK**F**DD**IDI**EV**YI**K**S**

BoNT A **NG**--**KK**YEL**D**KY**TM****FH**Y**LR**AQ**E**FER**GK**S**R**IA**L**T**NS**V**NE**AL**N**PS**R**V**Y**TF**FS**S**D**Y**V**K**V**N**K**A**TE**A**M**L**G**W**E**Q**L**V**Y**D**E**F**D**E  
 BoNT B **AI**--**KK**IF**T**D**EN**TI**EQ**LY**S**Q**TF**LD**IR**DIS**LT**SS**FD**D**ALL**PS**N**K**V**Y**S**FF**S**MD**Y**IK**T**ANK**V**Y**E**AG**L**AG**V**W**K**Q**I**V**ND**F**Y**IE  
 BoNT C **FY**N**R**T**Q**N**V**D**L**NS**Y**Y**LE**S**Q**KL**S**DN**V**ED**FT**FR**S**IE**EA**L**DN**S**A**K**V**Y**T**EP**T**-**L**ANK**V**N**AG**V**Q**GG**L**EL**M**W**AN**D**V**ED**FT**EN  
 BoNT D **FY**D**I**TY**K**V**D**Y**L**NS**Y**Y**LE**S**Q**KL**S**NN**V**EN**IT**LT**TS**V**EE**AL**G**YS**N**K**I**Y**T**EL**PS**-**L**AE**K**V**N**K**G**V**Q**AG**L**EL**N**W**AN**EV**ED**FT**EN**  
 BoNT E **TS**D**IE**Q**H**D**V**N**L**V**F**Y**L**DA**Q**K**V**EG**EN**V**N**L**T**SS**ID**T**AL**LE**Q**PK**I**Y**T**FF**SE**FE**FN**N**K**P**V**Q**A**AL**F**V**S**TI**Q**Q**L**V**D**FT**TE**  
 BoNT F **TS**E**LE**EH**N**V**D**L**N**V**F**Y**L**HA**Q**K**V**EG**ET**N**IS**L**T**SS**ID**T**AL**SE**S**Q**V**Y**T**FF**SE**FE**FN**I**N**K**P**V**HA**AL**F**IS**IN**Q**V**IR**D**FT**TE**  
 BoNT G **AL**--**KK**IF**V**D**G**DS**L**E**Y**L**HA**Q**T**F**S**N**I**EN**L**Q**L**T**NS**L**N**D**AL**R**NN**N**K**V**Y**TF**FS**ST**N**L**V**E**K**AN**T**V**V**G**A**S**L**V**N**V**K**G**V**ID**D**E**SE**

BoNT A **TSEV**ST**DD**KI**AD**IT**I**IP**Y**IG**PA**L**N**IG**N**ML**Y**K**DD**F**V**G**AL**IF**S**GA**V**IL**LE**FI**PE**IA**IP**V**L**GT**AL**V**S**TI---**AN**K**V**L**T**V**Q**TI  
 BoNT B **ANK**SN**MD**KI**AD**IS**LV**IP**Y**IG**LA**L**N**V**GN**ET**AK**GN**EA**FA**BI**AG**S**IL**LE**FI**PE**LI**P**V**V**GA**L**LE**ST**I---**DN**K**K**I**K**IT**I**  
 BoNT C **IL**R**K**D**TD**DK**IS**D**V**SA**TI**IP**Y**IG**PA**L**N**IS**NS**V**RR**GN**ET**EA**FA**VT**GV**IL**LE**FA**PE**FT**IP**AL**GA**R**V**I**Y**SK**V**---**Q**ER**NE**I**K**IT**I**  
 BoNT D **IM**R**K**D**TD**DK**IS**D**V**SV**TI**IP**Y**IG**PA**L**N**IG**NS**AL**R**GN**EQ**AF**TA**GA**V**AL**LE**GF**PE**FT**IP**AL**GV**TF**Y**SS**I**---**Q**ER**E**K**I**K**IT**I  
 BoNT E **AN**Q**S**TF**Q**KI**AD**IS**LV**VP**Y**IG**LA**L**N**IG**EA**Q**K**GN**FD**AL**EL**L**G**AG**IL**LE**EP**EL**LI**PT**IL**V**ET**IK**S**FL**GS**SD**N**K**K**V**IR**AI  
 BoNT F **AT**Q**S**TF**Q**KI**AD**IS**LV**VP**V**GL**AL**N**IG**EV**Q**EN**FK**EA**FL**L**G**AG**IL**LE**EP**EL**LI**PT**IL**V**ET**IK**S**FL**GS**SEN**K**K**I**RA**I**  
 BoNT G **ST**Q**S**TF**Q**DK**IS**D**V**SV**TI**IP**Y**IG**PA**L**N**V**GN**ET**AK**EN**FK**NA**FA**BI**G**GA**L**LE**EP**EL**LI**PT**IL**V**ET**IK**S**FL**GS**EN**K**K**I**RA**I**

BoNT A **DN**AL**S**K**NE**K**N**DE**V**Y**I**V**T**N**W**L**A**K**V**N**T**Q**ID**L**TR**KK**M**KE**AL**EN**Q**AE**AT**RA**IN**Y**Q**Y**N**Q**Y**TE**ER**N**N**TF---**N**ID**L**SS**Y**LM  
 BoNT B **DN**AL**T**K**NE**K**N**SD**M**Y**GL**I**V**A**Q**W**L**ST**V**N**T**Q**F**Y**TI**KE**G**Y**K**A**N**Y**Q**A**L**RE**IK**Y**R**Y**N**I**Y**S**E**R**E**K**S**HT**NI**--**D**F**N**D**I**NS**Y**LM  
 BoNT C **DN**GLE**Q**IK**R**K**N**K**D**S**Y**W**MM**G**T**W**L**S**R**IE**T**Q**F**N**N**I**SY**Q**Y**MD**S**Y**NY**Q**A**AI**K**AK**ID**LE**Y**K**K**Y**S**G**S**D**K**EN**T**K**S**--**Q**VEN**L**K**N**S**D**  
 BoNT D **EN**CLE**Q**V**K**R**K**N**K**D**S**Y**Q**W**M**V**S**N**W**L**S**R**IE**T**Q**F**N**H**I**NY**Q**Y**MD**S**Y**Q**A**DA**I**K**AK**ID**LE**Y**K**K**Y**S**G**S**D**K**EN**T**K**S**--**Q**V**EN**L**K**N**S**D**  
 BoNT E **NN**AL**K**ER**DE**K**W**KE**V**Y**S**F**I**V**S**N**W**M**TK**IN**T**Q**F**N**K**R**KE**Q**Y**Q**A**IQ**N**Q**V**NA**IK**T**IE**S**K**Y**NS**Y**T**LE**E**K**N**L**T**N**K**Y**D**I**Q**TE**N**L**N**  
 BoNT F **NN**SL**M**ER**ET**K**W**KE**I**NS**W**I**V**S**N**W**L**TR**IN**T**Q**F**N**K**R**KE**Q**Y**Q**A**I**Q**N**Q**V**DA**IK**TV**IE**Y**K**Y**N**Y**T**S**D**ER**R**L**E**SE**Y**NN**I**N**I**R**E**L**N**  
 BoNT G **SN**AL**K**K**D**Q**K**N**T**D**M**Y**GL**I**V**S**Q**W**L**ST**V**N**T**Q**F**Y**TI**KE**R**MY**NA**L**N**NS**Q**A**I**E**K**IE**D**Q**Y**N**Y**S**E**D**K**M**Y**NI--**D**F**N**D**I**D**F**RL**N**

BoNT A **ES**IN**K**AM**IN**IN**K**FL**N**Q**CS**V**S**Y**L**M**N**S**M**I**Y**G**V**K**R**L**ED**DA**S**L**K**DA**L**L**K**Y**I**D**N**R**GT**-**L**I**G**Q**V**D**R**L**D**K**V**N**N**T**L**ST**D**I**PF**FL**S**  
 BoNT B **EG**IN**Q**AL**DN**IN**N**ING**CS**V**S**Y**L**M**R**K**M**I**PL**AV**E**K**L**L**ED**DN**T**L**K**N**L**NY**ID**EN**K**LY**-**L**I**GS**A**Y**E**K**S**K**V**N**K**Y**L**K**T**IM**P**FD**S**  
 BoNT C **VE**I**SE**AM**NN**IN**K**FI**RE**CS**V**Y**L**FR**N**ML**PK**Y**IDE**L**N**EP**DR**N**TK**AK**L**IN**L**ID**-**SH**N**I**L**V**G**EV**D**L**K**AK**V**N**S**F**Q**N**I**PF**N**I**F**  
 BoNT D **VK**I**SE**AM**NN**IN**K**FI**RE**CS**V**Y**L**FR**N**ML**PK**Y**IDE**L**N**EP**DL**R**TK**TE**L**IN**L**ID**-**SH**N**I**L**V**G**EV**D**RL**K**AK**V**N**S**EF**EN**TP**EN**I**F**  
 BoNT E **QK**V**SI**AM**NN**ID**R**FL**ESS**I**SY**L**M**R**L**INE**V**K**IN**K**L**RE**Y**DEN**V**AT**Y**L**LN**Y**TI**Q**H**GS**I**-**L**GES**Q**Q**EL**N**S**M**V**T**D**T**NN**S**I**PF**FL**S  
 BoNT F **KE**V**SL**AM**EN**IER**FI**ESS**I**F**Y**L**M**R**L**INE**A**K**V**S**K**L**RE**Y**DE**G**V**KEY**L**LD**Y**ISE**HR**S**I**-**L**GS**N**S**V**Q**EL**ND**L**V**T**S**T**LN**N**S**I**PF**FL**S  
 BoNT G **QS**EN**L**AI**NN**ID**D**FI**N**Q**CS**I**SY**L**M**N**R**MI**PL**AV**K**K**L**K**D**ED**DN**L**K**R**D**L**LE**Y**ID**T**N**E**LY**-**L**L**DE**V**N**I**L**K**S**K**V**N**R**R**L**K**D**S**I**PF**FL**S

Receptor Binding Domain □

BoNT A **KV**Y**D**N**Q**R**L**LS**TE**TE**IK**N**I**NT**S**TL**N**LR**Y**ES**N**H**L**ID**SR**V**AS**K**IN**IG**S**K**V**N**F**D**ED**R**N**Q**I**Q**IF**N---**L**ES**S**K**IE**V**I**L**K**NA**V**  
 BoNT B **IY**EN**D**FL**IE**ME**FN**K**N**SE**IL**NN**I**TL**N**LR**Y**K**D**N**L**ID**SC**Y**G**AK**VE**Y**D**G**WE**LN---**D**K**N**Q**F**K**IT**SS**AN**--**S**K**TR**V**Q**N**Q**N**I**  
 BoNT C **SY**T**NN**S**L**L**K**D**I**INE**Y**FN**N**IND**S**K**L**SL**Q**R**K**N**TL**V**D**TS**G**Y**NA**E**V**SE**EG**D**V**Q**L**NE**IF**PF**DF**ER**IG**SS**GE**DR**G**V**I**V**T**Q**NE**N**V**  
 BoNT D **SY**T**NN**S**L**L**K**D**I**INE**Y**FN**S**IND**S**K**L**SL**Q**N**K**X**AL**V**D**TS**G**Y**NA**E**V**R**V**GD**N**V**Q**L**NT**I**Y**T**W**DF**K**SS**SG**D---**K**TI**V**N**L**N**K**W**L**  
 BoNT E **SY**T**DD**K**L**L**S**Y**EN**K**Y**FK**R**K**SS**S**V**LN**M**R**Y**K**K**D**K**Y**V**DT**SG**DS**N**IN**ING**D**V**Y**K**Y**E**TK**N**K**N**Q**F**GI**YN**--**D**L**K**SE**V**N**S**Q**ND**Y**I**  
 BoNT F **SY**T**ND**K**L**L**L**Y**EN**K**Y**K**K**D**NS**TL**D**M**R**Y**EN**K**K**FI**DT**SG**Y**S**N**IS**ING**D**V**Y**I**Y**ST**N**R**N**Q**F**GI**Y**SS**--**K**P**SE**V**N**IA**Q**ND**I**  
 BoNT G **LY**T**K**D**T**IL**I**Q**V**EN**N**I**SN**I**SS**NA**L**LS**SY**R**G**R**L**ID**SS**GY**G**AT**N**V**GS**D**V**IF**ND**I**G**N**G**Q**F**K**T**NN**SEN**--**S**N**T**AR**G**SK**T**V**V**

FIG. 1C

BoNT A YNSMVEFSFVWIRIPKVFNSISL--NNEYTIINCMEVN-SGWKSLNYGELIWTLDQTQEKDRVVFVNSQMINISDY  
 BoNT B FNSVFLDFSVFVWIRIPKVKNDGIQNIHNEYTIINCMEVN-SGWKSLRGNRIWTLDIINGKTRVFFVYNIREDISEY  
 BoNT C YNSMVEFSFVWIRIPKVFNSISL--GYTIIDSVKNN-SGWSIGIISNPLVFTLKQEDSEQVINSYDISNAPGY  
 BoNT D YSAIVENSSVFWIRIPKVFNSISL--NNEYTIINSIEQN-SGWKLCIRNGRIWTLDQVNRKYSLIFVYSELSHTGY  
 BoNT E YDNKVFNFSSVFWIRIPKVFNSISL--VNEYTIINCMEVN-SGWKSLNHNRIWTLDQVNRKYSLIFVYSELSHTGY  
 BoNT F YNGRQNFSSVFWIRIPKVFNSISL--NNEYTIIDGIRNNSGWKSLNHNRIWTLDQVNRKYSLIFVYSELSHTGY  
 BoNT G YDSMFDNFSVFWIRIPKVFNSISL--NNEYTIINCMEVN-SGWKSLNHNRIWTLDQVNRKYSLIFVYSELSHTGY

BoNT A INRWIEVTTIINRINNSKIYINGRLIDQKPISNLGNHASNNIIFKLDGCRDT-----HRYITIKYFNIFDKELNEIXE  
 BoNT B INEWFVTTIINRINNAKIYINGRLIESNTDKDIREVIANGELIFKLDGDIRT-----QFIYMKYFSTFNTLSQSN  
 BoNT C -NKWFFVTITNMMGMMKIYINGRLIDTIKVKELTCTNFSTITTEINKIPDGLITSDSDINMMWRDFTYFAKLDGKD  
 BoNT D TNKWFVTTITNMMGMMKIYINGRLKQSQKIEDLDEVKLDKTIVEGIDENIDE-----NQMLWRDFTYFAKLDGKD  
 BoNT E INKWFIEVTTITNDRIGDSKIYINGRLIDQKSIINLGNHVSNDNILEKIVNCSYH-----RYIGTRYFNFEDKELDETE  
 BoNT F INKWFIEVTTITNDRIGDSKIYINGRLIDEKSIINLGNHVSNDNILEKIVGNDT-----RYVGRYFNFEDTGLKTE  
 BoNT G INKWFIEVTTITNDRIGDSKIYINGRLKXSEKINLDRNSNDIDFKLINCTDT-----TKFVWIKDFNIFGRLNATE

BoNT A KDLVDNQNSGILKDFWGDYLYDQKPYVYMLNLYDPNKVVDVNVVGIKRYMYLKGPR-GSVMTN-IVLWSS----LYRG  
 BoNT B EERFYKIQSYSEYLRDFWGNPLMWNKYYMFMAGNNSYKLLKDSFVG-ELT-RSKYNQNSK-VINYRD---LYIG  
 BoNT C NIIENSLQYTVVVDYWGNDLRWNKYYMVNI---DYLNR-----YMYANS-ROIYFNTRR---NND---FNBG  
 BoNT D NIIIVEGQILRNVIKDYWGNPLKEDTEYIIND---NYIDR-----YTAPE-SNVLVLRV-VPDRSK---LYIG  
 BoNT E IOTLVSNEPNTNIIKDFWGNLYLVDKEYYLLNVLKPNFIDRRKDSL---SINNIRSTILLANR-----LYSG  
 BoNT F IETLVSDEPDPSILKDFWGNLYLVDKEYYLLNLLRDKSTQNSN---FLNINQOR-GVYQKPN-IFSWTR---LYIG  
 BoNT G VSSLVWISSTNTLKDFWGNPLRVDYQYLLFNQGMQNIYIK-----VFSKASMGET---APRTVFNNAAINYQNLVIG

BoNT A YFIIIRKYSAGN--KDNIVRNNDRVYIIV-VVKNKERYL-----ATNLSQAGV---EKILSALEIPDVGNLS----QV  
 BoNT B EYFIIRKNSNSQS--NDDIVRKEIYIYLDLDF-FNLNQEFV-----YTYKYFK-KEEKLFAPISDSDEFYNTI---QI  
 BoNT C YNIIIRIRIGNT--NDTRVRGGIILYFDM-TINNAIYFMKNEIMYADNHST---EDIYAIGLRE-----QT  
 BoNT D NPIITISVSDKNP--YSRLNGDNIIILHM-LYNSRKYMIIRDTEIYATQGG---ECSQNCVYALKL-----QS  
 BoNT E IIVKIQRVNNSST--NDNLVRKNDQVYIIVFVASKTHLFP-----YADTATNK--EKTIKISSSGNRIN-----QV  
 BoNT F VEVIRKNGSTDISNTDNFVRKNDLAIYIIV-VDRDVEYRL-----YADIS-IAKP-EKIKLIRTSNSNLSLGL---QI  
 BoNT G LRFILIKASNSRNINNDNIVREGIYIYIIMIDNISDESYRV-----YVLVNS--K--EIQTLFLAPINDPTFYDVLQI

BoNT A VVMK-----SKNDQGITNKCKNINL-----QDNMGND-IGFIFGHQFNNI-----AKLVASNNWVNRQI--ERS  
 BoNT B KEYD-----EQPTYSC--QLL---FKK--DESTEIGLIGIHRFYESGI-VFEEYKDYFCISKVYLK---EVK  
 BoNT C KDINDNIFQIQPMNNTYYAS--QIFKSNFN--GEN---ISGICSIG-----YRFRLLGGDNV--RHNVLVPT  
 BoNT D NLGNYGIGIFSIKNIIVSKNKYC--SQIF--SSFR---EN---TMLADI-----YKPWFSSFKNA---YT---PV  
 BoNT E VVMN-----SVGNNTMN-----FKNNNGNN---IGLLGFKA-----DTVYASTWY---YTHMR  
 BoNT F IYMD-----SIGNNTMN-----EQNNNGSN---IGLLGFHS-----NNLVASSWY---YNNIR  
 BoNT G KKY-----EKTTYNC--QIIC-----EKDKXTFGLFEGIKFVKDYGYVWDTYDNYFCISQWYLRNSENIN

BoNT A SRE-----LGSWEFIPVDDGEGRPL  
 BoNT B RKPYNLK-----LGCNWOEIPKDEGWE  
 BoNT C VKQGNYSALLESSTSTHWGVEVSE  
 BoNT D AVENYETKLL--STSEFWKFTSRDPGWVE  
 BoNT E DFN-----SNGCFWNEFSEEGWQNK  
 BoNT F KNTS-----SNGCFWSEFSEKHWQEN  
 BoNT G XLR-----LGCNWOEIPVDEGWE

FIG. 2A

Light Chain



BoNT A NPFVVKQFNITKDPVNGVDIAIKIENA-GQMVPKAKKIKNNKIWVPERDIF-TNPEEGDLNPPPEAKQVPVSYDSTLST  
 BoNT B NPVTENNENYNDPIDNNIIMNEPFAFGTGRYKAKKIKTDRIWIIPERTTFGTKPEDFN-KSSGIFNRDVCYEDDPIINT  
 BoNT C MPEITANNFNYSDPVNDKNIILDTHELTLANEPEKARITGNIWVLEDFSRNSNENLNK--PFRVTSK-SGYDPIYLSLST  
 BoNT D MFWPKDENISDPVNDNDILRLRIQNKLIITPVKAMITQNIWVLEDFSRNSNENLNK--PFRVTSKQS-YDPSYLSLST  
 BoNT E MPK-INSFNNDPVNDRTILYIKPG--GQEFYKSNINNNKIWVLEPERNVIETTPQDFH--PPTSLKNGDSSYDPIYLLQS  
 BoNT F MPVVINSFNNDPVNDDTILYMQIYEKSKKYKAEINRNVWVLEPERNVIETTPQDFD--PPASLENGSSAYDPIYLLT  
 BoNT G MPVNKKNFNNDPIDNDDIIMNEPNDPFGTYKAEIKIDRIWIIPERFVIGFQDQENA-STGVFSKDVYBYDPIYLLK

BoNT A DNEKDNLYLHGVTKLEFRITYSDDLGRMLLSTVIRGIPENGGSSTIDTELKVIDTNCINVI---QPDGYSRSEEL--NLYVIGF  
 BoNT B NDKKNIFNQMTKLFNRKSKPLLEKLMENIINGIPYIGDRRVLEEFNTNIA SVTKKLIENPQEVERKKGIFANLITFGP  
 BoNT C DSDKDPFLREIKLEKRIINSREIGELIYRLSTDIPEFGNNTPTINTEDFDVDFNSVDVKTRQGNWVYKGSITPESVITGFP  
 BoNT D DEQKDTFLRGIKLEKRIINERDIGKLIINLVVGSPEMGDSSSTPDEDFTRHTTNI AVEKFEKSKWVFNITPESVITGFP  
 BoNT E DEEKDRFLAIVKLEKRIINNSGGILLEELSKANPYLGNNTPDNQFHGDA-SAVEIKFNGSQD---ILLENVITMGA  
 BoNT F DAEKDRYLAITKLEKRIINSNPAGEVLLQETSYAKPYLGNHEPTINEFNPVTRTTSVNIKSGTN---VKSSITLNLVVLGA  
 BoNT G DAEKDKFLAITKLEKRIINSKPSGQRLLDMITVDALPYLGNASTPPDKFAANVANVSINKKIIPQAEADQIKGLMNTLITFGP

★★★

BoNT A SADTIQFEC----KSEGHVNLNLRNGIGSTQIIRFSDPFFGFEESLEVDTNPLLGAGKATDFAVTLAHELIRAGHRLY  
 BoNT B GQVLNENET----IDIGIQNFARREGFGIMQMKFCPEYVSVFNKVQENKNGASINRRGYFSDPALIIMHELITVHLGLY  
 BoNT C RENTDPEFS----TFKLTNTPFAAQEGFGALSITISPRFMLTYSKATNDVGEGRSKSECMDEITLIMHELNHAKENLY  
 BoNT D LENTDYDTAS----LTLQGGQSNPSEFGGTLSTLKYAPPELLTSDVTSNQSASVVGKSIKCMDEVALMHELITVHLGLY  
 BoNT E EPDL--FEFNSSNISLR--NHMPSNHGEFSIAHVTSEPTSFRENDSMN-----EITQDPALYIMHELITVHLGLY  
 BoNT F GPDLPENSSIPVKKLMDSGGVDPNDGEGSINIVTSEPEYETENDISGG---YNSTESIADPAISLAHELIRALHGLY  
 BoNT G GQVLSDNFT----DSMIMNGHSPISEGGARMIRFCPSCLNVFNKVQENKDTSESRRAFADPALYIMHELITVHLGLY

★

BoNT A GIAI-NPNRVYKVTNAYIEMSGLEYSVBEELRTEGGDAKFDLSLQENEFRLYYNNKFDIASTLHAKASI--VGTASLQY  
 BoNT B GIKV-DDLPIVPEKK-FFMQSTDAIQABELYTEGGDPSLITPSTDKSYDRIYQNFRTVDRNLKVLVCI-SDPNTNINI  
 BoNT C GIAIPNDQTISSVTSNTEISQINVKLEYAEIYAFGGPTIDITPKSARKYFEKALDYRSTAKRLNSITANPSSPNYIIGE  
 BoNT D GINIPSDKRIIPQVSEGEFSQDGPVQFEELYTEGGDVEIIPQIERSQLREKALGFKDIKRLNINKTIPSSWISNIDK  
 BoNT E GAKGITTKYITQKQNPILITNIRGTN-IEEFLTEGGDNLNITSAQSNITTNLADYKTIASKISKVQVS----NPLLNP  
 BoNT F GARGVYKETIKVKQAPLMAIK-PIRLBEFLTEGGDNLNITSAKKEKYNLNLANTEKIATRLSRVNSAPP---EYDINE  
 BoNT G GIKTISNLPITPNTKE--FFMQHSDPVQABELYTEGGDPSVYSPSTDMNLYNKALQNFQDLNRLNIVSSAQGS--GIDISL

★

BoNT A MNVYKELLLSDTSCKEFSVDKLEDRITKMLTEITTEDNYKFFVNLNKKTLNFDKA-VFKINIVPKVNYTIDGENLR  
 BoNT B YKNRKDKKFFVSDSECKYSIDVSEDKLYKSMFGTETETIENKIKTRASVFSDSLPPVKINLLEDNEIYTTTEGFNIS  
 BoNT C YKQKLIKKRFFVSESSGEVTVRRNKEVELTHELTIQIETETIATAKINNVQRKINLSNVTPTVA-NILDDNVYDQNGFNIP  
 BoNT D YKKIYSEKNTFRKNDTGNFVVIDKENSLSDLNVMSEVYISSQNVNRNTHVFSRYLPEFA-NILDDNYTTRDGENLT  
 BoNT E YKDYFEAKYGLDKDASGIIYSVNIKENDIFKKL-YSETEFDLATKEQVRCRQTIGQYKY-FKLSNLLNDSIVNTISEGNIN  
 BoNT F YKDYQWKYGLDXNADGSIYVNEKRENEYKKL-YSETEIDLANKFKVRCRNTYFIKYGK-IRVFNLLDDDIYTVSEGFNIG  
 BoNT G YKQIYKKNIDFVSDPNCKYSVDKLEDRITKALMFGTETETIENKIKTRISYFESBYLPPITRERLDDNTIYQNEGENIA

Heavy Chain



BoNT A NINLAANENGQWTEINNMNFTLKNFT---GSEFFYKL--LC--VREGIITSSTQSDQGINRCKALNDIC--IRVNN  
 BoNT B DKDMEKEYRGQKAIN----KQAYEISKEHLAV-YKIQ-MC-RSV-----KCK--APGIC--IDVDN  
 BoNT C KSNLNVLMGQNLRSNPAL-KVNPENMLY--L--FTN--FCHKAI-----DGQSL---YNTKCK--TLDCRELLVKN  
 BoNT D NKGFIENSQGWIERNPAL-QKLSSESVD--L--FTN--VC--LRLTK-----NSKCK--DDSTC--IRVKN  
 BoNT E --NLRVNRGQANLNP---RIITPITGR-GLVK--KIRFC-KNIVSVK-----GIRCK--KSIK--FEINN  
 BoNT F NLAV--NNRGQIKLNP---KIIDSIPDK-GLVE--KIVKFC-RSVIPRK-----GTCKK--APPRIC--IRVNN  
 BoNT G SKNLSTENGQKAVN----KEATYEISLHHLVI-VRIA-MC-KPV-MYKN-----TGCKK--SEQC--IRVNN



FIG. 2B

BoNT A WDLFFSPSEDNFTNLLNKGEETSDNIEAAEENISLDLIQQYLLTFNFDNEPENISIDMLSSSDIIGQLELMENIERFPNG-  
 BoNT B EDLEFFADKNFSFSDLLSKNERIEYNQSNYIENDFFINEL---ILDLDLISKIE-LPSENTESLTDFNV-DVPEVTEKQPAI-  
 BoNT C TDLFFIIGDISDVKTDLFLRKDINEEIVYIYPDNVSVDOV---ILSNITSEHGQ-L--DLLYPSIDSESEILPG-ENQVFI  
 BoNT D NRLLPYVADKDISQEIFENKIITDENVQVYSDNFSLDES---ILDGQVPIVPEIV--DPLLPVNMPEPLNLPG-EEIVFI  
 BoNT E GELEFFVASENSYNDNINTPKEIDDVYSNNYENDLDQV---ILNFNSEAPG-LSDKALNLTIQND-AIYPRKDSNGTSD  
 BoNT F RELEFFVASENSYNDNINTPKEIDDVYLNNNYRNLDQV---ILDVNSEIPIQ-TSNQTLNLTIVQDD-SIVPRDSNGTSE  
 BoNT G EDLEFFIANKDSFSKDLAKAETIAYNTQNNTEIENNFISIDQL---ILDNDLSSGID-LPNENTEPFTNFDDIDIPVYIKQSAL-

BoNT A -KKYELDKYTMFBYTRAQEFHGKSRIALTNSVNEALLNPSRVYTFSSDLYKVKVNRATEAAMFLGWEQLVYDFEDEFSEV  
 BoNT B -KKIFDQENTIEQYIYSQTFELDIRDISLTSFFDALLFSNKVYSFESMDIYKTNKVVVAGLEFAGVVKQIVNDFEIEANKS  
 BoNT C NRTQNVYILNSYIYLESQALSDNVEDFTFTRSIEBALDNSAKVYTFEPT-LANKVWAGVQGGLELMDANDVVEDEFTTNIIRK  
 BoNT D DITKYVYILNSYIYLESQALSNVVENITLTSVVEBALGYSNKIYTFELPS-LAEKVNKGVQAGLELNGANEVVEDEFTTNIIMK  
 BoNT E IEQHDVNELVVFFIYDAQVPEGENNVNLTSSIDTALLEQPKIYTFESSEFINNVNKPVQAALVSWIQVVLVDFTEANQK  
 BoNT F IEEHNVVLDNVFFIYEAQVPEGETNISLTSIDTALSSESQVYTFESSEFINNTPVHAALFISWLNQVIRDFTEATQK  
 BoNT G -XIFVYDGDSEFEYLEAQTFESNIENLQLTNSLNDALRNNKVVYTFESTNLVEKANTVWASLEVNKVVGVVDFTESESTQK

BoNT A STVDKIADITTIPIYIGPALNIGMLYKDDVVGALIFSGHVILLEFIPPIATPVIGTFALVSYI---ANKVLTQVTFIDNALS  
 BoNT B NNDKDIADISLIVPIYIGLALNVGNETAKGNENAEPIASISILLEFIPPELIPVVGAFLESYI---DKNKKIKTIDNAIT  
 BoNT C DLDKISDVSATIPYIGPALNISVVRGNTEAFAVTVITILEAPPEFTIPALGAFVYISKV---QERNKIKTIDNCE  
 BoNT D DLDKISDVSATIPYIGPALNIGSALRGNFQARATAVAFVLEGPPEFTIPALGAFVYISSYI---QEREKIKTIDNCE  
 BoNT E STVDKIADISIVVPIYIGLALNIGNEAKGNENADALLEGILLEFPELIPPTILVFTIKSFLGSSDNKKNKVIKATINNAIK  
 BoNT F STVDKIADISLVVPIYVGLALNIGNEVQKGNENARELLGAGILLEFPELIPPTILVFTIKSFLGSSDNKKNKVIKATINNSLM  
 BoNT G STVDKVSIVSIIPIYIGPALNVGNETAKENENAREIGGAIIIEFIPPELIPVIGVFFLESYVG---NKGHIMTISNAIK

BoNT A KNEKNDDEVYKIVVHWLAKVNTQIDLIRKMKKALENQAETRAIINYOYVTEEEKNNINF--NIDDLSSKLNESNKKA  
 BoNT B KNEKNSDMYGLIYVQWLSVNTQFYTIKEGWYKALNTQQAALKEIKRYMNIYSEKESNINI--DFNDNSKLNENGINQA  
 BoNT C QRILKRWKDSYQWVWNLSTRIITQFNNTSYQMYDSLNYQAGAKAKIDLEMKKYSGSDKENIKS--QVENLKNSLDVKISEA  
 BoNT D QRVKKRWKDSYQWVWNLSTRIITQFNNTSYQMYDSLSYQADAKAKIDLEMKKYSGSDKENIKS--QVENLKNSLDVKISEA  
 BoNT E ERDEKRWKDSYQWVWNLSTRIITQFNNTSYQMYQALQNVNNAIHTIESKNSYVTEERNEELNKKYDIKQENELNQQVSI  
 BoNT F ERDEKRWKDSYQWVWNLSTRIITQFNNTSYQMYQALQNVNNAIHTIEYKNNYVTEERNRLESEVNTNINREELNKKVSLA  
 BoNT G KEDQKRWDMYGLIYVQWLSVNTQFYTKERMYNANLNNSQALTEKIDQVNRYSSEDKMNNINI--DFNDIDFKLNQSNILA

BoNT A MININKFLNQCSVSLMNSMIEYGVKRIEDFDASLNDALTKYTYDRGT-LIGQVDRKDKVNNITLSTDIPEQLSRYVDNRQ  
 BoNT B IDMINNPIINGCSVSLMKKMIPLAVEKLLDFDNTLNKLNLYDENKLY-LIGSAEYKSKVNNYKLTIMPDLSTYVNTTI  
 BoNT C MNNINKFTIRECSVYLFKNMLPKVIDELNKFDRNTAKLTLNLD-SENILVGEVDRKAKVNSFQNTIPNIFSTYTNNSL  
 BoNT D MNNINKFTIRECSVYLFKNMLPKVIDELNKFDRNTAKLTLNLD-SENILVGEVDRKAKVNSFQNTIPNIFSTYTNNSL  
 BoNT E MNYIDRLETESSISYLMKLINEVKINKLREYDENVYTLNLYITQEGSI-LGESQQLNSMVTDTLNSIPELSSSTDDKI  
 BoNT F MEHTERLETESSISYLMKLINEAKVSKLREYDEGVYLLDYISEERSI-LGNSVQELNDLVSTLNSIPELSSSTDDKI  
 BoNT G INNIDDLINQCSISYLMNRMIELAVKLLKDFEDNTERDLEIYDTNELY-LDEVNIKSKVNRHLKDSIPDLSTYVNTTI

Receptor Binding Domain 

BoNT A LLSTFTYVYINNTIYTSILNRYEYSELDLSEYASXINIGSKVNFDPDKNQIQLEN--LESXTEVILKNAIVYNSMVEE  
 BoNT B LIEMFNKYNSEILNIIILNRYRDNLDLDSGYGKVEYVDGVLEL--DKNQFKLTSAN--SKLEVTQHQNTIYNSVFLDF  
 BoNT C LKDIINBYFNNINDSKILSLQNRNNTVDTSGYNAIYSEYDGVQINLTFPPEKIGSSGEDRGRVIVTQENIYNSMVEE  
 BoNT D LKDIINBYFNSINDSKILSLQNRNNAVDTSGYNAIYVRVGDNVQINLTYTNDKLSSSGD--KTIYVLLNNTIYSAIYENS  
 BoNT E LLSYFNKFFNNIKSSVLMNRYRDKYVDTSGYDSNININGDVYKIPYTKNNOFGIYN--DKLSEVNIYNDYTYNDKRYNF  
 BoNT F LLYFNKLYNNIKDNSILDMRYENKFDISGYGNSININGDVYIYSTNRNOFGIYSS--KPSVNIYQNDYTYNGRYQNE  
 BoNT G LIQVFNNTIYNSIYSSNAILSLSYRGGRLIDSYGYGATMNVGSDVIFNDLNGQFKLNNSEN--SNITAHQSKFVYDYSYDNE

FIG. 2C

BoNT A STSEWIRIPKYNFNSISL--NNEYTIINCMPNY-SGKKVSLNYGELIWTLQDTQEKQRVVFEXISQMINISDYINRWTFVTT  
 BoNT B SVSEWIRIPKYNNDGIQYIHNETHINCMPND-SGKKFSIRGNRIWTLLDINGKTKSVFEEYNIREDTSEYINRWTFVTT  
 BoNT C SISEWIRIPKYNFNSISL----GTTIIDSVRNN-SGWSIGIISNPLVFTLKQEDSEQSIINISYDISNNAFGY-NKWFVAV  
 BoNT D SVSEWIRIPKYNFNSISL----NETHIISIRGN-SGKKLCIRGNRIWTLLQDVNRKYSLEIDYSISLSHTGYNKKWFVTT  
 BoNT E SISEWIRIPKYNFNSISL----VNEYTIINCMPNDNSGKKVSLNHNETHIWTLQDNAGINQKLANVGNANGISDYINRWTFVTT  
 BoNT F SISEWIRIPKYNFNSISL----NNEYTIIDCIRNNNSGKKVSLNHNETHIWTLQDTAGNNQKLVFNVTQMSISDYINRWTFVTT  
 BoNT G SINEWVRIPKYNNDIQTLLQNETTISCIKND-SGKKVSIKGNRIWTLLDVNAKSKSIFHEYSIKDNIISDYINRWTFVTT

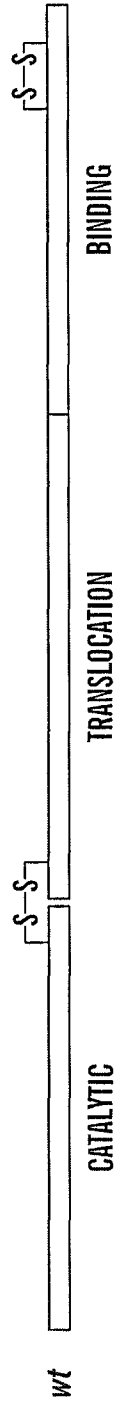
BoNT A TNNRLNWSKIYINGRIIDQKPISNLGNHASNNTLTKLDGCRDT-----HRIIWNHYFNLEDKELNEKEIKKLDYDNQSN  
 BoNT B TNDLNNAKIYINGRIESNTDIRDREVIANGELTKLDGDIQRT-----QFIWMNYESTFNTLSQSNIEERYKIQSY  
 BoNT C TNNMGNWSKIYINGRIIDTIKVKELTGNYFKTIIEINKIPDTGLITSDSDNINMNIHDFNIEAKELDGKDNILFNLSQY  
 BoNT D TNNMGNWSKIYINGRIKQSQKIEDLDEVKLDKTIIEGIDENIDE-----NQMLNIDHFNIEAKELSDNEDINIVVGGQIL  
 BoNT E TNDRLGDSKIYINGRIIDQKSIINLGNTEVSDNIEFKIYVGCST-----RIIGIEYFNLEDKELDETEIQTLYSNEPN  
 BoNT F TNNRLGNRIYINGRIIDQKSIINLGDTEVSDNIEFKIYVGCST-----RIVGIEYFNLEDKELDETEIQTLYSDEPD  
 BoNT G TNDRLGNRIYINGRIKKSSEKILNLDRTNSNDIEKRLINCTDT-----TKFVWTKDFNIEGRELNATEVSSLYNIIQSS

BoNT A SGIKDFWGDYIQYDNPYMLNLDIPNRYVDVHNVGIRCYMILKGP-GSVMTN-IYINSS----LYRGTKPIIKKIYASG  
 BoNT B SEYKDFWGNPIMNKEYYMFNAGNKNSSYIKLKKDSPVE-ELT-RSKYNQNSK--YINRD----LYICEKPIIKRKSNS  
 BoNT C TNYVVDYVGNDRYKKEYYMNVI----DYLNE-----YMYANS-QIIVFNTRE--NND----FNEGYKPIIKRIRGN  
 BoNT D RNVIKDYVGNPKFYDEYIIND----NYIDR-----YIAPESNVLVLR--YDPSK----LYTGNPITIKSVSDK  
 BoNT E TNLKDFWGNLYLDKEYYLLNVLKPNYIIEPKDSTL-----SANNIESTILLAN-----LYSGIYVNTIKRVVNS  
 BoNT F PSILKDFWGNLYLNRYVYLLNLLRDTKXSIQNSN----FLNINQQR-GVYQKPN-IFSNTR----LYTGVEYTIKRNKST  
 BoNT G TNLKDFWGNPFRYDTEQYVLEFNGMNIYIK-----YFSKASMGET--APRTNPNNAIINYQNLVGLRFTIKKASNS

BoNT A N---KDNIVRNNDRVYINNVVVKNKYEL-----ATNASQAGV---EKILSALZIPDVGNLS-----QVVVMK-----S  
 BoNT B QSI--RDDIVRKEIYIILDF-FNLNQEWV-----YTYKYPK-KEEKLILAPISDSDEFINTI--QIKEYD-----  
 BoNT C T---NDTRVRRGGDILTFDM-TINNKANNLFMKNEHMYADNHEST---EDIYAIGLRE-----QTKDINDNIIIFIQ  
 BoNT D NP---YSRILNGDNILHM-LYNSRKYMIIRDTHIYATQGG---ECSQNCVYALKL-----QSNLGNVIGIFPSI  
 BoNT E ST--NDNIVRKNDDQVYIIEVASKTEHLEFL-----YADTATTNK--EKTIKISSSGNREN-----QVVVMN-----  
 BoNT F DISNIDNIVRKNDLAIIINV-VDRDVEYEL-----YADIS-IAKP-EKIKLIRTSNSNNSLG---QIIVMD-----  
 BoNT G RNINNDNIVRREGDYIILNIDNISDESREY-----YVLVNS--K--EIQTQLFLAPINDDPTFYDVLQIKKIY-----

BoNT A KNDQGITNKCKMNL-----QDNMGND-LGFIQFHQNNI-----AKLVASNNYKROH--ERSSRA-----L  
 BoNT B ---EQPTTSC--QLH--EKK--DEESTDEIGLIGTHRYESGI-VPEEYKDFICISKWLK-----EVKRKPYNLK-----L  
 BoNT C PMNNTYYIAS--QIFKSNEN--GER---ISGICSIG-----TYRFRGGDNT-RHNLYVPTVKQGNVYASLIESP  
 BoNT D KNIVSKNYIC-SQIF-SSER---EV---TMLLADI-----YKPRRESFKNA--IT-----PVAVTNKEYTKLI-SI  
 BoNT E ----SVGNNTMN-----FRNNNGNN--IGLLGFKA-----DTVVASTWY---YTEMRDHFN-----SN  
 BoNT F ----SIGNNTMN-----FONNNGGN--IGLLGFHS-----NNLVASSWY---YNNIRKNTS-----SN  
 BoNT G ---EKTITNC--QLC-----EKDXTFGLFEGIKVVDYGYVVDYTDNYFCISQWYLRRESENINKLR-----L

BoNT A GCSWEFIPVDDCWGERPL  
 BoNT B GCNWFQFIPKDEGWTE  
 BoNT C STEWGFVAVSE  
 BoNT D SSFVNFISRDPCWVE  
 BoNT E GCFVNFISEEHGWQEK  
 BoNT F GCFVNFISKEHGWQEN  
 BoNT G GCNWFQFIPVDDCWTE



**FIG. 3A**

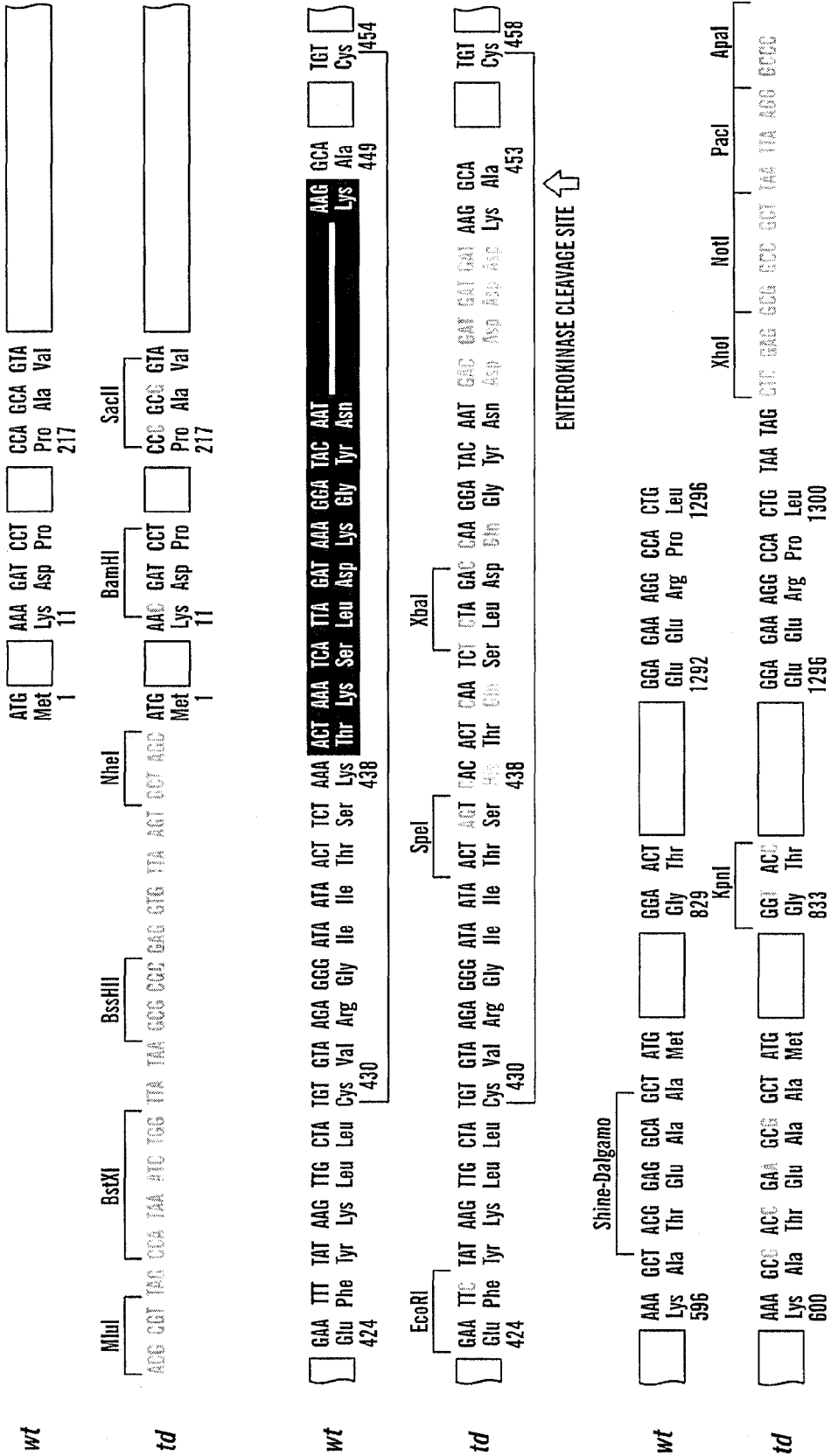
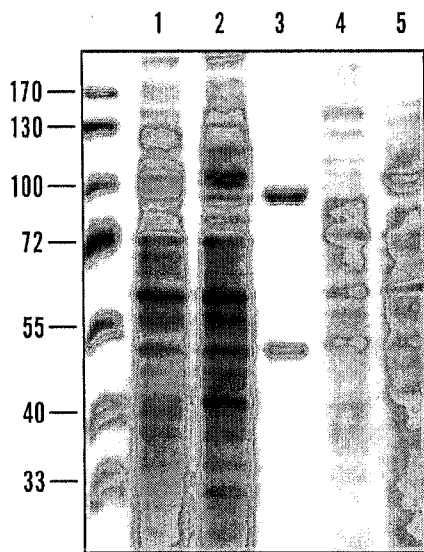
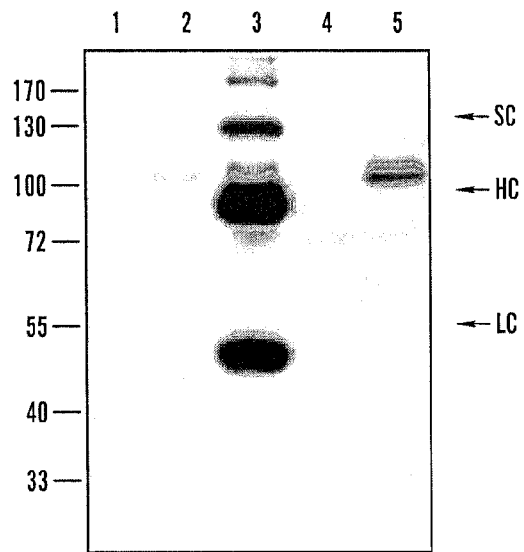


FIG. 3B



**FIG. 4A**



**FIG. 4B**

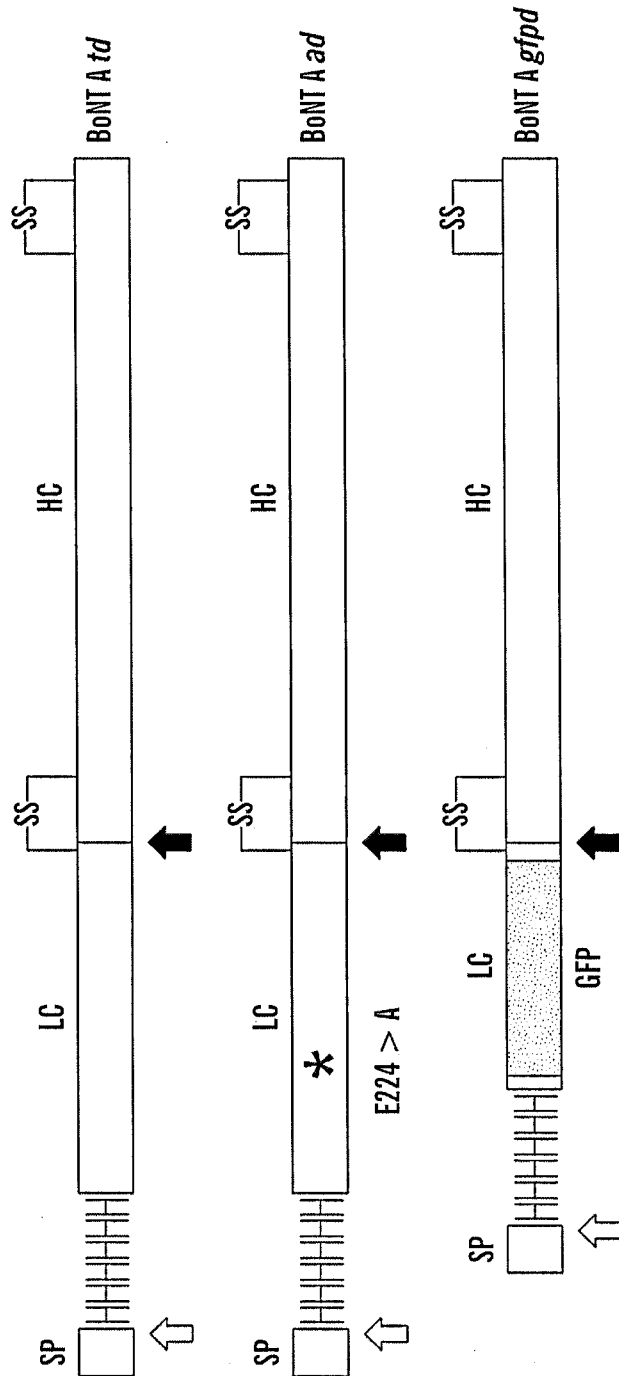
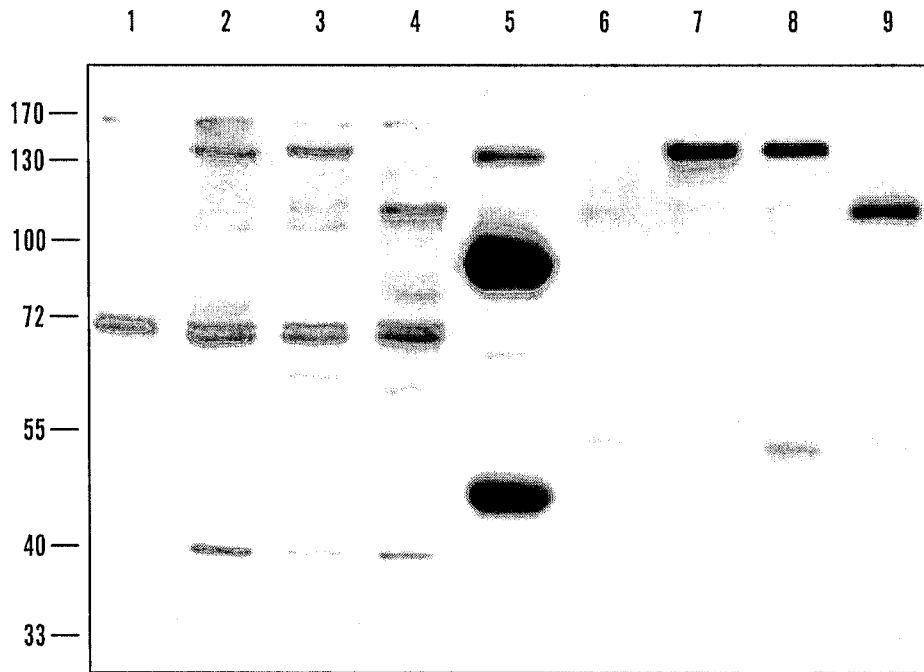
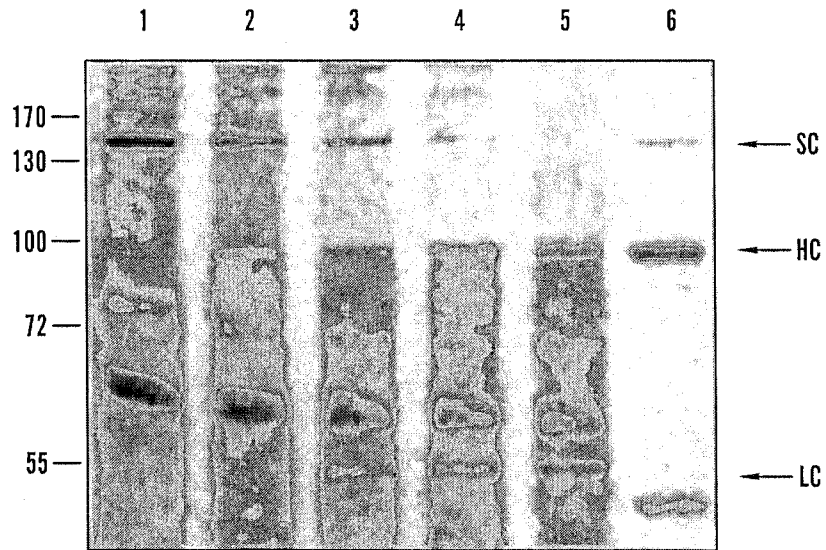


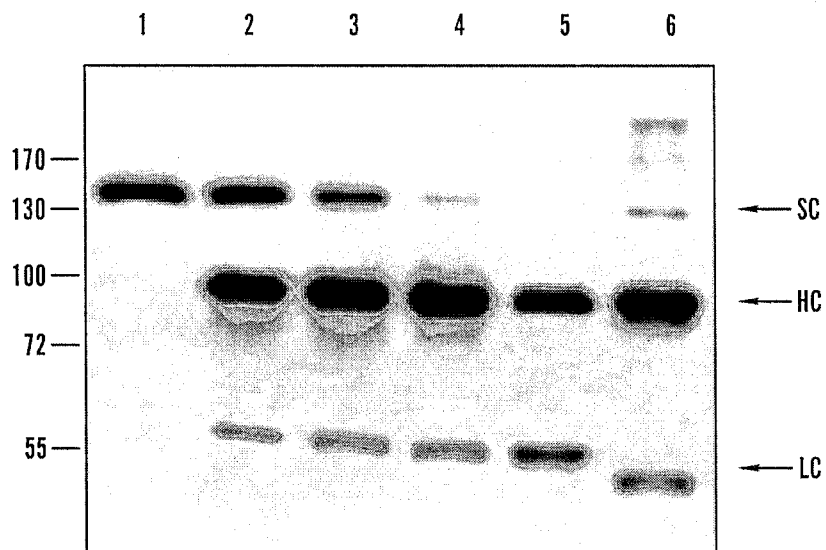
FIG. 5



**FIG. 6**

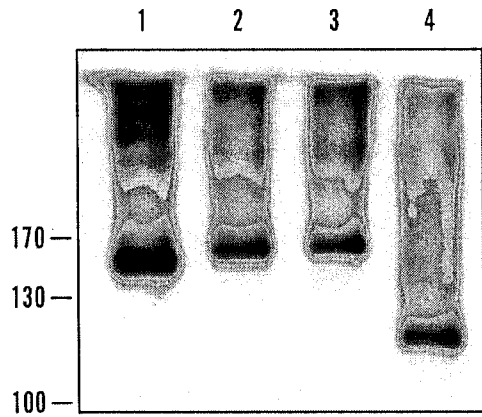


**FIG. 7A**

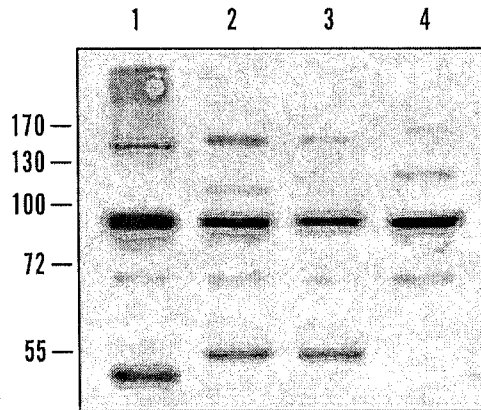


**FIG. 7B**

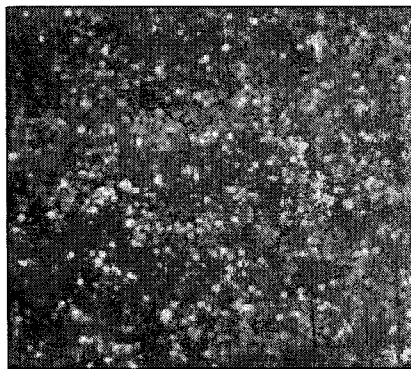




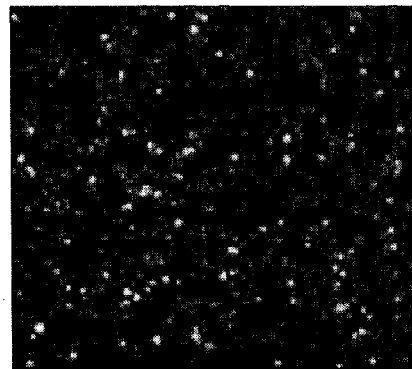
**FIG. 8A**



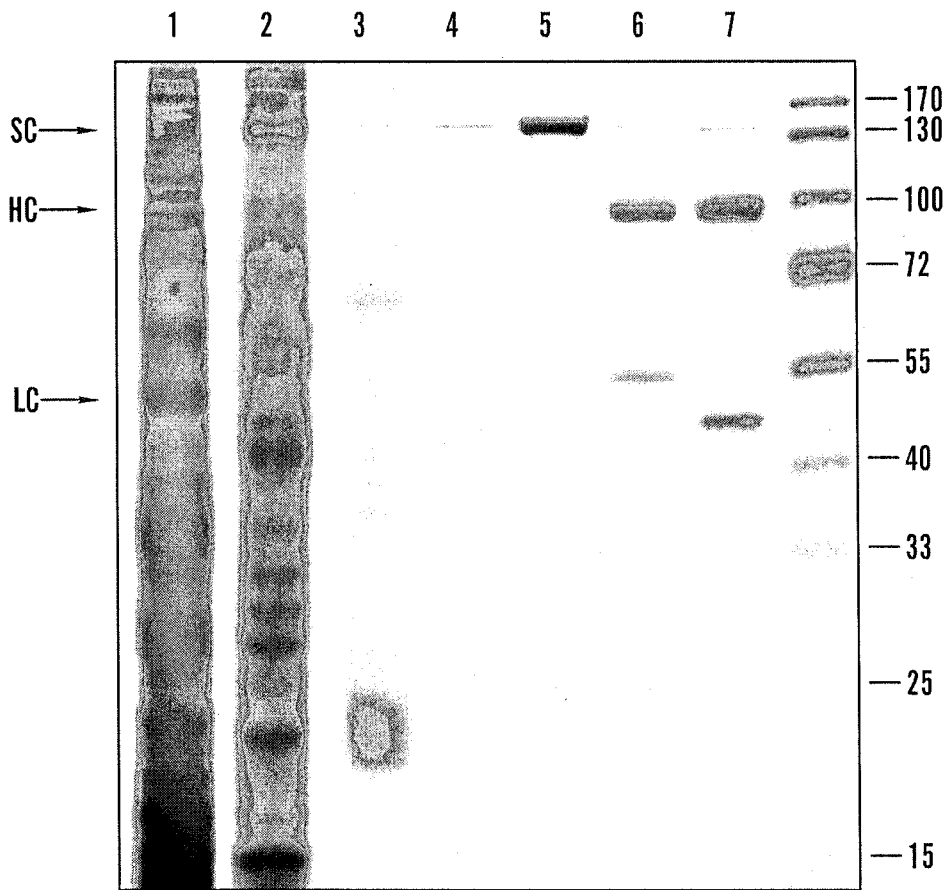
**FIG. 8B**



**FIG. 8C**

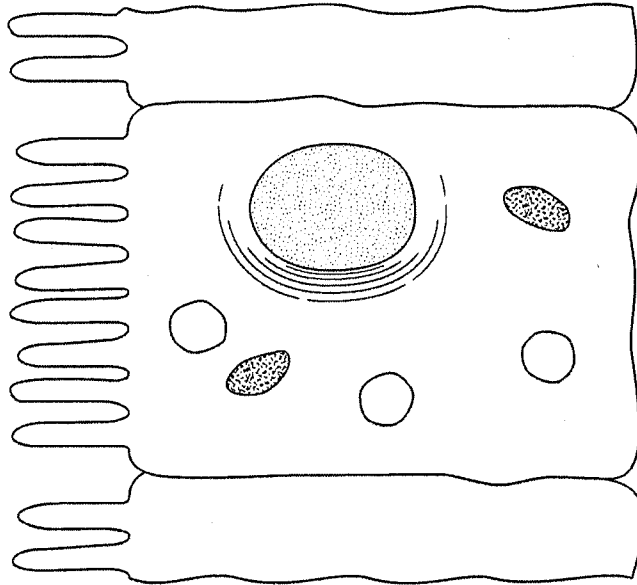


**FIG. 8D**



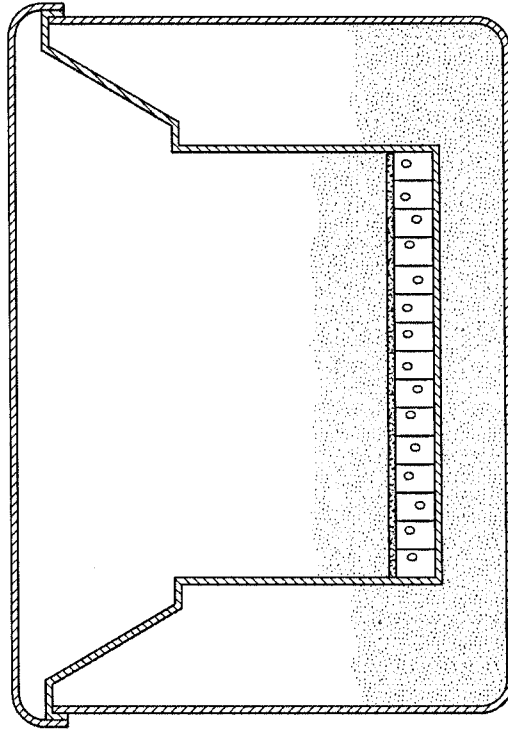
**FIG. 9**

APICAL (MUCOSAL)



BASAL (Serosal)

**FIG. 10A**



TRANSWELL APPARATUS

**FIG. 10B**



FIG. 11B

BoNT A LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 1 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 2 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 3 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 4 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 5 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 6 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 7 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 8 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL  
 Chimera 9 LNKGEEITSDTNIEAAEENISLDLIQQYYLTFNFDNEPENIS IENLSSDIIGQLELMPNIE RFPNGKKYELDKYTMFHYL

BoNT A RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 1 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 2 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 3 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 4 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 5 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 6 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 7 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 8 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII  
 Chimera 9 RAQFEFHGKSRIAL TNSVNEALLNPSRVYTFSSDYVKKVNKATEAAMFLGNVEQLVYDF TDETSEVSTTDKIADITIII

BoNT A PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 1 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 2 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 3 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 4 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 5 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 6 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 7 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 8 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV  
 Chimera 9 PYIGPALNIGNMMLYKDDFVGALIFSGAVILLEFPEIAIPVLGTFALVSYIANKVLT VQTDNALSKRNEKWDVEVYKYIV

BoNT A TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 1 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 2 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 3 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 4 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 5 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 6 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 7 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 8 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS  
 Chimera 9 TNWLAQVNTQIDLIRKKMKEALENQAEATKAIINYQYNQYTEEEKNNINFNIDDLSSKLNESINKAMININKFLNQCQSVS

BoNT A YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 1 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 2 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 3 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 4 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 5 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 6 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 7 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 8 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII  
 Chimera 9 YLMNSMIPYGVKRLLED F DASLKDALLKYYIDNRGTLIGQVDR LKDKVNNTLSTDI PFQLSKYVDNQ RLLSTFTEY IKNII

BoNT A NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 1 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 2 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 3 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 4 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 5 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 6 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 7 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 8 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN  
 Chimera 9 NTSILNLRYESNHLIDL SRYASKINIGSKVNFDPIDKNQIQLFNLES SKIEVILKNAIVYNSMYENFSTSFWIRIPKYPFN

FIG. 11C

BoNT A SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 1 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 2 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 3 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 4 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 5 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 6 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 7 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 8 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR  
 Chimera 9 SISLNNEYTTIINCMEENNSGKWSLNYGEIIWTLQDTQEIKQRVVFKEYSQMINISDYINRWIFVVTITNNRLNNSKIYINGR

BoNT A LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 1 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 2 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 3 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 4 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 5 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 6 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 7 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 8 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML  
 Chimera 9 LIDQKPI SNLGNIHASNNIMFKLDGCRDTRHYIWKYFNLFDKELNEKEIKDLYDNQSN SGILKDFWGDYLYQDKPYYML

BoNT A NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 1 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 2 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 3 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 4 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 5 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 6 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 7 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 8 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA  
 Chimera 9 NLYDPNKYVDVNNVVGIRGYMYLKGPRGSMVTNIIYLNSSLYRGTKFIIKKYASGNKDNIVRNNDRVYINVVVKNKEYRLA

BoNT A TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 1 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 2 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 3 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 4 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 5 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 6 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 7 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 8 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS  
 Chimera 9 TNASQAGVERILSALEIPDVGNLSQVVMKSKNDQGITNKCKMNLQDNNGNDIGFIGFHQFNNAKLVASNWNRQIERS

BoNT A SRTLGCSEFIPVDDGWERPL  
 Chimera 1 SRTLGCSEFIPVDDGWERPL  
 Chimera 2 SRTLGCSEFIPVDDGWERPL  
 Chimera 3 SRTLGCSEFIPVDDGWERPL  
 Chimera 4 SRTLGCSEFIPVDDGWERPL  
 Chimera 5 SRTLGCSEFIPVDDGWERPL  
 Chimera 6 SRTLGCSEFIPVDDGWERPL  
 Chimera 7 SRTLGCSEFIPVDDGWERPL  
 Chimera 8 SRTLGCSEFIPVDDGWERPL  
 Chimera 9 SRTLGCSEFIPVDDGWERPL

**GENETICALLY ENGINEERED  
CLOSTRIDIAL GENES, PROTEINS  
ENCODED BY THE ENGINEERED GENES,  
AND USES THEREOF**

This application is a division of U.S. patent application Ser. No. 12/762,909, filed Apr. 19, 2010, which is a division of U.S. patent application Ser. No. 11/284,930, filed Nov. 22, 2005, which claims the priority benefit of U.S. Provisional Patent Application Ser. No. 60/630,175, filed Nov. 22, 2004, which are hereby incorporated by reference in their entirety.

FIELD OF THE INVENTION

This invention relates to isolated Clostridial propeptides and neurotoxins, vaccines or antidotes thereof, methods of immunizing and treating subjects, isolated nucleic acid molecules encoding Clostridial propeptides and neurotoxins, methods of expression, chimeric proteins, and treatment methods.

BACKGROUND OF THE INVENTION

The Clostridial neurotoxins are a family of structurally similar proteins that target the neuronal machinery for synaptic vesicle exocytosis. Produced by anaerobic bacteria of the *Clostridium* genus, *botulinum* neurotoxins ("BoNT"s, seven immunologically distinct subtypes, A-G) and Tetanus neurotoxin ("TeNT") are the most poisonous substances known on a per-weight basis, with an LD<sub>50</sub> in the range of 0.5-2.5 ng/kg when administered by intravenous or intramuscular routes (*National Institute of Occupational Safety and Health*, "Registry of Toxic Effects of Chemical Substances (R-TECS)," Cincinnati, Ohio: National Institute of Occupational Safety and Health (1996)). BoNTs target cholinergic nerves at their neuromuscular junction, inhibiting acetylcholine release and causing peripheral neuromuscular blockade (Simpson, "Identification of the Major Steps in *Botulinum* Toxin Action," *Annu. Rev. Pharmacol. Toxicol.* 44:167-193 (2004)). BoNT serotypes A, B, and E are considered to represent the most significant threat to military and civilian populations, particularly because they can be aerosolized and delivered by inhalation (Amon et al., "*Botulinum* Toxin as a Biological Weapon: Medical and Public Health Management," *JAMA* 285:1059-1070 (2001)).

Though much work has been done to develop vaccines or antidotes which are effective against poisoning with Clostridial neurotoxins, the effectiveness of available products is limited because the available inactivated toxin preparations do not optimally mimic the native toxin. No therapeutic antidotes or vaccines have been approved for widespread use, though some preparations are available for limited use under specific circumstances. The NIAID Biodefense Research Agenda has identified the development of countermeasures against Clostridial neurotoxins as one of its most pressing goals (*National Institute of Allergy and Infectious Diseases*, "NIAID Biodefense Research Agenda for CDC category A Agents" NIH Publication #03-5308 (2002)). A prime target is understanding and preventing neurotoxin entry into target cells. Immunological approaches have utilized passive protection via injection of antibodies as antitoxins, or active immunization via vaccination with toxoids, toxins chemically or genetically transformed to render them non-toxic but still immunogenic (Ramon et al., "Sur L'immunization Antitetanique et sur la Production de L'antitoxine Tetanique," *Compt. Rend. Soc. Biol.* 93:508-598 (1925)). Antibody-based anti-toxins are available in limited

quantities, but no protective vaccine against Clostridial neurotoxins has been approved. A pentavalent *botulinum* toxoid (ABCDE), consisting of toxins inactivated by temperature or cross-linked with formaldehyde, is available in limited quantities, and has been shown to induce antibodies in laboratory workers and military personnel (*National Institute of Allergy and Infectious Diseases*, "NIAID Biodefense Research Agenda for CDC category A Agents. Progress Report," NIH Publication #03-5435 (2003)). An inactivated heavy chain toxoid administered by inhalation was found to protect animals against inhaled toxin doses 10<sup>4</sup> times the LD<sub>50</sub> (Park et al., "Inhalational Poisoning by *Botulinum* Toxin and Inhalation Vaccination with Its Heavy-Chain Component," *Infect. Immun.* 71:1147-1154 (2003)). An investigational heptavalent antitoxin (A-G reactive, equine origin) against BoNT is being developed by the U.S. Department of Defense and is now being tested. Initial data demonstrate the general safety of this antitoxin, though it displays some cross-species reactivity in humans. Another investigational BoNT antitoxin is based on a combination of three recombinant monoclonal antibodies, which neutralize BoNT A with a high potency (Nowakowski et al., "Potent Neutralization of *Botulinum* Neurotoxin by Recombinant Oligoclonal Antibody," *Proc. Natl. Acad. Sci. USA* 99:11346-11350 (2002)). Development and testing of human monoclonal antibodies to BoNT B-G is also currently in progress and supported by NIAID (*National Institute of Allergy and Infectious Diseases*, "NIAID Biodefense Research Agenda for CDC category A Agents. Progress Report," NIH Publication #03-5435 (2003)).

Several laboratories are attempting to develop recombinant Clostridial toxin genes or fragments thereof. The Department of Defense has developed a vaccine based on expression of the receptor-binding domain of the BoNT A heavy chain (*National Institute of Allergy and Infectious Diseases*, "NIAID Biodefense Research Agenda for CDC Category A Agents. Progress Report," NIH Publication #03-5435 (2003); Byrne et al., "Purification, Potency, and Efficacy of the *Botulinum* Neurotoxin Type A Binding Domain from *Pichia pastoris* as a Recombinant Vaccine Candidate," *Infect. Immun.* 66:4817-4822 (1998); and Pless et al., "High-Affinity, Protective Antibodies to the Binding Domain of *Botulinum* Neurotoxin Type A," *Infect. Immun.* 69:570-574 (2001)). A similar approach with a recombinant BoNT F fragment expressed in *Salmonella typhimurium* was found to provide partial protection of animals against the toxin (Foynes et al., "Vaccination Against Type F *Botulinum* Toxin Using Attenuated *Salmonella enterica* var Typhimurium Strains Expressing the BoNT/F H<sub>C</sub> Fragment," *Vaccine* 21:1052-1059 (2003)). A catalytically active non-toxic derivative of BoNT A expressed in *E. coli* was reported to induce toxin-neutralizing antibodies and protect animals from a BoNT challenge (Chaddock et al., "Expression and Purification of Catalytically Active, Non-Toxic Endopeptidase Derivatives of *Clostridium botulinum* Toxin Type A," *Protein Expr. Purif.* 25:219-228 (2002)). A catalytically inactive, full-length derivative of BoNT C expressed in *E. coli* was immunogenic in mice, though limitations of this system hinder expression of full-length native and active recombinant toxin (Kiyatkin et al., "Induction of an Immune Response by Oral Administration of Recombinant *Botulinum* Toxin," *Infect. Immun.* 65:4586-4591 (1997)). Rummel et al. ("Synaptotagmins I and II Act as Nerve Cell Receptors for *Botulinum* Neurotoxin G," *J. Biol. Chem.* 279:30865-30870 (2004) ("Rummel I")) and Rummel et al. ("The H<sub>cc</sub>-domain of *Botulinum* Neurotoxins A and B Exhibit a Singular Ganglioside Binding Site Displaying Serotype-Specific Carbohydrate Interaction," *Mol. Microbiol.*

51:631-643 (2004) ("Rummel II"), report full-length BoNT A, B, and G neurotoxins expressed in an *E. coli* from plasmids encoding the respective full-length genes. Rummel I and Rummel II also report several derivatives of BoNT genes. The neurotoxins described in Rummel I and Rummel II are active only at very high concentrations. This is likely due to the fact that the neurotoxins expressed by Rummel I and Rummel II are denatured during expression, extraction, and purification from *E. coli* and achieve low physiological activity of the single chain BoNT propeptide due to improper disulfide bonding. Thus, although Rummel I and Rummel II may in fact have produced full-length recombinant BoNT peptides of serotypes A, B, and G, the properties of the neurotoxins described do not possess native structures and physiological activity.

The widely used *E. coli* expression system may be problematic for some proteins, because the *E. coli* cytosol may not provide the non-reducing environment needed for maintenance of disulfide bridges critical to the native toxin structure (Alberts et al., *Molecular Biology of the Cell*, Third Edition, Garland Publishing Inc., 112, 113, 488, 589). In addition, *E. coli* based expression systems also present practical problems associated with endotoxin removal. These limitations emphasize the importance of selecting an expression system capable of producing recombinant molecules that retain the native toxin structure and biological activity.

Data from multiple laboratories suggest that the C-terminal moiety of Clostridial toxin heavy chains ("Hc"), or the intact heavy chain ("HC") expressed or prepared by reduction/denaturation from native toxins, are functionally altered and therefore require a ~10,000-fold molar excess to delay the onset of toxin-induced paralysis (Li et al., "Recombinant Forms of Tetanus Toxin Engineered for Examining and Exploiting Neuronal Trafficking Pathways," *J. Biol. Chem.* 276:31394-31401 (2001); Lalli et al., "Functional Characterization of Tetanus and *Botulinum* Neurotoxins Binding Domains," *J. Cell Sci.* 112:2715-2724 (1999)). Some of these preparations have been completely inactive in this assay (Daniels-Holgate et al., "Productive and Non-Productive Binding of *Botulinum* Neurotoxin A to Motor Nerve Endings are Distinguished by Its Heavy Chain," *J. Neurosci. Res.* 44:263-271 (1996)). The low efficiency of HC and Hc may be due to either their increased binding affinity to non-productive sites on cells normally mediating toxin trafficking or their conformational differences from the native toxin which results in a low binding affinity for the specific binding sites at the target cells. In either case, incorrect folding, altered post-translational modification, a requirement for the N-terminal portion of the molecule (Koriazova et al., "Translocation of *Botulinum* Neurotoxin Light Chain Protease through the Heavy Chain Channel," *Nat. Struct. Biol.* 10:13-18 (2003)), or multiple other changes, may be responsible for these functionally important deficiencies. These facts suggest that the currently available preparations of BoNT or its derivatives are poor mimics of the native toxin, which may limit their therapeutic potential.

The methods currently available to produce inactivated derivatives of BoNTs as vaccines or antidotes to BoNT poisoning have met with limited success. This can be due to several factors. First, the methods used to inactivate BoNT prepared from Clostridial cultures are harsh, and may alter the toxin's native conformation in ways that may influence its immunogenicity or trafficking and absorption. Second, methods based on producing recombinant toxins have thus far only succeeded in producing either inactive toxin molecules or fragments of its protein domains. In both cases, the recombinant molecules produced are by definition significantly dif-

ferent from native toxin, particularly with respect to post-translational processing and disulfide bonding. Though inactivated toxins and toxin fragments have been shown to be immunogenic, the pool of polyclonal antibodies they generate will include a fraction recognizing epitopes present only on misfolded toxins.

Another area in which Clostridial neurotoxins have been extensively studied relates to their clinical use to treat dystonias, and to temporarily correct aesthetic defects in skin. These indications are specific to the neurotoxins produced by strains of *Clostridium botulinum* (BoTox), because they can be used at extremely small doses to locally paralyze specific muscles and thereby achieve therapeutic goals. All of the current products used for this indication are produced from Clostridial cultures, and there have been no reports of an active BoTox molecule produced using any type of genetic engineering technology.

A further area of interest is derived from the ability of Clostridial neurotoxins to pass undegraded through epithelial barriers via transcytosis, and specifically target nervous tissue. This has led to suggestions that Clostridial neurotoxins can be used to enable oral and inhalational carriers for therapeutic agents that cannot normally be delivered via these routes of administration, and delivery vehicles which can specifically target the peripheral and central nervous system.

The present invention is directed to overcoming these and other limitations in the art.

#### SUMMARY OF THE INVENTION

One aspect of the present invention relates to an isolated Clostridial neurotoxin propeptide. The propeptide has a light chain region, a heavy chain region, where the light and heavy chain regions are linked by a disulfide bond, and an intermediate region connecting the light and heavy chain regions. The intermediate region has a highly specific protease cleavage site which has three or more specific adjacent amino acid residues that are recognized by the highly specific protease in order to enable cleavage.

Another aspect of the present invention relates to an isolated nucleic acid molecule encoding the above Clostridial neurotoxin propeptide as well as expression systems and host cells containing this nucleic acid molecule.

A further aspect of the present invention relates to an isolated, physiologically active Clostridial neurotoxin produced by cleaving the above Clostridial neurotoxin propeptide. The propeptide is cleaved at the highly specific protease cleavage site. The light and heavy chain regions are linked by a disulfide bond.

Yet another aspect of the present invention relates to a vaccine or antidote including the above physiologically active, atoxic, Clostridial neurotoxin produced by cleaving the isolated Clostridial neurotoxin propeptide at the highly specific protease cleavage site. The light and heavy chain regions are linked by a disulfide bond.

Still another aspect of the present invention relates to method of immunizing a subject against toxic effects of a Clostridial neurotoxin. This method involves administering the above vaccine to the subject under conditions effective to immunize the subject against toxic effects of Clostridial neurotoxin.

Yet a further aspect of the present invention relates to a method of treating a subject for toxic effects of a Clostridial neurotoxin. This method involves administering an antidote comprising the above physiologically active, atoxic, Clostridial neurotoxin produced by cleaving the isolated



Clostridial neurotoxin propeptide under conditions effective to treat the subject for toxic effects of Clostridial neurotoxin.

Still a further aspect of the present invention relates to a chimeric protein including a first protein or protein fragment having a heavy chain region of a Clostridial neurotoxin and a second protein or protein fragment linked to the first protein or protein fragment.

Another aspect of the present invention relates to a method of expressing a recombinant physiologically active Clostridial neurotoxin. This method involves providing a nucleic acid construct having a nucleic acid molecule encoding an isolated Clostridial neurotoxin propeptide. The nucleic acid construct has a heterologous promoter operably linked to the nucleic acid molecule and a 3' regulatory region operably linked to the nucleic acid molecule. The nucleic acid construct is introduced into a host cell under conditions effective to express the physiologically active Clostridial neurotoxin.

A further aspect of the present invention relates to a treatment method. This method involves contacting a patient with an isolated, physiologically active, toxic, Clostridial neurotoxin produced by cleaving the above isolated Clostridial neurotoxin propeptide.

The present invention relates to a genetic engineering platform that enables rationale design of therapeutic agents based on Clostridial toxin genes. The genetic engineering scheme is based on a two-step approach. For each Clostridial toxin serotype, gene constructs, expression systems, and purification schemes are designed that produce physiologically active, recombinant Clostridial neurotoxin. This ensures that the recombinant toxin derivatives retain structural features important for developing therapeutic candidates, or useful biologic reagents. Using the genetic constructs and expression systems developed by this paradigm, selective point mutations are then introduced to create atoxic recombinant derivatives. This two-step approach is designed to ensure that the recombinant toxin derivatives retain the immunogenicity, absorption profile, and trafficking pathways of native toxin, allowing the atoxic derivatives to have optimized therapeutic and biological properties. They also enable useful chimeric proteins to be created.

Genetically engineered forms of recombinant toxins which structurally and functionally mimic native toxins are superior to the toxoids currently in development for therapeutic purposes. They provide new approaches which can produce customized toxin derivatives in large quantities, and with mutations specifically targeted to the creation of vaccines and toxin antidotes. By focusing on solving the problems associated with producing recombinant toxins, which are physiologically active, the inactivated toxin derivatives of the present invention have distinct advantages over currently available alternatives. This is particularly true with respect to their immunogenic activity and their ability to compete with native toxin for cellular binding sites.

The methodology described herein has additional scientific and practical value because it provides a broad platform enabling facile manipulation and expression of Clostridial toxin genes. This will facilitate studies of the mechanism of Clostridial toxin action, their intracellular trafficking, and the factors responsible for their ability to transit through specific cell types without activation or toxic consequences. In addition, the BoNT constructs created can provide new tools for delivering specific reagents or drugs via oral or inhalation routes, or specifically into peripheral neurons, and enable their controlled activation at the site of intended action. Other approaches to engineer delivery tools based on chemically modified heavy chains from Clostridial neurotoxins have had limited success, possibly because the methods used to inac-

tivate the toxin interfere with protein spatial structure (Good-nough et al., "Development of a Delivery Vehicle for Intracellular Transport of *botulinum* Neurotoxin Antagonists," *FEBS Lett.* 513:163-168 (2002), which is hereby incorporated by reference in its entirety).

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1A-C show comparative alignment of amino acid sequences of the seven wildtype *botulinum* neurotoxin serotypes, including *Clostridium botulinum* serotype A (SEQ ID NO: 1), *Clostridium botulinum* serotype B (SEQ ID NO: 2), *Clostridium botulinum* serotype C (SEQ ID NO: 3), *Clostridium botulinum* serotype D (SEQ ID NO: 4), *Clostridium botulinum* serotype E (SEQ ID NO: 5), *Clostridium botulinum* serotype F (SEQ ID NO: 6), and *Clostridium botulinum* serotype G (SEQ ID NO: 7). Gaps have been introduced to maximize homology. Amino acids identical in  $\geq 50\%$  of compared sequences are shown in black boxes. Amino acids constituting the active site of the catalytic domain of metalloprotease are marked by stars. Disulfide bridge between neurotoxin cysteine residues of the light and heavy chain are shown as a long horizontal bracket. The amino acid residues constituting the minimal catalytic domain of the light chain are hatched. The first amino acid of the C-terminal part of the protein heavy chain (N872 for BoNT A), constituting receptor-binding domain are shown with the arrow. Amino acids, absent in the mature dichain BoNT A molecule along with the aligned amino acids of the other BoNT serotypes are boxed. The white arrow is positioned at the first amino acid of the neurotoxins' heavy chain.

FIGS. 2A-C show comparative alignment, using the Clustal Program, of amino acid sequences of the seven *botulinum* neurotoxin serotypes, including *Clostridium botulinum* serotype A (SEQ ID NO: 8), *Clostridium botulinum* serotype B (SEQ ID NO: 9), *Clostridium botulinum* serotype C (SEQ ID NO: 10), *Clostridium botulinum* serotype D (SEQ ID NO: 11), *Clostridium botulinum* serotype E (SEQ ID NO: 12), *Clostridium botulinum* serotype F (SEQ ID NO: 13), and *Clostridium botulinum* serotype G (SEQ ID NO: 14), which have been slightly modified in accordance with the present invention. Gaps have been introduced to maximize homology. Amino acids identical in  $\geq 50\%$  of compared sequences are shown in black boxes. Amino acids constituting the active site of the catalytic domain of metalloprotease are marked by stars. Disulfide bridge between neurotoxin cysteine residues of the light and heavy chain are shown as a long horizontal bracket. The amino acid residues constituting the minimal catalytic domain of the light chain are hatched. The first amino acid of the C-terminal part of the protein heavy chain (N876 for BoNT A), constituting receptor-binding domain are shown with the arrow. Amino acids, absent in the mature dichain BoNT A molecule along with the aligned amino acids of the other BoNT serotypes are boxed. The white arrow is positioned at the first amino acid of the neurotoxins' heavy chain. Amino acid residues are modified in comparison with the wild type sequence to restrict trypsin-like proteolysis. Amino acids which constitute the insertion/modification into the wild type amino acid residues and represent an enterokinase cleavage site are also shown.

FIGS. 3A-B illustrate features of the wild type BoNT A protein and gene (wt), and its toxic recombinant derivative (td). FIG. 3A is a schematic representation of the native BoNT A (wt) dimer, illustrating the catalytic (~50 kDa), translocation (~50 kDa), and receptor-binding (~50 kDa) domains. FIG. 3B is a comparison of the nucleotide (SEQ ID NO: 65 (wt) and SEQ ID NO: 66 (td)) and amino acid (SEQ

ID NO: 1 (wt) and SEQ ID NO: 8 (td) sequences of the native BoNT A (wt) and its recombinant toxic derivative (td), as generated in plasmid pLitBoNTA. Sequences common to both the wt and td genes are shown as black letters on a white background, or as white boxes. White letters on a black background represent the amino acids excised from the toxin propeptide to generate the mature wt toxin. The disulfide bonds joining the LC and HC are shown as long horizontal brackets. Grey letters indicate the unique endonuclease restriction sites introduced into non-coding portions of the td DNA sequence and the Shine-Dalgarno region of the wt sequence. All other mutations introduced to modify the construct properties are also shown in grey letters. The de novo enterokinase cleavage site inserted into the td propeptide is shown by an arrow. Amino acids proximal to conceived (wt) or executed (td) mutations are numbered.

FIGS. 4A-B show expression and purification of the toxic derivative of BoNT A (td) in *E. coli*. FIG. 4A shows 8% PAGE stained with Coomassie G-250. FIG. 4B shows a Western blot of the PAG shown in FIG. 4A, probed with polyclonal antibodies raised against the full-length BoNT A toxoid. Samples were treated with  $\beta$ -mercaptoethanol before separation. The protein molecular weight standards are shown to the far left. Lanes 1 and 2 are cleared lysate of *E. coli* transformed with pETcoco2 empty vector (Lane 1) or pETcocoBoNTA (Lane 2). Lane 3 is a purified preparation of native BoNT A used as positive control. Lane 4 and 5 are eluates from the Ni-NTA affinity purification of cleared *E. coli* lysates which have been transformed with pETcoco2 (Lane 4) or pETcocoBoNTA (Lane 5). SC: single chain propeptide. HC: Heavy Chain. LC: Light Chain.

FIG. 5 is a schematic representation of the three recombinant BoNT A derivatives expressed in a baculovirus system. BoNT A td: toxic derivative of BoNT A. BoNT A ad: atoxic derivative of BoNT A. BoNT A gfpd: green fluorescent protein (GFP) derivative of BoNT A. Further modifications introduced into the td sequence depicted in FIG. 3 include the introduction of a signal sequence and a hexahistidine tag (e.g., SEQ ID NO: 45) in front of the first native methionine for affinity purification. The difference between td and ad is a single amino acid substitution, E224>A, in the active center of toxin's catalytic domain. To create BoNT A gfpd, amino acids Tyr<sub>10</sub>-Leu<sub>416</sub> of the native toxin's minimal catalytic domain were substituted with GFP. White and black arrows represent secretase and enterokinase cleavage sites, respectively.

FIG. 6 shows expression of BoNT A derivatives in a baculovirus system by Western blot, probed with polyclonal antibodies raised against full-length BoNT A toxoid. Samples were treated with  $\beta$ -mercaptoethanol before separation. Protein molecular weight standards are shown on the left. Lane 1, 2, 3, and 4: conditioned media from Sf9 cells infected with empty bacmid (Lane 1), or recombinant bacmids derived from pFBSBoNTA (Lane 2), pFBSBoNTAME224A (Lane 3) or pFBSGFPBoNTAHC (Lane 4). Lane 5 is native BoNT A as a positive control. Lanes 6, 7, 8, and 9: eluate after Ni-NTA affinity purification of conditioned media from Sf9 cells transfected with empty bacmid (Lane 6), or recombinant bacmids derived from pFBSBoNTA (Lane 7), pFBSBoNTAME224A (Lane 8), or pFBSGFPBoNTAHC (Lane 9).

FIGS. 7A-B illustrate the concentration of recombinant enterokinase (rEK) required to effect complete cleavage of BoNT A toxic derivative (td) propeptide. FIG. 7A shows 8% PAGE stained with Coomassie G-250. FIG. 7B shows a Western blot of the gel in FIG. 7A, probed with polyclonal antibodies raised against full-length BoNT A toxoid. Samples

were treated with (3-mercaptoethanol before the separation. Protein molecular weight standards are shown on the left. Different amounts of rEK were added to 1  $\mu$ g of BoNT A td in rEK cleavage buffer and incubated at 20° C. for 8 hours. 10% of each reaction mixture was loaded per lane. The number of rEK units added per 1 g of BoNT A td were: no rEK added (Lane 1); 0.05 U of rEK (Lane 2); 0.1 U of rEK (Lane 3); 0.25 U of rEK (Lane 4); 0.5 U of rEK (Lane 5). Lane 6 is the positive control, with 0.1  $\mu$ g of native BoNT A. The recombinant light chain is larger than the control because of construct design.

FIGS. 8A-D show selected features of the recombinant BoNT A derivatives illustrating their native disulfide bonding (FIGS. 8A and 8B), and the use of a signal sequence to increase secretion of the toxin derivative into the culture medium (FIGS. 8C and 8D). FIGS. 8A and 8B show PAGE of the indicated BoNT derivatives run on 10% PAGE gels, followed by Western blotting using polyclonal antibodies raised against full-length BoNT A toxoid. A protein molecular weight ladder is shown on the left. In FIG. 8A, the PAGE was run under non-reducing conditions before transfer to the nitrocellulose. In FIG. 8B, samples were treated with  $\beta$ -mercaptoethanol and run under reducing conditions before transfer to the nitrocellulose for Western blotting. Lane 1: Positive control, purified native BoNT A; Lane 2: BoNT A td cleaved with rEK; Lane 3: BoNT A ad cleaved with rEK; Lane 4: BoNT A gfpd cleaved with rEK. FIGS. 8C and 8D are fluorescent images of the adherent layer of Sf9 cells ( $2.10^5/cm^2$ ) in the SF 900 II medium at 12 hours post-infection (MOI~0.1) with recombinant baculovirus expressing BoNT A gfpd containing the signal peptide for secretion (FIG. 8C), or the control recombinant baculovirus expressing GFP without added signal peptide (FIG. 8D). Emission wavelength 508 nm, magnification factor  $\times 200$ , exposure time 0.1 sec.

FIG. 9 is a BoNT A td purification table of 8% PAGE stained with Coomassie G-250. Samples were separated in the presence of  $\beta$ -mercaptoethanol. Lane 1: concentrated and dialyzed Sf9 medium, loaded on DEAE Sepharose; Lane 2: 100 mM NaCl eluate from DEAE Sepharose; Lane 3: 200 mM NaCl eluate from MonoS column; Lane 4: 60 mM imidazole eluate from Ni-NTA agarose; Lane 5: material, eluted from the FPLC gel-filtration column; Lane 6: material, eluted from the FPLC gel-filtration column and digested with rEK; Lane 7: positive control, purified native BoNT A. Protein molecular weight ladder is shown on the right.

FIGS. 10A-B illustrate a transcytosis assay for polarized cells. Human gut epithelial cells (T-84) or canine kidney cells (MDCK) will be grown subject to conditions that promote differentiation and polarization of the cell monolayer (FIG. 10A). An example of a polarized cell illustrating orientation of the apical membrane toward the top (accessible to medium in the insert) and the basal membrane oriented toward the bottom (accessible to medium in the well) (FIG. 10B). Cells will be grown on polycarbonate membranes coated with collagen in Transwell® porous bottom inserts. The inserts suspend the cell monolayer above the bottom of the well, enabling cells to feed from the top and the bottom, and to be exposed to toxin from the top and the bottom. Cultures grown in this manner differentiate into a polarized membrane with tight junctions.

FIGS. 11A-C illustrate the amino acid sequences of nine BoNT A chimeric proteins containing SNARE motif peptides substituted for alpha-helix domains in the light chain region aligned against the BoNT A ad protein (SEQ ID NO: 8). Chimera 1 (SEQ ID NO: 15) contains the full-length sequence of BoNT A ad with three SNARE motif peptides substituting light chain alpha-helix 1. Chimera 2 (SEQ ID

NO: 16) contains the full-length sequence of BoNT A ad with two SNARE motif peptides substituting light chain alpha-helix 4. Chimera 3 (SEQ ID NO: 17) contains the full-length sequence of BoNT A ad with five SNARE motif peptides substituting light chain alpha-helices 1 and 4. Chimera 4 (SEQ ID NO: 18) contains the full-length sequence of BoNT A ad with three SNARE motif peptides substituting light chain alpha-helices 4 and 5. Chimera 5 (SEQ ID NO: 19) contains the full length sequence of BoNT A ad with six SNARE motif peptides substituting light chain alpha-helices 1, 4, and 5. Chimera 6 (SEQ ID NO: 20) contains the full length sequence of BoNT A ad with four SNARE motif peptides substituting light chain alpha-helices 4, 5, and 6. Chimera 7 (SEQ ID NO: 21) contains the full length sequence of BoNT A ad with five SNARE motif peptides substituting light chain alpha-helices 4, 5, 6, and 7. Chimera 8 (SEQ ID NO: 22) contains the full length sequence of BoNT A ad with seven SNARE motif peptides substituting light chain alpha-helices 1, 4, 5, and 6. Chimera 9 (SEQ ID NO: 23) contains the full length sequence of BoNT A ad with eight SNARE motif peptides substituting light chain alpha-helices 1, 4, 5, 6, and 7.

#### DETAILED DESCRIPTION OF THE INVENTION

One aspect of the present invention relates to an isolated Clostridial neurotoxin propeptide. The propeptide has a light chain region, a heavy chain region, where the light and heavy chain regions are linked by a disulfide bond, and an intermediate region connecting the light and heavy chain regions. The intermediate region has a highly specific protease cleavage site which has three or more specific adjacent amino acid residues that are recognized by the highly specific protease in order to enable cleavage.

In a preferred embodiment, the isolated Clostridial neurotoxin propeptide is from *Clostridium botulinum*. *Clostridium botulinum* has multiple serotypes (A-G). Although the Clostridial neurotoxin propeptides of the present invention may be from any of the *Clostridium botulinum* serotypes, preferable serotypes are serotype A, serotype B, and serotype E.

Common structural features of the wild-type *Clostridium botulinum* neurotoxin propeptides are shown in FIG. 1. These structural features are illustrated using BoNT A propeptide as an example, and are generalized among all *Clostridium botulinum* serotypes. BoNT A propeptide has two chains, a light chain ("LC") of Mr~50,000 and a heavy chain ("HC") of Mr~100,000, linked by a disulfide bond between Cys<sub>429</sub> and Cys<sub>453</sub>. As illustrated in FIG. 1, all seven BoNT serotype propeptides have a light chain region and a heavy chain region linked by a disulfide bond. Two essential Cys residues, one adjacent to the C-terminus of the light chain, and a second adjacent to the N-terminus of the heavy chain are present in all seven BoNT serotypes. These two Cys residues form the single disulfide bond holding the HC and LC polypeptides together in the mature neurotoxin. This disulfide bond enables the mature neurotoxin to accomplish its native physiological activities by permitting the HC and LC to carry out their respective biological roles in concert. The disulfide bond between HC and LC polypeptides in all seven serotypes is illustrated in FIG. 1 by the solid line joining the involved Cys residues. The outlined box in FIG. 1 illustrates the intermediate region defined by amino acid residues Lys<sub>438</sub>-Lys<sub>448</sub> of BoNT A. This intermediate region identifies the amino acids eliminated during maturation of wild-type BoNT A, and believed to be excised by a protease endogenous to the host microorganism. This cleavage event, described infra, generates the biologically active BoNT HC-LC dimer. The outlined

amino acid residues in FIG. 1, representing amino acid residues numbered approximately in the 420 to 450 range for all seven BoNT serotypes, can be considered as a region "non-essential" to the toxins' physiological activity and, therefore, represents targets for directed mutagenesis in all seven BoNT serotypes.

All seven BoNT serotypes contain Lys or Arg residues in the intermediate region defined by the box in FIG. 1 which make the propeptides susceptible to activation by trypsin. Native BoNT A propeptide recovered from young bacterial cultures can be activated by trypsinolysis, with production of intact, S—S bound light and heavy chain. Though multiple additional trypsin-susceptible sites are present in the propeptides, they are resistant to proteolysis due to their spatial positions within the native toxin molecule (Dekleva et al., "Nicking of Single Chain *Clostridium botulinum* Type A Neurotoxin by an Endogenous Protease," *Biochem. Biophys. Res. Commun.* 162:767-772 (1989); Lacy et al., "Crystal Structure of *Botulinum* Neurotoxin Type A and Implications for Toxicity," *Nat. Struct. Biol.* 5:898-902 (1998), which are hereby incorporated by reference in their entirety). A second site in the native propeptide of several BoNT serotypes can be susceptible to trypsin cleavage when subjected to higher enzyme concentrations or incubation times (Chaddock et al., "Expression and Purification of Catalytically Active, Non-Toxic Endopeptidase Derivatives of *Clostridium botulinum* Toxin Type A," *Protein Expr. Purif.* 25:219-228 (2002), which is hereby incorporated by reference in its entirety). This trypsin-susceptible site is located in the region adjacent to the toxin receptor binding domain. This region of the HC peptide is found to be exposed to solvent in BoNT serotypes for which information is available on their 3-D crystal structure (Lacy et al., "Crystal Structure of *Botulinum* Neurotoxin Type A and Implications for Toxicity," *Nat. Struct. Biol.* 5:898-902 (1998); Swaminathan et al., "Structural Analysis of the Catalytic and Binding Sites of *Clostridium botulinum* Neurotoxin B," *Nat. Struct. Biol.* 7:693-699 (2000), which are hereby incorporated by reference in their entirety).

In a preferred embodiment, the propeptide of the present invention has an intermediate region connecting the light and heavy chain regions which has a highly specific protease cleavage site and no low-specificity protease cleavage sites. For purposes of the present invention, a highly specific protease cleavage site has three or more specific adjacent amino acid residues that are recognized by the highly specific protease in order to permit cleavage (e.g., an enterokinase cleavage site). In contrast, a low-specificity protease cleavage site has two or less adjacent amino acid residues that are recognized by a protease in order to enable cleavage (e.g., a trypsin cleavage site).

In all seven BoNT serotypes, the amino acid preceding the N-terminus of the heavy chain is a Lys or Arg residue which is susceptible to proteolysis with trypsin. This trypsin-susceptible site can be replaced with a five amino acid enterokinase cleavage site (i.e., DDDDK (SEQ ID NO: 24)) upstream of the heavy chain's N-terminus, as illustrated for the seven serotypes in FIG. 2. This modification enables standardization activation with enterokinase. In serotypes A and C, additional Lys residues within this region are mutated to either Gln or His, thereby eliminating additional trypsin-susceptible sites which might result in undesirable non-specific activation of the toxin. Trypsin-susceptible recognition sequences also occur upstream of the heavy chain's receptor-binding domain in serotypes A, E, and F. This region's susceptibility to proteolysis is consistent with its exposure to solvent in the toxin's 3-D structure, as shown by X-ray crystallography analysis. Therefore, in serotypes A, E, and F, the susceptible residues

are modified to Asn (FIG. 2). Signal peptides and N-terminal affinity tags are also preferably introduced, as required, to enable secretion and recovery.

In a preferred embodiment, the isolated Clostridial neurotoxin propeptide of the present invention has light and heavy chain regions which are not truncated.

As described in greater detail infra, the isolated Clostridial neurotoxin propeptide of the present invention may include a disabling mutation in an active metalloprotease site of the propeptide. The amino acid residues constituting the minimal catalytic domain of the light chain of the propeptide are illustrated in FIG. 1 and FIG. 2 by hatching. Specific amino acid residues constituting the active site of the catalytic domain of the metalloprotease are marked by stars in FIG. 1 and FIG. 2.

The Clostridial neurotoxin propeptide of the present invention may also possess a non-native motif in the light chain region that is capable of inactivating light chain metalloprotease activity in a toxic Clostridial neurotoxin. Suitable non-native motifs capable of inactivating light chain metalloprotease activity of a toxic Clostridial neurotoxin include, without limitation, SNARE motifs, metalloprotease inhibitor motifs, such as those present in the protein family known as Tissue Inhibitors of Metalloprotease (TIMP) (Mannello et al., "Matrix Metalloproteinase Inhibitors as Anticancer Therapeutics," *Curr. Cancer Drug Targets* 5:285-298 (2005); Emonard et al., "Regulation of Matrix Metalloproteinase (MMP) Activity by the Low-Density Lipoprotein Receptor-Related Protein (LRP). A New Function for an 'Old Friend,'" *Biochimie* 87:369-376 (2005); Maskos, "Crystal Structures of MMPs in Complex with Physiological and Pharmacological Inhibitors," *Biochimie* 87:249-263 (2005), which are hereby incorporated by reference in their entirety), zinc chelating motifs based on suitably positioned sulfhydryl (preferably methionine) and acidic amino acids which become exposed upon binding of the chimeric antagonist to the active LC metalloprotease, and peptide motifs corresponding to the cleavage site on the substrate of LC metalloproteases, including transition state analogs of said cleavage site (Sukonpan et al., "Synthesis of Substrates and Inhibitors of *Botulinum* Neurotoxin Type A Metalloprotease," *J. Peptide Res.* 63:181-193 (2004); Hayden et al., "Discovery and Design of Novel Inhibitors of Botulinus Neurotoxin A: Targeted 'Hinge' Peptide Libraries," *Journal of Applied Toxicology* 23:1-7 (2003); Oost et al., "Design and Synthesis of Substrate-Based Inhibitors of *Botulinum* Neurotoxin Type B Metalloprotease," *Biopolymers (Peptide Science)* 71:602-619 (2003), which are hereby incorporated by reference in its entirety).

SNARE motif peptides have been shown to prevent cleavage of synaptic complex components in *Aplysia* neurons (Rossetto et al., "SNARE Motif and Neurotoxins," *Nature* 372:415-416 (1994), which is hereby incorporated by reference in its entirety). SNARE motif peptides are common to the substrate binding site of known BoNT serotypes, and have been shown to inhibit the toxic LC when injected into BoNT-affected neurons (Rossetto et al., "SNARE Motif and Neurotoxins," *Nature* 372:415-416 (1994), which is hereby incorporated by reference in its entirety).

In a preferred embodiment, the Clostridial neurotoxin propeptide light chain region has one or more non-native motifs (e.g., SNARE motif peptides), which replace surface alpha-helix domains of the native propeptide. Seven surface alpha-helix domains in the light chain region of *Clostridium botulinum* serotypes are identified in FIG. 11.

A variety of Clostridial neurotoxin propeptides with light chain regions containing non-native motifs (e.g., SNARE

motif peptides) in place of surface alpha-helix domains can be created. As described in greater detail below, these non-native motif bearing propeptides are generated by altering the nucleotide sequences of nucleic acids encoding the Clostridial neurotoxin propeptides.

Another aspect of the present invention relates to an isolated nucleic acid molecule encoding an isolated Clostridial neurotoxin propeptide of the present invention.

Nucleic acid molecules encoding full-length toxic Clostridial neurotoxins are well known in the art (See e.g., GenBank Accession Nos. M81186 (BoNT B); D90210 (BoNT C); S49407 (BoNT D); D90210 (BoNT E); X81714 (BoNT F); and X74162 (BoNT G)).

Nucleic acid molecules of the present invention preferably encode the amino acid sequences of FIG. 2. In particular, the nucleic acid molecules of the present invention are modified from the wild type BoNT serotype sequences to have one or more characteristics selected from the group consisting of a mutation which renders the encoded propeptide resistant to low-specificity proteolysis, one or more silent mutations that inactivate putative internal DNA regulatory elements, and one or more unique restriction sites. In particular, and as illustrated for each BoNT serotype in FIG. 2, mature neurotoxin stability and yield are optimized by site-directed mutation of residues within the intermediate region of the propeptide, thereby reducing the propeptides' susceptibility to non-specific proteolysis and poisoning of the host organism used for expression by the mature neurotoxin. Also, silent mutations are introduced into DNA regulatory elements that can affect RNA transcription or expression of the Clostridial neurotoxin propeptide in the system of choice. In addition, unique endonuclease restriction sites are introduced to enable creation of chimeric proteins.

A nucleic acid molecule of the present invention may also have a disabling mutation in a region encoding an active metalloprotease site of the propeptide, as described supra.

A nucleic acid molecule of the present invention may also have a mutation in a region encoding the light chain region, such that the nucleic acid molecule encodes, in the light chain region, a non-native motif capable of inactivating light chain metalloprotease activity in a toxic clostridial neurotoxin. Suitable non-native motifs are described supra.

A further aspect of the present invention relates to an expression system having a nucleic acid molecule encoding an isolated Clostridial neurotoxin propeptide of the present invention in a heterologous vector.

Yet another aspect of the present invention relates to a host cell having a heterologous nucleic acid molecule encoding an isolated Clostridial neurotoxin propeptide of the present invention.

Still another aspect of the present invention relates to a method of expressing a recombinant physiologically active Clostridial neurotoxin of the present invention. This method involves providing a nucleic acid construct having a nucleic acid molecule encoding an isolated Clostridial neurotoxin propeptide of the present invention. The nucleic acid construct has a heterologous promoter operably linked to the nucleic acid molecule and a 3' regulatory region operably linked to the nucleic acid molecule. The nucleic acid construct is then introduced into a host cell under conditions effective to express the physiologically active Clostridial neurotoxin.

In a preferred embodiment, the expressed neurotoxin is contacted with a highly specific protease under conditions effective to effect cleavage at the intermediate region. Preferably, the intermediate region of the Clostridial neurotoxin

propeptide is not cleaved by proteases endogenous to the expression system or the host cell.

Expression of a Clostridial neurotoxin of the present invention can be carried out by introducing a nucleic acid molecule encoding a Clostridial neurotoxin propeptide into an expression system of choice using conventional recombinant technology. Generally, this involves inserting the nucleic acid molecule into an expression system to which the molecule is heterologous (i.e., not normally present). The introduction of a particular foreign or native gene into a mammalian host is facilitated by first introducing the gene sequence into a suitable nucleic acid vector. "Vector" is used herein to mean any genetic element, such as a plasmid, phage, transposon, cosmid, chromosome, virus, virion, etc., which is capable of replication when associated with the proper control elements and which is capable of transferring gene sequences between cells. Thus, the term includes cloning and expression vectors, as well as viral vectors. The heterologous nucleic acid molecule is inserted into the expression system or vector in proper sense (5'→3') orientation and correct reading frame. The vector contains the necessary elements for the transcription and translation of the inserted Clostridial neurotoxin propeptide-coding sequences.

U.S. Pat. No. 4,237,224 to Cohen and Boyer, which is hereby incorporated by reference in its entirety, describes the production of expression systems in the form of recombinant plasmids using restriction enzyme cleavage and ligation with DNA ligase. These recombinant plasmids are then introduced by means of transformation and replicated in unicellular cultures including prokaryotic organisms and eukaryotic cells grown in tissue culture.

Recombinant genes may also be introduced into viruses, including vaccinia virus, adenovirus, and retroviruses, including lentivirus. Recombinant viruses can be generated by transfection of plasmids into cells infected with virus.

Suitable vectors include, but are not limited to, the following viral vectors such as lambda vector system gt11, gt WES.tB, Charon 4, and plasmid vectors such as pBR322, pBR325, pACYC177, pACYC184, pUC8, pUC9, pUC18, pUC19, pLG339, pR290, pKC37, pKC101, SV 40, pBlue-script II SK +/- or KS +/- (see "Stratagene Cloning Systems" Catalog (1993) from Stratagene, La Jolla, Calif., which is hereby incorporated by reference in its entirety), pQE, pIH821, pGEX, pFastBac series (Invitrogen), pET series (see F. W. Studier et. al., "Use of T7 RNA Polymerase to Direct Expression of Cloned Genes," *Gene Expression Technology* Vol. 185 (1990), which is hereby incorporated by reference in its entirety), and any derivatives thereof. Recombinant molecules can be introduced into cells via transformation, particularly transduction, conjugation, mobilization, or electroporation. The DNA sequences are cloned into the vector using standard cloning procedures in the art, as described by Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Springs Laboratory, Cold Springs Harbor, N.Y. (1989), which is hereby incorporated by reference in its entirety.

A variety of host-vector systems may be utilized to express the Clostridial neurotoxin propeptide-encoding sequence in a cell. Primarily, the vector system must be compatible with the host cell used. Host-vector systems include but are not limited to the following: bacteria transformed with bacteriophage DNA, plasmid DNA, or cosmid DNA; microorganisms such as yeast containing yeast vectors; mammalian cell systems infected with virus (e.g., vaccinia virus, adenovirus, etc.); insect cell systems infected with virus (e.g., baculovirus); and plant cells infected by bacteria. The expression elements of these vectors vary in their strength and specificities. Depend-

ing upon the host-vector system utilized, any one of a number of suitable transcription and translation elements can be used.

Different genetic signals and processing events control many levels of gene expression (e.g., DNA transcription and messenger RNA ("mRNA") translation).

Transcription of DNA is dependent upon the presence of a promoter which is a DNA sequence that directs the binding of RNA polymerase and thereby promotes mRNA synthesis. The DNA sequences of eukaryotic promoters differ from those of prokaryotic promoters. Furthermore, eukaryotic promoters and accompanying genetic signals may not be recognized in or may not function in a prokaryotic system, and, further, prokaryotic promoters are not recognized and do not function in eukaryotic cells.

Similarly, translation of mRNA in prokaryotes depends upon the presence of the proper prokaryotic signals which differ from those of eukaryotes. Efficient translation of mRNA in prokaryotes requires a ribosome binding site called the Shine-Dalgarno ("SD") sequence on the mRNA. This sequence is a short nucleotide sequence of mRNA that is located before the start codon, usually AUG, which encodes the amino-terminal methionine of the protein. The SD sequences are complementary to the 3'-end of the 16S rRNA (ribosomal RNA) and probably promote binding of mRNA to ribosomes by duplexing with the rRNA to allow correct positioning of the ribosome. For a review on maximizing gene expression see Roberts and Lauer, *Methods in Enzymology*, 68:473 (1979), which is hereby incorporated by reference in its entirety.

Promoters vary in their "strength" (i.e., their ability to promote transcription). For the purposes of expressing a cloned gene, it is desirable to use strong promoters in order to obtain a high level of transcription and, hence, expression of the gene. Depending upon the host cell system utilized, any one of a number of suitable promoters may be used. For instance, when cloning in *E. coli*, its bacteriophages, or plasmids, promoters such as the PH promoter, T7 phage promoter, lac promoter, trp promoter, recA promoter, ribosomal RNA promoter, the P<sub>R</sub> and P<sub>L</sub> promoters of coliphage lambda and others, including but not limited, to lacUV5, ompF, bla, lpp, and the like, may be used to direct high levels of transcription of adjacent DNA segments. Additionally, a hybrid trp-lacUV5 (tac) promoter or other *E. coli* promoters produced by recombinant DNA or other synthetic DNA techniques may be used to provide for transcription of the inserted gene.

Bacterial host cell strains and expression vectors may be chosen which inhibit the action of the promoter unless specifically induced. In certain operons, the addition of specific inducers is necessary for efficient transcription of the inserted DNA. For example, the lac operon is induced by the addition of lactose or IPTG (isopropylthio-beta-D-galactoside). A variety of other operons, such as trp, pro, etc., are under different controls.

Specific initiation signals are also required for efficient gene transcription and translation in prokaryotic cells. These transcription and translation initiation signals may vary in "strength" as measured by the quantity of gene specific messenger RNA and protein synthesized, respectively. The DNA expression vector, which contains a promoter, may also contain any combination of various "strong" transcription and/or translation initiation signals. For instance, efficient translation in *E. coli* requires a Shine-Dalgarno ("SD") sequence about 7-9 bases 5' to the initiation codon (ATG) to provide a ribosome binding site. Thus, any SD-ATG combination that can be utilized by host cell ribosomes may be employed. Such combinations include but are not limited to the SD-ATG combination from the cro gene or the N gene of coliphage

lambda, or from the *E. coli* tryptophan E, D, C, B or A genes. Additionally, any SD-ATG combination produced by recombinant DNA or other techniques involving incorporation of synthetic nucleotides may be used.

Depending on the vector system and host utilized, any number of suitable transcription and/or translation elements, including constitutive, inducible, and repressible promoters, as well as minimal 5' promoter elements may be used.

The Clostridial neurotoxin-encoding nucleic acid, a promoter molecule of choice, a suitable 3' regulatory region, and if desired, a reporter gene, are incorporated into a vector-expression system of choice to prepare a nucleic acid construct using standard cloning procedures known in the art, such as described by Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Third Edition, Cold Spring Harbor: Cold Spring Harbor Laboratory Press, New York (2001), which is hereby incorporated by reference in its entirety.

The nucleic acid molecule encoding a Clostridial neurotoxin is inserted into a vector in the sense (i.e., 5'→3') direction, such that the open reading frame is properly oriented for the expression of the encoded Clostridial neurotoxin propeptide under the control of a promoter of choice. Single or multiple nucleic acids may be ligated into an appropriate vector in this way, under the control of a suitable promoter, to prepare a nucleic acid construct.

Once the isolated nucleic acid molecule encoding the Clostridial neurotoxin propeptide has been inserted into an expression vector, it is ready to be incorporated into a host cell. Recombinant molecules can be introduced into cells via transformation, particularly transduction, conjugation, lipofection, protoplast fusion, mobilization, particle bombardment, or electroporation. The DNA sequences are incorporated into the host cell using standard cloning procedures known in the art, as described by Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Springs Laboratory, Cold Springs Harbor, N.Y. (1989), which is hereby incorporated by reference in its entirety. Suitable hosts include, but are not limited to, bacteria, virus, yeast, fungi, mammalian cells, insect cells, plant cells, and the like. Preferable host cells of the present invention include, but are not limited to, *Escherichia coli*, insect cells, and *Pichia pastoris* cells.

Typically, an antibiotic or other compound useful for selective growth of the transformed cells only is added as a supplement to the media. The compound to be used will be dictated by the selectable marker element present in the plasmid with which the host cell was transformed. Suitable genes are those which confer resistance to gentamycin, G418, hygromycin, puromycin, streptomycin, spectinomycin, tetracycline, chloramphenicol, and the like. Similarly, "reporter genes" which encode enzymes providing for production of an identifiable compound, or other markers which indicate relevant information regarding the outcome of gene delivery, are suitable. For example, various luminescent or phosphorescent reporter genes are also appropriate, such that the presence of the heterologous gene may be ascertained visually.

In a preferred embodiment of the present invention, the expressed neurotoxin propeptide is contacted with a highly specific protease (e.g., enterokinase) under conditions effective to enable cleavage at the intermediate region of the propeptide of the present invention. Preferably, the expressed neurotoxin propeptide has one or more disulfide bridges.

Another aspect of the present invention relates to an isolated, physiologically active Clostridial neurotoxin produced by cleaving an isolated Clostridial neurotoxin propeptide of the present invention. The propeptide is cleaved at the highly

specific protease cleavage site. The light and heavy chain regions are linked by a disulfide bond.

As discussed supra, Clostridial neurotoxins are synthesized as single chain propeptides which are later activated by a specific proteolysis cleavage event, generating a dimer joined by a disulfide bond. These structural features can be illustrated using BoNT A as an example, and are generally applicable to all *Clostridium botulinum* serotypes. The mature BoNT A is composed of three functional domains of Mr~50,000 (FIG. 3A), where the catalytic function responsible for toxicity is confined to the light chain (residues 1-437), the translocation activity is associated with the N-terminal half of the heavy chain (residues 448-872), and cell binding is associated with its C-terminal half (residues 873-1,295) (Johnson, "Clostridial Toxins as Therapeutic Agents: Benefits of Nature's Most Toxic Proteins," *Annu. Rev. Microbiol.* 53:551-575 (1999); Montecucco et al., "Structure and Function of Tetanus and *Botulinum* Neurotoxins," *Q. Rev. Biophys.* 28:423-472 (1995), which are hereby incorporated by reference in their entirety).

Optimized expression and recovery of recombinant neurotoxins for BoNT serotypes in a native and physiologically active state is achieved by the introduction of one or more alterations to the nucleotide sequences encoding the BoNT propeptides, as discussed supra. These mutations are designed to maximize yield of recombinant Clostridial neurotoxin, while retaining the native toxins structure and biological activity.

Isolated, full-length Clostridial neurotoxins of the present invention are physiologically active. This physiological activity includes, but is not limited to, toxin immunogenicity, trans- and intra-cellular trafficking, and cell recognition.

The mechanism of cellular binding and internalization of Clostridial toxins is still poorly understood. No specific receptor has been unambiguously identified, and the binding constants have not been characterized. The C-terminal portion of the heavy chain of all Clostridial neurotoxins binds to gangliosides (sialic acid-containing glycolipids), with a preference for gangliosides of the G<sub>1b</sub> series (Montecucco et al., "Structure and Function of Tetanus and *Botulinum* Neurotoxins," *Q. Rev. Biophys.* 28:423-472 (1995); Montecucco, "How Do Tetanus and *Botulinum* Toxins Bind to Neuronal Membranes?" *TIBS* 11:314-317 (1986); and Van Heyningen et al., "The Fixation of Tetanus Toxin by Ganglioside," *J. Gen. Microbiol.* 24:107-119 (1961), which are hereby incorporated by reference in their entirety). The sequence responsible for ganglioside binding has been identified for the structurally similar TeNT molecule, and is located within the 34 C-terminal amino acid residues of its heavy chain. BoNT A, B, C, E, and F share a high degree of homology with TeNT in this region (FIG. 1) (Shapiro et al., "Identification of a Ganglioside Recognition Domain of Tetanus Toxin Using a Novel Ganglioside Photoaffinity Ligand," *J. Biol. Chem.* 272:30380-30386 (1997), which is hereby incorporated by reference in its entirety). Multiple types of evidence suggest the existence of at least one additional component involved in the binding of Clostridial neurotoxins to neuronal membranes (Montecucco et al., "Structure and Function of Tetanus and *Botulinum* Neurotoxins," *Q. Rev. Biophys.* 28:423-472 (1995); Montecucco, "How Do Tetanus and *Botulinum* Toxins Bind to Neuronal Membranes?" *TIBS* 11:314-317 (1986), which are hereby incorporated by reference in their entirety). In two reports (Nishiki et al., "The High-Affinity Binding of *Clostridium Botulinum* Type B Neurotoxin to Synaptotagmin II Associated with Gangliosides G<sub>T1b</sub>/G<sub>D1a</sub>," *FEBS Lett.* 378:253-257 (1996); Dong et al., "Synaptotagmins I and II Mediate Entry of *Botulinum* Neurotoxin B into Cells," *J. Cell Biol.*

162:1293-1303 (2003), which are hereby incorporated by reference in their entirety), synaptotagmins were identified as possible candidates for the auxiliary BoNT B receptor, and synaptotagmins I and II were implicated as neuronal receptors for BoNT G (Rummel et al., "Synaptotagmins I and II Act as Nerve Cell Receptors for *Botulinum* Neurotoxin G," *J. Biol. Chem.* 279:30865-30870 (2004), which is hereby incorporated by reference in its entirety). However despite the structural similarity in the putative receptor-binding domain of Clostridial neurotoxins, other toxin subtypes show no affinity for synaptotagmins or synaptotagmin-related molecules. Lipid rafts (Herreros et al., "Lipid Rafts Act as Specialized Domains for Tetanus Toxin Binding and Internalization into Neurons," *Mol. Biol. Cell* 12:2947-2960 (2001), which is hereby incorporated by reference in its entirety) have been implicated as a specialized domain involved in TeNT binding and internalization into neurons, but these domains are widely distributed on multiple cell types, and therefore cannot simply explain the high specificity of the toxins for neurons.

Clostridial neurotoxins are internalized through the presynaptic membrane by an energy-dependent mechanism (Montecucco et al., "Structure and Function of Tetanus and *Botulinum* Neurotoxins," *Q. Rev. Biophys.* 28:423-472 (1995); Matteoli et al., "Synaptic Vesicle Endocytosis Mediates the Entry of Tetanus Neurotoxin into Hippocampal Neurons," *Proc. Natl. Acad. Sci. USA* 93:13310-13315 (1996); and Mukherjee et al., "Endocytosis," *Physiol. Rev.* 77:759-803 (1997), which are hereby incorporated by reference in their entirety), and rapidly appear in vesicles where they are at least partially protected from degradation (Dolly et al., "Acceptors for *Botulinum* Neurotoxin Reside on Motor Nerve Terminals and Mediate Its Internalization," *Nature* 307:457-460 (1984); Critchley et al., "Fate of Tetanus Toxin Bound to the Surface of Primary Neurons in Culture: Evidence for Rapid Internalization," *J. Cell Biol.* 100:1499-1507 (1985), which are hereby incorporated by reference in their entirety). The BoNT complex of light and heavy chains interacts with the endocytic vesicle membrane in a chaperone-like way, preventing aggregation and facilitating translocation of the light chain in a fashion similar to the protein conducting/translocating channels of smooth ER, mitochondria, and chloroplasts (Koriazova et al., "Translocation of *Botulinum* Neurotoxin Light Chain Protease through the Heavy Chain Channel," *Nat. Struct. Biol.* 10:13-18 (2003), which is hereby incorporated by reference in its entirety). Acidification of the endosome is believed to induce pore formation, which allows translocation of the light chain to the cytosol upon reduction of the interchain disulfide bond (Hoch et al., "Channels Formed by *Botulinum*, Tetanus, and Diphtheria Toxins in Planar Lipid Bilayers: Relevance to Translocation of Proteins Across Membranes," *Proc. Natl. Acad. Sci. USA* 82:1692-1696 (1985), which is hereby incorporated by reference in its entirety). Within the cytosol, the light chain displays a zinc-endopeptidase activity specific for protein components of the synaptic vesicle exocytosis apparatus. TeNT and BoNT B, D, F, and G recognize VAMP/synaptobrevin. This integral protein of the synaptic vesicle membrane is cleaved at a single peptide bond, which differs for each neurotoxin. BoNT A, C, and E recognize and cleave SNAP-25, a protein of the presynaptic membrane, at two different sites within the carboxyl terminus. BoNT C also cleaves syntaxin, another protein of the nerve plasmalemma (Montecucco et al., "Structure and Function of Tetanus and *Botulinum* Neurotoxins," *Q. Rev. Biophys.* 28:423-472 (1995); Sutton et al., "Crystal Structure of a SNARE Complex Involved in Synaptic Exocytosis at 2.4 Å Resolution," *Nature* 395:347-353 (1998), which are hereby

incorporated by reference in their entirety). The cleavage of any component of the synaptic release machinery results in inhibition of acetylcholine release, ultimately leading to neuromuscular paralysis.

In one embodiment of the present invention, the isolated Clostridial neurotoxin is toxic. The toxicity of Clostridial neurotoxins is a result of a multi-step mechanism. From the circulation, BoNT targets the pre-synaptic membrane of neuromuscular junctions, where it is internalized to directly exert its toxic effect on the peripheral nervous system (Dolly et al., "Acceptors for *Botulinum* Neurotoxin Reside on Motor Nerve Terminals and Mediate Its Internalization," *Nature* 307:457-460 (1984), which is hereby incorporated by reference in its entirety). Toxicity at the neuromuscular junction involves neuron binding; internalization into endocytic vesicles, similar to those involved in synaptic vesicle recycling; activation within an acidic compartment to the proteolytically active toxin which then penetrates into the neuronal cytoplasm; and target recognition and catalytic cleavage of substrates in the neuronal machinery for synaptic vesicle exocytosis.

In an alternative embodiment of the present invention, the isolated Clostridial neurotoxin is physiologically active and atoxic. The endopeptidase activity responsible for Clostridial neurotoxin toxicity is believed to be associated with the presence of a HEXxHxxH (SEQ ID NO: 25) motif in the light chain, characteristic of metalloproteases (FIG. 1). Mutagenesis of BoNT A light chain, followed by microinjection of the corresponding mRNA into presynaptic cholinergic neurons of *Aplysia californica*, allowed the minimal essential domain responsible for toxicity to be identified (Kurazono et al., "Minimal Essential Domains Specifying Toxicity of the Light Chains of Tetanus Toxin and *Botulinum* Neurotoxin Type A," *J. Biol. Chem.* 267:14721-14729 (1992), which is hereby incorporated by reference in its entirety). Site-directed mutagenesis of BoNT A light chain pinpointed the amino acid residues involved in Zn<sup>2</sup> coordination, and formation of the active metalloendoprotease core which cleaves SNAP-25 (Rigoni et al., "Site-Directed Mutagenesis Identifies Active-Site Residues of the Light Chain of *Botulinum* Neurotoxin Type A," *Biochem. Biophys. Res. Commun.* 288:1231-1237 (2001), which is hereby incorporated by reference in its entirety). The three-dimensional structures of Clostridial neurotoxins and their derivatives confirmed the mutagenesis results, and detailed the spatial organization of the protein domains. For the BoNT A holotoxin, crystal structure was obtained to a resolution of 3.3 Å (Lacy et al., "Crystal Structure of *Botulinum* Neurotoxin Type A and Implications for Toxicity," *Nat. Struct. Biol.* 5:898-902 (1998), which is hereby incorporated by reference in its entirety). The BoNT B holotoxin crystal structure was determined at 1.8 and 2.6 Å resolution (Swaminathan et al., "Structural Analysis of the Catalytic and Binding Sites of *Clostridium Botulinum* Neurotoxin B," *Nat. Struct. Biol.* 7:693-699 (2000), which is hereby incorporated by reference in its entirety). Recently, a crystal structure for BoNT E catalytic domain was determined to 2.1 Å resolution (Agarwal et al., "Structural Analysis of *Botulinum* Neurotoxin Type E Catalytic Domain and Its Mutant Glu212>Gln Reveals the Pivotal Role of the Glu212 Carboxylate in the Catalytic Pathway," *Biochemistry* 43:6637-6644 (2004), which is hereby incorporated by reference in its entirety). The later study provided multiple interesting structural details, and helps explain the complete loss of metalloendoproteolytic activity in the BoNT E LC E212>Q mutant. The availability of this detailed information on the relationship between the amino acid sequence and

biological activities of Clostridial toxins enables the design of modified toxin genes with properties specifically altered for therapeutic goals.

Thus, in a preferred embodiment, the physiologically active and atoxic Clostridial neurotoxin of the present invention has a disabling mutation in an active metalloprotease site.

The physiologically active and atoxic Clostridial neurotoxin of the present invention may also have a non-native motif (e.g., a SNARE motif) in the light chain region that is capable of inactivating light chain metalloprotease activity in a toxic Clostridial neurotoxin. FIG. 11 illustrates the sequences of nine chimeric proteins, which are physiologically active and atoxic Clostridial neurotoxins containing at least one non-native motif in the light chain region that is capable of inactivating light chain metalloprotease activity in a toxic Clostridial neurotoxin. The non-native motifs are substituted for alpha-helix domains. When present in the physiologically active and atoxic Clostridial neurotoxin, the non-native protein motifs enable the neurotoxin to bind, inactivate, or otherwise mark the toxic light chain region of a wild type Clostridial neurotoxin for elimination from the cytosol of neurotoxin-affected neurons. As such, a physiologically active and atoxic Clostridial neurotoxin having a non-native motif in the light chain region that is capable of inactivating light chain metalloprotease activity in a toxic Clostridial neurotoxin is useful as an antidote to effectively target the cytoplasm of neurotoxin-affected neurons. Administration of such antidotes is described in greater detail below.

Yet a further aspect of the present invention relates to a vaccine or antidote having an isolated, physiologically active, atoxic, Clostridial neurotoxin produced by cleaving an isolated Clostridial neurotoxin propeptide of the present invention. The propeptide is cleaved at the highly specific protease cleavage site. The light and heavy chain regions are linked by a disulfide bond.

Developing effective vaccines and antidotes against Clostridial neurotoxins requires the preservation of structural features important to toxin trafficking and immunogenicity. From a practical perspective, this is most easily achieved by first producing recombinant molecules that retain the structural features and toxicity of native toxin, followed by selective modification to eliminate toxicity and introduce therapeutic utility. To achieve this goal, a versatile platform for the genetic manipulation of Clostridial toxin genes and for their selective modification was developed (described infra). The genetic engineering scheme can produce full-length toxic and atoxic derivatives of BoNT A, which retains important aspects of the wild toxin's native structure. This methodology can be generalized across the entire family of Clostridial neurotoxins because of their structural similarities (See FIGS. 1-2).

Thus, in a preferred embodiment, the vaccine or antidote of the present invention is a physiologically active and atoxic Clostridial neurotoxin from *Clostridium botulinum*, such as from *Clostridium botulinum* serotypes A-G. As described supra, the vaccine or antidote has the physiological activity of a wild Clostridial neurotoxin, which activity includes, but is not limited to, toxin immunogenicity, trans- and intra-cellular trafficking, and cell recognition. The Clostridial neurotoxin of the vaccine or antidote is rendered atoxic by a mutation in its active metalloprotease site, as described supra. Additional mutations may be introduced to ensure atoxicity and introduce new biological activities, while preserving systemic trafficking and cellular targeting of the vaccine or antidote. As has also been described, the vaccine or antidote may possess non-native motifs in the light chain region that are

capable of inactivating light chain metalloprotease activity in a toxic Clostridial neurotoxin.

Atoxic Clostridial neurotoxins can be tested as candidate vaccines and antidotes to BoNT poisoning. Atoxic derivatives are created using the BoNT toxic derivative constructs developed under the methods described infra. Point mutations are introduced into the toxin's active metalloprotease site to eliminate toxicity while maintaining native toxin structure, immunogenicity, trans- and intra-cellular trafficking, and cell recognition. Expression systems and purification schemes are optimized as described infra. Derivatives found to completely lack toxicity yet retain relevant biological activities of the native toxin, are evaluated for their potential as either vaccines or antidotes to BoNT poisoning. Parenteral routes of administration are tested first, followed by evaluation of oral and inhalational routes as applicable. Utility as a vaccine is determined by immunogenicity and challenge studies in mice. Utility as an antidote is first evaluated in vitro by testing the ability of atoxic derivatives to prevent neuromuscular blockade in the mouse phrenic-nerve hemidiaphragm, and to inhibit native toxin trafficking in the transcytosis assay. Effective in vitro antagonists are tested as in vivo antidotes, and may be superior to antibody-based antidotes because they more effectively mimic native toxin absorption and trafficking pathways. Antidote effectiveness in vivo is first evaluated using simultaneous dosing. Additional dosage and timing parameters relevant to using antidotes under crisis situations is further evaluated for atoxic derivatives found to be effective when administered simultaneously with toxin. Using these procedures, a series of atoxic derivatives and fusion proteins are created and their biological activities systematically catalogued. The availability of these well characterized BoNT gene constructs and toxin derivatives enables the rational design of new anti-BoNT therapeutics. Dose-response analyses and challenge studies against active neurotoxin provide data that allows the best candidate vaccines and antidotes to be selected for further development.

A further aspect of the present invention relates to method of immunizing a subject against toxic effects of a Clostridial neurotoxin. This method involves administering a vaccine of the present invention to the subject under conditions effective to immunize the subject against toxic effects of Clostridial neurotoxin.

The subject administered the vaccine may further be administered a booster of the vaccine under conditions effective to enhance immunization of the subject.

Another aspect of the present invention relates to a method of treating a subject for toxic effects of a Clostridial neurotoxin. This method involves administering an antidote comprising an isolated, physiologically active, atoxic, Clostridial neurotoxin produced by cleaving the isolated Clostridial neurotoxin propeptide of the present invention to the subject under conditions effective to treat the subject for toxic effects of Clostridial neurotoxin.

A vaccine or antidote of the present invention can be administered to a subject orally, parenterally, for example, subcutaneously, intravenously, intramuscularly, intraperitoneally, by intranasal instillation, or by application to mucous membranes, such as, that of the nose, throat, and bronchial tubes. The vaccine or antidote may be administered alone or with suitable pharmaceutical carriers, and can be in solid or liquid form such as, tablets, capsules, powders, solutions, suspensions, or emulsions.

The vaccine or antidote of the present invention may be orally administered, for example, with an inert diluent, or with an assimilable edible carrier, or may be enclosed in hard or soft shell capsules, or may be compressed into tablets, or



may be incorporated directly with the food of the diet. For oral therapeutic administration, the vaccine or antidote may be incorporated with excipients and used in the form of tablets, capsules, elixirs, suspensions, syrups, and the like. Such compositions and preparations should contain at least 0.1% of active compound. The percentage of the compound in these compositions may, of course, be varied and may conveniently be between about 2% to about 60% of the weight of the unit. The amount of active compound in such therapeutically useful compositions is such that a suitable dosage will be obtained. Preferred compositions according to the present invention are prepared so that an oral dosage unit contains between about 1 and 250 mg of active compound.

The tablets, capsules, and the like may also contain a binder such as gum tragacanth, acacia, corn starch, or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid; a lubricant such as magnesium stearate; and a sweetening agent such as sucrose, lactose, or saccharin. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier, such as a fatty oil.

Various other materials may be present as coatings or to modify the physical form of the dosage unit. For instance, tablets may be coated with shellac, sugar, or both. A syrup may contain, in addition to active ingredient, sucrose as a sweetening agent, methyl and propylparabens as preservatives, a dye, and flavoring such as cherry or orange flavor.

The vaccine or antidote may also be administered parenterally. Solutions or suspensions can be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions can also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof in oils. Illustrative oils are those of petroleum, animal, vegetable, or synthetic origin, for example, peanut oil, soybean oil, or mineral oil. In general, water, saline, aqueous dextrose and related sugar solution, and glycols such as, propylene glycol or polyethylene glycol, are preferred liquid carriers, particularly for injectable solutions. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases, the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (e.g., glycerol, propylene glycol, and liquid polyethylene glycol), suitable mixtures thereof, and vegetable oils.

The vaccine or antidote of the present invention may also be administered directly to the airways in the form of an aerosol. For use as aerosols, the vaccine or antidote of the present invention in solution or suspension may be packaged in a pressurized aerosol container together with suitable propellants, for example, hydrocarbon propellants like propane, butane, or isobutane with conventional adjuvants. The vaccine or antidote of the present invention also may be administered in a non-pressurized form such as in a nebulizer or atomizer.

A further aspect of the present invention relates to a chimeric protein having a first protein or protein fragment having a heavy chain region of a Clostridial neurotoxin and a second protein or protein fragment linked to the first protein or protein fragment.

In a preferred embodiment, the second protein or protein fragment has therapeutic functionality which can target specific steps in a trafficking pathway of the Clostridial neurotoxin.

BoNTs pass across epithelial surfaces without being destroyed or causing local toxicity. Passage across epithelia is believed to occur by specific binding and transcytosis. The ability of intact BoNT A to pass through pulmonary epithelia and resist proteolytic inactivation was demonstrated in rat primary alveolar epithelial cells and in immortalized human pulmonary adenocarcinoma (Calu-3) cells. The rate of transport was greater in the apical-to-basolateral direction than in the basolateral-to-apical direction, and it was blocked by serotype-specific toxin antibodies (Park et al., "Inhalational Poisoning by *Botulinum* Toxin and Inhalation Vaccination with Its Heavy-Chain Component," *Infect. Immun.* 71:1147-1154 (2003), which is hereby incorporated by reference in its entirety).

The ability of Clostridial neurotoxins to pass undegraded through epithelial barriers via transcytosis and to specifically target nervous tissue makes Clostridial neurotoxins useful in the development of oral and inhalational carriers for therapeutic agents that cannot normally be delivered via these routes of administration, and as delivery vehicles which can specifically target the peripheral and central nervous system.

Still another aspect of the present invention relates to a treatment method. This method involves contacting a patient with an isolated, physiologically active, toxic, Clostridial neurotoxin produced by cleaving an isolated Clostridial neurotoxin propeptide according to the present invention, under conditions effective to treat the patient.

By treatment, it is meant aesthetic treatment (See e.g., Carruthers et al., "*Botulinum* Toxin A in the Mid and Lower Face and Neck," *Dermatol. Clin.* 22:151-158 (2004); Lang, "History and Uses of BOTOX (*Botulinum* Toxin Type A)," *Lippincotts Case Manag.* 9:109-112 (2004); Naumann et al., "Safety of *Botulinum* Toxin Type A: A Systematic Review and Meta-Analysis," *Curr. Med. Res. Opin.* 20:981-990 (2004); Vartanian et al., "Facial Rejuvenation Using *Botulinum* Toxin A: Review and Updates," *Facial Plast. Surg.* 20:11-19 (2004), which are hereby incorporated by reference in their entirety) as well as therapeutic treatment (See e.g., Bentsianov et al., "Noncosmetic Uses of *Botulinum* Toxin," *Clin. Dermatol.* 22:82-88 (2004); Carruthers et al., "Botox: Beyond Wrinkles," *Clin. Dermatol.* 22:89-93 (2004); Jankovic, "*Botulinum* Toxin In Clinical Practice," *J. Neurol. Neurosurg. Psychiatry* 75:951-957 (2004); Klein, "The Therapeutic Potential of *Botulinum* Toxin," *Dermatol. Surg.* 30:452-455 (2004); Schurch, "The Role of *Botulinum* Toxin in Neurology," *Drugs Today (Barc)* 40:205-212 (2004), which are hereby incorporated by reference in their entirety).

Preferred treatment methods of the present invention include, but are not limited to, dermatologic, gastroenterologic, genitourinaric, and neurologic treatment.

Dermatologic treatment includes, but is not limited to, treatment for Rhytides (wrinkles) (Sadick et al., "Comparison of *Botulinum* Toxins A and B in the Treatment of Facial Rhytides," *Dermatol. Clin.* 22:221-226 (2004), which is hereby incorporated by reference in its entirety), including glabellar (Carruthers et al., "*Botulinum* Toxin type A for the Treatment of Glabellar Rhytides," *Dermatol. Clin.* 22:137-144 (2004); Ozsoy et al., "Two-Plane Injection of *Botulinum* Exotoxin A in Glabellar Frown Lines," *Aesthetic Plast. Surg.* 28:114-115 (2004); which are hereby incorporated by reference in their entirety), neck lines (Brandt et al., "*Botulinum* Toxin for the Treatment of Neck Lines and Neck Bands," *Dermatol. Clin.* 22:159-166 (2004), which is hereby incor-

porated by reference in its entirety), crows feet (Levy et al., "Botulinum Toxin A: A 9-Month Clinical and 3D In Vivo Profilometric Crow's Feet Wrinkle Formation Study," *J. Cosmet. Laser Ther.* 6:16-20 (2004), which is hereby incorporated by reference in its entirety), and brow contour (Chen et al., "Altering Brow Contour with Botulinum Toxin," *Facial Plast. Surg. Clin. North Am.* 11:457-464 (2003), which is hereby incorporated by reference in its entirety). Other dermatologic treatment includes treatment for hypertrophic masseter muscles in Asians (Ahn et al., "Botulinum Toxin for Masseter Reduction in Asian Patients," *Arch. Facial Plast. Surg.* 6:188-191 (2004), which is hereby incorporated by reference in its entirety) and focal hyperhidrosis (Glogau, "Treatment of Hyperhidrosis with Botulinum Toxin," *Dermatol. Clin.* 22:177-185, vii (2004), which is hereby incorporated by reference in its entirety), including axillary ("Botulinum Toxin (Botox) for Axillary Hyperhidrosis," *Med. Lett. Drugs Ther.* 46:76 (2004), which is hereby incorporated by reference in its entirety) and genital (Lee et al., "A Case of Foul Genital Odor Treated with Botulinum Toxin A," *Dermatol. Surg.* 30:1233-1235 (2004), which is hereby incorporated by reference in its entirety).

Gastroenterologic treatment includes, but is not limited to, treatment for esophageal motility disorders (Achem, "Treatment of Spastic Esophageal Motility Disorders," *Gastroenterol. Clin. North Am.* 33:107-124 (2004), which is hereby incorporated by reference in its entirety), pharyngeal-esophageal spasm (Bayles et al., "Operative Prevention and Management of Voice-Limiting Pharyngoesophageal Spasm," *Otolaryngol. Clin. North Am.* 37:547-558 (2004); Chao et al., "Management of Pharyngoesophageal Spasm with Botox," *Otolaryngol. Clin. North Am.* 37:559-566 (2004), which are hereby incorporated by reference in their entirety), and anal fissure (Brisinda et al., "Botulinum Neurotoxin to Treat Chronic Anal Fissure: Results of a Randomized 'Botox vs. Dysport' Controlled Trial," *Ailment Pharmacol. Ther.* 19:695-701 (2004); Jost et al., "Botulinum Toxin A in Anal Fissure: Why Does it Work?" *Dis. Colon Rectum* 47:257-258 (2004), which are hereby incorporated by reference in their entirety).

Genitourinaric treatment includes, but is not limited to, treatment for neurogenic dysfunction of the urinary tract ("Botulinic Toxin in Patients with Neurogenic Dysfunction of the Lower Urinary Tracts," *Urologia* July-August:44-48 (2004); Giannantoni et al., "Intravesical Resiniferatoxin Versus Botulinum-A Toxin Injections for Neurogenic Detrusor Overactivity: A Prospective Randomized Study," *J. Urol.* 172:240-243 (2004); Reitz et al., "Intravesical Therapy Options for Neurogenic Detrusor Overactivity," *Spinal Cord* 42:267-272 (2004), which are hereby incorporated by reference in their entirety), overactive bladder (Cruz, "Mechanisms Involved in New Therapies for Overactive Bladder," *Urology* 63:65-73 (2004), which is hereby incorporated by reference in its entirety), and neuromodulation of urinary urge incontinence (Abrams, "The Role of Neuromodulation in the Management of Urinary Urge Incontinence," *BJU Int.* 93:1116 (2004), which is hereby incorporated by reference in its entirety).

Neurologic treatment includes, but is not limited to, treatment for tourettes syndrome (Porta et al., "Treatment of Phonic Tics in Patients with Tourette's Syndrome Using Botulinum Toxin Type A," *Neurol. Sci.* 24:420-423 (2004), which is hereby incorporated by reference in its entirety) and focal muscle spasticity or dystonias (MacKinnon et al., "Corticospinal Excitability Accompanying Ballistic Wrist Movements in Primary Dystonia," *Mov. Disord.* 19:273-284 (2004), which is hereby incorporated by reference in its

entirety), including, but not limited to, treatment for cervical dystonia (Haussermann et al., "Long-Term Follow-Up of Cervical Dystonia Patients Treated with Botulinum Toxin A," *Mov. Disord.* 19:303-308 (2004), which is hereby incorporated by reference in its entirety), primary blepharospasm (Defazio et al., "Primary Blepharospasm: Diagnosis and Management," *Drugs* 64:237-244 (2004), which is hereby incorporated by reference in its entirety), hemifacial spasm, post-stroke (Bakheit, "Optimising the Methods of Evaluation of the Effectiveness of Botulinum Toxin Treatment of Post-Stroke Muscle Spasticity," *J. Neurol. Neurosurg. Psychiatry* 75:665-666 (2004), which is hereby incorporated by reference in its entirety), spasmodic dysphonia (Bender et al., "Speech Intelligibility in Severe Adductor Spasmodic Dysphonia," *J. Speech Lang. Hear. Res.* 47:21-32 (2004), which is hereby incorporated by reference in its entirety), facial nerve disorders (Finn, "Botulinum Toxin Type A: Fine-Tuning Treatment of Facial Nerve Injury," *J. Drugs Dermatol.* 3:133-137 (2004), which is hereby incorporated by reference in its entirety), and Rasmussen syndrome (Lozsadi et al., "Botulinum Toxin A Improves Involuntary Limb Movements in Rasmussen Syndrome," *Neurology* 62:1233-1234 (2004), which is hereby incorporated by reference in its entirety). Other neurologic treatments include treatment for amputation pain (Kern et al., "Effects of Botulinum Toxin Type B on Stump Pain and Involuntary Movements of the Stump," *Am. J. Phys. Med. Rehabil.* 83:396-399 (2004), which is hereby incorporated by reference in its entirety), voice tremor (Adler et al., "Botulinum Toxin Type A for Treating Voice Tremor," *Arch. Neurol.* 61:1416-1420 (2004), which is hereby incorporated by reference in its entirety), crocodile tear syndrome (Kyrmizakis et al., "The Use of Botulinum Toxin Type A in the Treatment of Frey and Crocodile Tears Syndrome," *J. Oral Maxillofac. Surg.* 62:840-844 (2004), which is hereby incorporated by reference in its entirety), marginal mandibular nerve paralysis, and pain control (Cui et al., "Subcutaneous Administration of Botulinum Toxin A Reduces Formalin-Induced Pain," *Pain* 107:125-133 (2004), which is hereby incorporated by reference in its entirety), including but not limited to pain after mastectomy (Layeeque et al., "Botulinum Toxin Infiltration for Pain Control After Mastectomy and Expander Reconstruction," *Ann. Surg.* 240:608-613 (2004), which is hereby incorporated by reference in its entirety) and chest pain of esophageal origin (Schumulson et al., "Current and Future Treatment of Chest Pain of Presumed Esophageal Origin," *Gastroenterol. Clin. North Am.* 33:93-105 (2004), which is hereby incorporated by reference in its entirety). Another neurologic treatment amenable to the methods of the present invention is headache (Blumenfeld et al., "Botulinum Neurotoxin for the Treatment of Migraine and Other Primary Headache Disorders," *Dermatol. Clin.* 22:167-175 (2004), which is hereby incorporated by reference in its entirety).

The methods of the present invention are also suitable for treatment of cerebral palsy (Balkrishnan et al., "Longitudinal Examination of Health Outcomes Associated with Botulinum Toxin Use in Children with Cerebral Palsy," *J. Surg. Orthop. Adv.* 13:76-80 (2004); Berweck et al., "Use of Botulinum Toxin in Pediatric Spasticity (Cerebral Palsy)," *Mov. Disord.* 19:S162-S167 (2004); Pidcock, "The Emerging Role of Therapeutic Botulinum Toxin in the Treatment of Cerebral Palsy," *J. Pediatr.* 145:S33-S35 (2004), which are hereby incorporated by reference in their entirety), hip adductor muscle dysfunction in multiple sclerosis (Wissel et al., "Botulinum Toxin Treatment of Hip Adductor Spasticity in Multiple Sclerosis," *Wien Klin Wochenschr* 4:20-24 (2001), which is hereby incorporated by reference in its entirety), neurogenic pain and inflammation, including arthritis, iatro-

genic parotid sialoceles (Capaccio et al., "Diagnosis and Therapeutic Management of Iatrogenic Parotid Sialoceles," *Ann. Otol. Rhinol. Laryngol.* 113:562-564 (2004), which is hereby incorporated by reference in its entirety), and chronic TMJ displacement (Aquilina et al., "Reduction of a Chronic Bilateral Temporomandibular Joint Dislocation with Intermaxillary Fixation and *Botulinum* Toxin A," *Br. J. Oral Maxillofac. Surg.* 42:272-273 (2004), which is hereby incorporated by reference in its entirety). Other conditions that can be treated by local controlled delivery of pharmaceutically active toxin include intra-articular administration for the treatment of arthritic conditions (Mahowald et al., "Long Term Effects of Intra-Articular BoNT A for Refractory Joint Pain," *Annual Meeting of the American College of Rheumatology* (2004), which is hereby incorporated by reference in its entirety), and local administration for the treatment of joint contracture (Russman et al., "Cerebral Palsy: A Rational Approach to a Treatment Protocol, and the Role of *Botulinum* Toxin in Treatment," *Muscle Nerve Suppl.* 6:S181-S193 (1997); Pucinelli et al., "Botulinic Toxin for the Rehabilitation of Osteoarthritis Fixed-Flexion Knee Deformity," *Annual Meeting of the Osteoarthritis Research Society International* (2004), which are hereby incorporated by reference in their entirety). The methods of the present invention are also suitable for the treatment of pain associated with various conditions characterized by the sensitization of nociceptors and their associated clinical syndromes, as described in Bach-Rojecky et al., "Antinociceptive Effect of *Botulinum* Toxin Type A In Rat Model of Carrageenan and Capsaicin Induced Pain," *Croat. Med. J.* 46:201-208 (2005); Aoki, "Evidence for Antinociceptive Activity of *Botulinum* Toxin Type A in Pain Management," *Headache* 43 Suppl 1:S9-15 (2003); Kramer et al., "*Botulinum* Toxin A Reduces Neurogenic Flare But Has Almost No Effect on Pain and Hyperalgesia in Human Skin," *J. Neurol.* 250:188-193 (2003); Blersch et al., "*Botulinum* Toxin A and the Cutaneous Nociception in Humans: A Prospective, Double-Blind, Placebo-Controlled, Randomized Study," *J. Neurol. Sci.* 205:59-63 (2002), which are hereby incorporated by reference in its entirety.

The methods and products of the present invention may be customized to optimize therapeutic properties (See e.g., Chaddock et al., "Retargeted Clostridial Endopeptidases Inhibition of Nociceptive Neurotransmitter Release In Vitro, and Antinociceptive Activity in In Vivo Models of Pain," *Mov. Disord.* 8:S42-S47 (2004); Finn, "*Botulinum* Toxin Type A: Fine-Tuning Treatment of Facial Nerve Injury," *J. Drugs Dermatol.* 3:133-137 (2004); Eleopra et al., "Different Types of *Botulinum* Toxin in Humans," *Mov. Disord.* 8:S53-S59 (2004); Flynn, "Myobloc," *Dermatol. Clin.* 22:207-211 (2004); and Sampaio et al., "Clinical Comparability of Marketed Formulations of *Botulinum* Toxin," *Mov. Disord.* 8:S129-S136 (2004), which are hereby incorporated by reference in their entirety).

## EXAMPLES

The following examples are provided to illustrate embodiments of the present invention but are by no means intended to limit its scope.

### Example 1

#### SDS PAGE

Samples from all intermediate purification steps, as well as pure recombinant protein, were routinely separated and visualized on 8% separating polyacrylamide gels, according to

Laemmli procedure (Laemmli, "Cleavage of Structural Proteins During the Assembly of the Head of Bacteriophage T4," *Nature* 227:680-685 (1970), which is hereby incorporated by reference in its entirety). Protein bands were visualized by Bio-Safe Coomassie G-250 Stain (Bio-Rad, Cat. #161-0786).

### Example 2

#### Western Blotting

Samples for Western blot analysis were separated on 8% SDS-polyacrylamide gels. Followed by separation, proteins were transferred to the Hybond-C nitrocellulose membrane (Amersham Biosciences, Cat. #RPN303C) in 1× Tris/Glycine buffer (Bio-Rad, Cat. #161-0734) supplemented with 20% methanol at 100 volts for 2 hours, 4° C. After the transfer, membrane was rinsed in distilled water and protein bands were visualized by staining with 0.2% Ponceau S in 1% acetic acid for 1 minute. Dye from the membrane was washed away in the Tris-buffered saline/0.1% Tween-20 buffer, pH 7.5, followed by incubation of the membrane in the blocking reagent (5% non-fat powdered milk in Tris-buffered saline/0.1% Tween-20 buffer, pH 7.5) for 16 hours at 4° C. For immunodetection, membrane was incubated with primary antibodies/immune serum at 1:7,000 dilution, in 0.5% non-fat milk in Tris-buffered saline/0.1% Tween-20 buffer, pH 7.5 at room temperature for 2 hours. Membrane was washed (6×5 min) and incubated with secondary antibody at 1:10,000 dilution at room temperature for 25 minutes. After the series of additional washing (6×5 min), immunoreactive bands were visualized using ECL (enhanced chemiluminescence) Plus Western Blotting Reagent (Amersham Biosciences, Cat. #RPN2124) according to manufacturer instructions. Hyperfilm ECL (Amersham Biosciences, Cat. #RPN1674K) was used for autoradiography with the exposure time adequate to visualize chemiluminescent bands. The proteins were identified by comparison with the positive controls and molecular weight protein standards.

### Example 3

#### Evaluation of Recombinant Toxin Yield

The protein concentration of the purified recombinant protein fractions was determined using the BCA Protein assay reagent (Pierce, Cat. #23225) with bovine serum albumin used as standard.

### Example 4

#### In Vitro Toxicity Assay on the Mouse Phrenic Nerve-Hemidiaphragm Preparation

The toxicity of the various recombinant proteins is bioassayed on the mouse phrenic nerve-hemidiaphragm preparation (Simpson et al., "Isolation and Characterization of a Novel Human Monoclonal Antibody that Neutralizes Tetanus Toxin," *J. Pharmacol. Exp. Ther.* 254:98-103 (1990), which is hereby incorporated by reference in its entirety). Tissues are excised and suspended in physiological buffer, aerated with 95% O<sub>2</sub>, 5% CO<sub>2</sub>, and maintained at 35° C. The physiological solution has the following composition: 137 mM NaCl, 5 mM KCl, 1.8 mM CaCl<sub>2</sub>, 1 mM MgSO<sub>4</sub>, 24 mM NaHCO<sub>3</sub>, 1 mM NaH<sub>2</sub>PO<sub>4</sub>, 11 mM D-glucose, and 0.01% gelatin. Phrenic nerves are stimulated continuously (1.0 Hz; 0.1-0.3 msec duration), and muscle twitch is recorded. Toxin-induced

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paralysis is measured as a 50% reduction in muscle twitch response to neurogenic stimulation.

## Example 5

## In Vitro Transcytosis Assay

Cells are grown on polycarbonate membranes with a 0.4  $\mu\text{m}$  pore size in Transwell® porous bottom inserts (Corning-Costar) (FIG. 10) (Zweibaum et al., "Use of Cultured Cell Lines in Studies of Intestinal Cell Differentiation and Function," In: *Handbook of Physiology*, Section 6: "The Gastrointestinal System," Edited by Schulz et al., American Physiological Society, Bethesda, Vol. IV, 223-255; Dharmasathaphorn et al., "A Human Colonic Tumor Cell Line that Maintains Vectorial Electrolyte Transport," *Am. J. Physiol.* 246:G204-G208 (1984); and Dharmasathaphorn et al., "Established Intestinal Cell Lines as Model Systems for Electrolyte Transport Studies," *Methods Enzymol.* 192:354-389 (1990), which are hereby incorporated by reference in their entirety). The cell growth area within each insert is equivalent to 1  $\text{cm}^2$ . Prior to seeding the cells, insert membranes are coated with 10  $\mu\text{g}/\text{cm}^2$  rat tail collagen type I. Collagen stock solution (6.7 mg/ml) are prepared in sterile 1% acetic acid and stored at 4° C. The collagen stock solution is diluted as needed in ice cold 60% ethanol, and 150  $\mu\text{l}$  of the resulting solution containing 10  $\mu\text{g}$  of diluted collagen is added to each well ( $\text{cm}^2$ ).

The collagen solution is allowed to dry at room temperature overnight (ca. 18 hours). After drying, the wells are sterilized under UV light for one hour, followed by a preincubation with cell culture medium (30 minutes). The preincubation medium is removed immediately prior to addition of cells and fresh medium. Cells are plated in the Transwells® at confluent density. The volumes of medium added will be 0.5 ml to the upper chamber and 1.0 ml to the bottom chamber. Culture medium is changed every two days. The cultures maintained in 12 well plates are allowed to differentiate a minimum of 10 days before use. The integrity of cell monolayers and formation of tight junctions is visualized by monitoring the maintenance of a slightly higher medium meniscus in the inserts as compared to the bottom wells.

Formation of tight junctions is confirmed experimentally by assay of the rate of [ $^3\text{H}$ ]-inulin diffusion from the top well into the bottom chamber or by measurement of transepithelial resistance across the monolayer. Transcytosis is assayed by replacement of medium, usually in the top well, with an appropriate volume of medium containing various concentrations of [ $^{125}\text{I}$ ]-labeled protein of interest. Iodination is performed according to Park et al., "Inhalational Poisoning by *Botulinum* Toxin and Inhalation Vaccination with Its Heavy-Chain Component," *Infect. Immun.* 71:1147-1154 (2003), which is hereby incorporated by reference in its entirety. Transport of radiolabeled protein is monitored by sampling the entire contents of opposite wells, which is usually the bottom wells. Aliquots (0.5 ml) of the sampled medium are filtered through a Sephadex G-25 column, and 0.5 ml fractions are collected. The amount of radioactivity in the fractions is determined in a  $\gamma$ -counter. The amount of transcytosed protein is normalized and expressed as fmole/hr/ $\text{cm}^2$ . A minimum of two replicates per condition is included in each experiment, and experiments typically are reproduced at least three times.

## Example 6

## In Vivo Toxicity Assay in Mice

The toxicity of proteins of interest are bioassayed in mice. Proteins are diluted in phosphate buffered saline, including 1

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mg/ml bovine serum albumin, and injected intraperitoneally (i.p.) into animals. The proteins are administered in a 100  $\mu\text{l}$  aliquot of solution at concentrations of 1-100 ng per animal (average weight ~25 g). Any animals that survive exposure to the toxic derivatives are monitored for a total of 2 weeks to detect any non-specific toxicity.

## Example 7

## The BoNT Substrate-Cleavage Assay

Engineered proteins are assayed for endoprotease activity using either mouse brain synaptosomes and recombinant SNAP-25 for BoNT A and BoNT E as the source of the substrate. Native or reduced proteins are incubated with 10 to 50  $\mu\text{g}$  of synaptosomal membranes in reaction buffer containing 50 mM HEPES, pH 7.1, 20  $\mu\text{M}$   $\text{ZnCl}_2$ , and 1% N-octyl- $\beta$ -D-glucopyranoside. Reduced protein are prepared by incubation with DTT (20 mM; 1 hr; room temperature) in phosphate buffered saline. The cleavage reaction is initiated by addition of engineered protein (200 nM final concentration) to substrate, and the reaction is allowed to proceed for 3 hours at 37° C. Endoprotease activity is assayed using Western blot analysis and anti-C-terminal SNAP-25 antibodies (StressGen) for immunodetection of substrate. For visualization of SNAP-25, samples are separated on 16.5% Tris-tricine gels. After separation, proteins are transferred to nitrocellulose membranes (Micron Separations) in Tris-glycine transfer buffer at 50 volts for 1 hr. Blotted membranes are rinsed in distilled water and stained for 1 min with 0.2% Ponceau S in 1% acetic acid. Following a brief rinse with distilled water, molecular weight markers and transferred proteins are visualized. Membranes are destained in phosphate buffered saline-Tween (pH 7.5; 0.1% Tween 20), then blocked with 5% non-fat powdered milk in phosphate buffered saline-Tween for 1 hr at room temperature. Subsequently, membranes are incubated in 0.5% milk with a 1:5,000 dilution of anti-SNAP-25 polyclonal antibody. Secondary antibody is used at 1:20,000 dilution. Membranes are washed again (5 $\times$ ) and visualized using enhanced chemiluminescence (SuperSignal® West Pico, Pierce) according to manufacturer's instructions. Membranes are exposed to film (Hyperfilm ECL, Amersham Biosciences) for times adequate to visualize chemiluminescence bands. Peptides are identified by comparison with known standards. The BoNT B substrate-cleavage assay is performed according to the published protocol (Caccin et al., "VAMP/Synaptobrevin Cleavage by Tetanus and *Botulinum* Neurotoxins is Strongly Enhanced by Acidic Liposomes," *FEBS Lett.* 542:132-136 (2003), which is hereby incorporated by reference in its entirety).

## Example 8

## Cloning Procedures

## Preparation of the DNA Template for PCR

Outlined in detail infra are the procedures used to engineer BoNT A derivatives. A similar strategy for engineering all BoNT derivatives can be carried out.

25  $\mu\text{g}$  of the pure *Clostridium botulinum* type A (Hall strain) genomic DNA was isolated from bacterial pellet separated from the 100 ml of the culture according to Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, Plainview, N.Y.: Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference in its

entirety. DNA was precipitated and dissolved in 1×TE, pH 8.0, at concentration ~0.8 mg/ml.

Genomic DNA, isolated from the mixture of the anaerobic bacteria from the soil, was prepared according to the following protocol: 1000 g of the soil taken from Central Park, New York, were triturated in 2 liters of Dulbecco's phosphate-buffered saline (DPBS) (Invitrogen, Cat. #14190-144). Crude extract was filtered through Kimwipes EX-L wipes (Kimberly-Clark, Neenah, Wis.) and concentrated on a stirred ultrafiltration cell (Millipore (Billerica, Mass.), Cat. #5123) with Ultracel 100-KDa cutoff membrane (Millipore, Cat. #14432) to a final volume of 5 ml. Four liters of cooked meat medium (Difco (Franklin Lakes, N.J.), Cat. #226720), prepared according to manufacturer's protocol were inoculated with 5 ml of concentrated soil extract. After 168-hour incubation at 37° C. without agitation or aeration, a mixture of anaerobic bacteria was separated from the supernatant by centrifugation on Sorwall GS3 rotor (7000 rpm, 25 min., 4° C.) and processed for the isolation of the total genomic DNA on Qiagen (Valencia, Calif.) Genomic tips (Cat. #10262), with additional components also purchased from Qiagen (Cat. #19060, Cat. #19133, Cat. #19101), according to manufacturer's protocol (Qiagen Genomic DNA Handbook). From the cells recovered from 4 liters of the media on ten Qiagen Genomic tips, 6 mg of the genomic DNA were isolated. DNA was precipitated and dissolved in 1×TE, pH 8.0 at concentration ~1 mg/ml.

#### Example 9

##### PCR Amplification of BoNT DNA

25 µg of the mixed genomic DNA or 5 µg of the pure *Clostridium botulinum* type A genomic DNA were used per one 100-µl PCR reaction setting. Reaction conditions were designed according to manufacturer's protocols supplied with Platinum®Pfx polymerase (Invitrogen, Cat. #11708-021). All oligonucleotides and linkers were designed according to the sequence of *botulinum* Neurotoxin type A cDNA obtained from Genebank (Accession #: M30196). Annealing temperatures were deduced from the structure of each set of the oligonucleotides used for the PCR.

#### Example 10

##### Engineering of Non-Expression Vector pLitBoNTA, Carrying Coding Part of BoNT A td

Plasmid encoding *botulinum* Neurotoxin A light chain (pLitBoNTALC) was obtained by the following protocol: The annealed phosphorylated linkers

(SEQ ID NO: 26)  
CBA1: 5' -pCTAGCATGCCATTTGTTAATAAACAATTTAATTATAAG  
and

(SEQ ID NO: 27)  
CBA2: 5' -pGATCCTTATAATTAATTTGTTTATTAAACAAATGGCATG

were subcloned into vector pcDNA3.1/Zeo(+) (Invitrogen, Cat. #V86020), pre-digested with the restriction endonucleases NheI and BamHI and dephosphorylated, resulting in plasmid pcDBoNTALC1. The 620 b.p. PCR product, obtained on genomic DNA as a template with the oligonucleotides

CBA03: (SEQ ID NO: 28)  
5' -TATCTGCAGGGATCCTGTAATGGTGTGGATATTGCTT

ATATAAAAAATTC

and

CBA04: (SEQ ID NO: 29)  
5' -TATGAATTCACCGGTCCGCGGGATCTGTAGCAAATTT

GCCTGCACC

was digested with the restriction endonucleases BamHI and EcoRI and subcloned into pre-digested plasmid pcDBoNTALC1, resulting in plasmid pcDBoNTALC2. The 630 b.p. PCR product, obtained on genomic DNA as a template with the oligonucleotides

CBA05: (SEQ ID NO: 30)  
5' -TATACCGCGGTAACATTAGCACATGAACCTTATACA

TGCTGGACATAGATTATATG

and

CBA06: (SEQ ID NO: 31)  
5' -CATAGAATTCAAACAATCCAGTAAATTTTTAGTTT

AGTAAATTCATATTATTAATTTCTGTATTTTGACC,

was digested with the restriction endonucleases SacII and EcoRI and subcloned into pre-digested plasmid pcD-BoNTLC2, resulting in plasmid pcDBoNTLC3. The annealed phosphorylated linkers

CBA8: (SEQ ID NO: 32)  
5' -pAAT TCTATAAGTTGCTATGTGTAAGAGGGATAAT

ACTAGTCACACTCAATCT

and

CBA9: (SEQ ID NO: 33)  
5' -pCTAGAGATTGAGTGTGACTAGTTATTATCCCTCTTA

CACATAGCAACTTATAG

were subcloned into vector pcDBoNTLC3, pre-digested with the restriction endonucleases EcoR and XbaI and dephosphorylated, resulting in plasmid pcDBoNTALC. The annealed phosphorylated linkers

CBA10: (SEQ ID NO: 34)  
5' -pCGCGTTAGCCATAAATCTGGTTATAAGCGCGC

GAGGTGTTAAGTG

and

CBA11: (SEQ ID NO: 35)  
5' -pCTAGCACTTAACACCTCGCGCCTTATAACCAGA

TTTATGGCTAA

were subcloned into vector pLitmus38i (New England Biolabs, Cat. #N3538S), pre-digested with the restriction endonucleases MluI and NheI and dephosphorylated, resulting in plasmid pLit38iMod. The 1230 b.p. DNA fragment, isolated from the plasmid pcDBoNTALC after its digest with

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restriction endonucleases NheI and ApaI was subcloned into pre-digested and dephosphorylated vector pLit38iMod, resulting in plasmid pLitBoNTALC.

Plasmid encoding *botulinum* Neurotoxin A heavy chain (pLitBoNTAHC) was obtained by the following protocol: The 1450 b.p. PCR product obtained on the genomic DNA as a template with the oligonucleotides

CBA12: (SEQ ID NO: 36)  
5' - AATCTGCAGCCACAGCTGTGGGTACCTTAATTGGTCA

AGTAGATAGATTAAGATAAAGTTAATAATACACTTAGTACAGA

TATACC  
and

CBA13: (SEQ ID NO: 37)  
5' - ATTAGGGCCCTTAATTAAGCGCCGCTCGAGC

TATTACAGTGGCCTTTCTCCCATCCATCATCTACAGGAATAAATTC

was digested with restriction endonucleases ApaI and PstI and subcloned into pre-digested and dephosphorylated vector pLitmus38i, resulting in plasmid pLitBoNTAHC1. Two PCR products, 490 b.p., obtained on the genomic DNA as a template with the oligonucleotides

CBA14: (SEQ ID NO: 38)  
5' - ATACTGCAGTCTAGACCAAGGATACAATGACGATG

ATGATAAGGCA TTAAATGATTTATGTATCAAAGTTAATAATTGGG  
and

CBA15: (SEQ ID NO: 39)  
5' - GCCTAAAAACATAGCCGCTTCGGTCGCTTTATTAAGT

TCTTTACATAGTCTGAAG

and 720 b.p., obtained on genomic DNA as a template with the oligonucleotides

CBA16: (SEQ ID NO: 40)  
5' - TAATAAAGCGACCGAAGCGGCTATGTTTTTAGGCT

GGGTAGAACAATTAG  
and

CBA17: (SEQ ID NO: 41)  
5' - TATAGGGCCCCCTAGGGGTACCTCTATTATCATATATAT

ACTTTAATAATGCATCTTTAAGAC

were mixed with the molar ratio 1:1 and re-PCR'd with oligonucleotides CBA14 and CBA17, resulting in 1170 b.p. PCR product, which was digested with restriction endonucleases PstI and KpnI and subcloned into pre-digested and dephosphorylated vector pLitBoNTAHC1, leading to plasmid pLitBoNTAHC.

Plasmid pLitBoNTA, encoding the entire sequence of BoNT A was obtained by ligating a 2615 b.p. DNA fragment from the vector pLitBoNTAHC, digested with restriction endonucleases XbaI and ApaI into pre-digested and dephosphorylated vector pLitBoNTALC. The size of pLitBoNTA is 6712 b.p. with 3900 b.p. of BoNT A coding sequence.

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## Example 11

Engineering Plasmid pETCBoNTA for the BoNT A td Expression in *E. coli*

pETCBoNTA was obtained by subcloning DNA fragment obtained after the digest of pLitBoNTA vector with NheI and NotI into pre-digested and dephosphorylated expression vector pETcoco2 (Novagen (San Diego, Calif.), Cat. #71148-3) and resulted in 16,194 b.p. BoNT A td expression vector pETCBoNTA.

## Example 12

## Engineering Donor Plasmid pFBSecBoNTA for the Expression of BoNT A td in Insect Cells

pFBSecBoNTA was obtained by the following protocol: 112 b.p. PCR product, synthesized on plasmid pBac-3 (Novagen, Cat. #70088-3) with oligonucleotides

CBA 22: (SEQ ID NO: 42)  
5' - TAAGCGCGCAGAATTCTCTAGAAT GCCCATGTTAAGCGCTATTG  
and

CBA23: (SEQ ID NO: 43)  
5' - TAAGCTAGCGTGATGGTGGTGATGATGGACCATGGCC

and digested with restriction endonucleases BssHIII and NheI was subcloned into pre-digested and dephosphorylated vector pLitBoNTA, resulting in plasmid pLitSecBoNTA. DNA fragment, isolated from pLitSecBoNTA digested with BssHIII and NotI was subcloned into pre-digested and dephosphorylated vector pFastBac™1 (Invitrogen, Cat. #10360-014), resulting in 8764 b.p. plasmid pFBSecBoNTA.

## Example 13

## Engineering the BoNT A Coding Sequence to Enable Expression of Toxin Derivatives

The DNA template was obtained as either pure genomic DNA isolated from *Clostridium botulinum* type A cultures, or as mixed genomic DNA isolated from anaerobic bacteria of soil. BoNT A DNA was amplified by PCR using the high fidelity Platinum Pfx polymerase (Invitrogen, Carlsbad, Calif.). The full-length coding sequence of BoNT A toxic derivative (td) was obtained after consecutive subcloning of five PCR fragments and two phosphorylated linkers into the modified vector pLitmus38i (New England Biolabs, Beverly, Mass.), resulting in plasmid pLitBoNTA. This strategy was used to minimize infidelity during the PCR reaction and to enable the introduction of targeted mutations and endonuclease restriction sites for subsequent engineering of expressed toxin products.

The details of the final construct (td) in comparison with native BoNT A (wt) are shown in FIG. 3, illustrating new restriction sites, eliminated alternative translation sites, and amino acids inserted or substituted. The construct encoding full-length BoNT A td was obtained by ligation of the DNA inserts from the plasmid encoding the toxin heavy chain ("HC") into the plasmid encoding the toxin light chain ("LC"). Plasmid encoding the LC of BoNT A td was generated by consecutive ligation of two PCR products and two phosphorylated linkers into vector pLitmus38i. It contains multiple unique restriction sites upstream from the 5'-end of

the LC sequence, the unique endonuclease restriction site NheI upstream from the first methionine codon, and endonuclease restriction sites for BamHI and EcoRI introduced by silent mutations flanking the minimal catalytic domain (Kadkhodayan et al., "Cloning, Expression, and One-Step Purification of the Minimal Essential Domain of the Light Chain of *Botulinum* Neurotoxin Type A," *Protein Expr. Purif.* 19:125-130 (2000), which is hereby incorporated by reference in its entirety) of the protein at the codons for Lys<sub>11</sub> and Phe<sub>425</sub>. Two additional mutations encoding substitutions Lys<sub>438</sub>>His and Lys<sub>440</sub>>Gln were introduced to minimize non-specific proteolysis of the BoNT A td propeptide during expression. A unique restriction site for XbaI was introduced by silent mutation at the codon. Plasmid, encoding the HC of BoNT A td, was generated by consecutive ligation of two PCR products into the pLitmus38i vector. First, the PCR product encoding the receptor-binding domain of BoNT A was subcloned into the vector pLitmus38i. Second, the PCR product encoding the toxin's translocation domain, obtained by re-PCR of two smaller PCR products was subcloned into plasmid encoding the toxin's receptor binding domain. The final plasmid contains a unique XbaI site at the 5'-end of the coding sequence introduced by silent mutation of the codon Asp<sub>443</sub>, mutation of Lys<sub>444</sub>>Gln to minimize non-specific proteolysis of the BoNT A td propeptide, insertion of codons for four aspartic acid residues between Asn<sub>447</sub> and Lys<sub>448</sub> to create an enterokinase cleavage site, four silent mutations at Ala<sub>597</sub>, Thr<sub>598</sub>, Glu<sub>599</sub>, and Ala<sub>600</sub> to inactivate the putative internal DNA regulatory element, a unique KpnI site introduced at the codon for Gly<sub>829</sub> by silent mutagenesis, and multiple unique restriction sites at the 3'-end of the construct after the stop codon. DNA encoding the Ala<sub>597</sub>-Ala<sub>699</sub> sequence was mutated, because it contains an internal Shine-Dalgarno sequence upstream from internal methionine codon which can result in co-translational contamination of recombinant protein expressed in *E. coli* (Lacy et al., "Recombinant Expression and Purification of the *Botulinum* Neurotoxin Type A Translocation Domain," *Protein Expr. Purif.* 11:195-200 (1997), which is hereby incorporated by reference in its entirety), the initial choice for an expression system to test.

A second full-length BoNT A gene derivative was designed to render the BoNT A atoxic (ad, atoxic derivative). Using site-directed mutagenesis with two synthetic oligonucleotides, a single point mutation, E<sub>224</sub>>A, was introduced into plasmid pLitBoNTA to inactivate the proteolytic activity responsible for BoNT A neurotoxicity resulting in plasmid pLitBoNTAME224A (Kurazono et al., "Minimal Essential Domains Specifying Toxicity of the Light Chains of Tetanus Toxin and *Botulinum* Neurotoxin Type A," *J. Biol. Chem.* 267:14721-14729 (1992); Lacy et al., "Crystal Structure of *Botulinum* Neurotoxin Type A and Implications for Toxicity," *Nat. Struct. Biol.* 5:898-902 (1998); Agarwal et al., "Structural Analysis of *Botulinum* Neurotoxin Type E Catalytic Domain and Its Mutant Glu212>Gln Reveals the Pivotal Role of the Glu212 Carboxylate in the Catalytic Pathway," *Biochemistry* 43:6637-6644 (2004), which are hereby incorporated by reference in their entirety). Atoxic derivatives produced in this way will better preserve the structural moieties responsible for toxin immunogenicity, trafficking, and cell recognition sites.

A third full-length BoNT A derivative was designed to test the utility of the genetic engineering methodology to produce fusion proteins, using GFP as an example. The sequence encoding the minimal catalytic domain of the BoNT A LC (Kadkhodayan et al., "Cloning, Expression, and One-Step Purification of the Minimal Essential Domain of the Light Chain of *Botulinum* Neurotoxin Type A," *Protein Expr. Purif.*

19:125-130 (2000), which is hereby incorporated by reference in its entirety) was excised from plasmid pLitBoNTA and replaced with a GFP-coding sequence to create plasmid pLitGFPBoNTAHC encoding a GFP derivative of BoNT A (gfpd). The GFP-encoding sequence was obtained by PCR with two synthetic oligonucleotides on the plasmid pEGFP-N3 (Clontech, Palo Alto, Calif.). The fusion protein was specifically designed to preserve structural features responsible for cell binding and intracellular trafficking.

All intermediate DNA constructs as well as plasmids pLit-BoNTA, pLitBoNTAME224A, and pLitGFPBoNTAHC were checked by multiple restriction digests. pLitBoNTA and pLitBoNTAME224A were sequenced with twelve BoNT A-specific synthetic oligonucleotides, while pLitGFPBoNTAHC was sequenced with ten BoNT A-specific synthetic oligonucleotides and two GFP-specific oligonucleotides as primers, resulting in a set of overlapping sequences which covered all coding parts of all of the above plasmids. All sequences were demonstrated to be free of unexpected mutations.

#### Example 14

##### Expression of the Recombinant BoNT A Derivatives in *E. coli*

Expression plasmids were transfected into *E. coli* Rosetta-gami B (DE3) competent cells (Novagen, Cat. #71136-3) by the heat-shock method according to manufacturer protocol. Bacterial cultures were grown in LB media containing 50 mg/l carbenicillin, 15 mg/l kanamycin, 12.5 mg/l tetracycline and 34 mg/l chloramphenicol. Various conditions, affecting the plasmid copy number per cell without and with addition of L-arabinose (0.01% final concentration) to the bacterial medium were tested. All bacterial cultures used for protein expression were grown at 37° C. until reaching OD@600 nm ~0.3-0.4. Prior to the induction of the expression, bacterial cultures were split to test influence of the temperature on the yield and quality of the expressed product. Upon induction (OD@600 nm ~0.5-0.7), cultures were grown at 37° C., 25° C., and 12° C. Final IPTG concentration in the growth medium used for induction was 0.5 mM. For the time-course study, samples of the culture at 1, 3, 6, 9, and 12 hours after induction were collected and analyzed. Under the optimal conditions the *E. coli* cultures were incubated overnight in the presence of L-arabinose at 37° C. until reaching OD ~0.4@600 nm. The temperature of the bacterial suspensions was then lowered to 12° C. over one hour, and IPTG was added to a final concentration of 0.5 mM. After induction, culture growth was allowed to continue in a shaker incubator at 12° C. for six more hours. The bacterial pellet was then harvested by centrifugation on Sorwall GS3 rotor (7000 rpm, 25 min., 4° C.) and processed for recombinant protein isolation. Cells kept on ice were resuspended in BugBuster lysis reagent (Novagen, Cat. #70584-4) with the volume ratio cell paste:BugBuster solubilization reagent 1:5. The nucleic acid degradation reagent benzonase was used instead of the mixture sonication (Novagen, Cat. #70746-3), 1000 U/ml final concentration, recombinant lysozyme (Novagen, Cat. #71110-4), 50 U/ml final concentration, and a cocktail of protease inhibitors "Complete" (Roche (Switzerland), Cat. #1697498), 1 tablet/50 ml final concentration were added simultaneously to the paste in the process of resuspension. Approximately 30 minutes after resuspension, the non-viscous lysate was cleared by centrifugation on Sorwall SS34 rotor (17000 rpm, 25 min, 4° C.) and processed for the further protein purification.

An *E. coli* expression system was the first tested for a number of reasons. First, other laboratories have reported expression of recombinant partial length BoNT A domains in this system (Rigoni et al., "Site-Directed Mutagenesis Identifies Active-Site Residues of the Light Chain of *Botulinum Neurotoxin Type A*," *Biochem. Biophys. Res. Commun.* 288: 1231-1237 (2001); Chaddock et al., "Expression and Purification of Catalytically Active, Non-Toxic Endopeptidase Derivatives of *Clostridium Botulinum Toxin Type A*," *Protein Expr. Purif.* 25:219-228 (2002); Lalli et al., "Functional Characterization of Tetanus and *Botulinum Neurotoxins Binding Domains*," *J. Cell Sci.* 112:2715-2724 (1999); Kadkhodayan et al., "Cloning, Expression, and One-Step Purification of the Minimal Essential Domain of the Light Chain of *Botulinum Neurotoxin Type A*," *Protein Expr. Purif.* 19:125-130 (2000); Lacy et al., "Recombinant Expression and Purification of the *Botulinum Neurotoxin Type A Translocation Domain*," *Protein Expr. Purif.* 11:195-200 (1997), which are hereby incorporated by reference in their entirety). A second reason for selecting an *E. coli* expression system is that many recombinant proteins can be expressed in *E. coli* with good yield and stability. A third reason is that non-canonical *E. coli* codons in the BoNT A sequence can be overcome by utilizing a bacterial strain carrying a plasmid encoding tRNA for the rare codons. A fourth reason is that toxicity of the full-length BoNT A to the host can be minimized by using an expression plasmid that allows regulation of the transition from low to medium plasmid copy numbers. Fifth, proper disulfide bridge formation in the recombinant proteins can be optimized by utilizing an *E. coli* strain with *trxB*<sup>-</sup> *gor*<sup>-</sup> mutations. Sixth, degradation of recombinant proteins in the host can be minimized by utilizing an *E. coli* Rosetta-gami strain with *lon*<sup>-</sup> *ompT*<sup>-</sup> mutations, in which two major proteolytic enzymes are inactivated.

Expression plasmids were obtained by single-step subcloning of the coding portion of BoNT A derivatives—td, ad, and *gfpd* into the expression vector pETcoco2 (Novagen, San Diego, Calif.). The resulting constructs contain DNA, encoding sequence MHHHHHHGAS . . . (SEQ ID NO: 44) and flanked with *NheI* unique restriction site in front of the first native methionine codon. The pETcoco system combines the advantages of T7 promoter-driven protein expression with the ability to control plasmid copy number. The pETcoco vectors are normally maintained at one copy per cell. In the single-copy state, pETcoco clones are extremely stable, which is especially important for target genes that are toxic to the host. Copy number can be amplified to 20-50 copies per cell by the addition of L-arabinose to the culture medium. The pETcoco vectors in  $\lambda$ DE3 lysogenic hosts can be induced to increase expression of the target gene by as much as 2,500-fold over background when IPTG is added to the culture media. A 6-His tag was added to each recombinant protein to enable affinity purification. The affinity tag was added to the N-terminus, because prior studies found that addition of an affinity tag to the C-terminus results in loss of the toxin's physiological activity (Shapiro et al., "Identification of a Ganglioside Recognition Domain of Tetanus Toxin Using a Novel Ganglioside Photoaffinity Ligand," *J. Biol. Chem.* 272:30380-30386 (1997), which is hereby incorporated by reference in its entirety), while adding a hexahistidine tag to the N-terminus allowed expression and purification of the light chain domain with retained enzymatic activity (Kadkhodayan et al., "Cloning, Expression, and One-Step Purification of the Minimal Essential Domain of the Light Chain of *Botulinum Neurotoxin Type A*," *Protein Expr. Purif.* 19:125-130 (2000), which is hereby incorporated by reference in its entirety).

All expression constructs were transformed into *E. coli* Rosetta-gami B (DE3) competent cells (Novagen) and were grown in LB media containing ampicillin, kanamycin, tetracycline, and chloramphenicol. Ampicillin was added to select for colonies carrying pETcoco derived bla marker, kanamycin and tetracycline were added to select for thioredoxin (*trxB*) and glutathione reductase (*gor*) mutations, thus improving the chances for proper disulfide bond formation in the *E. coli* cytoplasm (Derman et al., "Mutations that Allow Disulfide Bond Formation in the Cytoplasm of *Escherichia Coli*," *Science* 262:1744-1747 (1993); Prinz et al., "The Role of the Thioredoxin and Glutaredoxin Pathways in Reducing Protein Disulfide Bonds in the *Escherichia Coli* Cytoplasm," *J. Biol. Chem.* 272:15661-15667 (1997), which are hereby incorporated by reference in their entirety). Chloramphenicol was added to the medium to select for colonies containing helper plasmids that provide tRNAs for rare codons, thereby increasing the expression of proteins such as BoNT A encoded by DNA with codons non-canonical for *E. coli*.

Multiple conditions were tested to optimize expression of the BoNT A full length derivatives. Cultures were grown with and without L-arabinose in the media, and different IPTG concentrations were evaluated for induction. Incubation temperatures and time were also optimized for BoNT derivative expression. Under optimal conditions, the *E. coli* cultures were incubated overnight in the presence of L-arabinose at 37° C. until reaching OD ~0.4 at 600 nm. The temperature of the bacterial suspensions was then lowered to 12° C. over one hour, and IPTG was added to a final concentration 0.5 mM. After induction, culture growth was allowed to continue in a shaker incubator at 12° C. for six more hours. The bacterial pellet was then harvested by centrifugation, lysed with Bug-Buster lysis reagent (Novagen) in the presence of nucleic acid degradation reagent benzonase (Novagen), lysozyme, and a cocktail of protease inhibitors. The lysate was cleared by centrifugation and purified by incubation with a Ni-NTA affinity resin. The supernatant and eluate from the Ni-NTA agarose were run on 8% SDS PAGE gels, and analyzed by Western blotting with polyclonal antibodies raised against the full-length BoNT A inactivated toxoid. Rosetta-gami B (DE3) *E. coli* transformed with the empty vector was used as the negative control. Native BoNT A in SDS-PAGE loading buffer was used as the positive control.

FIG. 4 illustrates the results of *E. coli* expression and purification protocols for BoNT A td. The expressed protein was soluble and could be purified using the chelate affinity tag. However, the molecular weight of the recombinant BoNT A td full length propeptide expressed was significantly lower than that of the native full-length BoNT A propeptide. Extensive proteolysis was observed with all purification and expression protocols tested in *E. coli*, even when the toxin derivatives were expressed by the cells in the single-copy plasmid state. This instability may be related to the systems available in *E. coli* for post-translational processing of proteins, with improper folding and disulfide bonding making the recombinant toxins susceptible to degradation. Similar results were obtained when attempting to express the atoxic (ad) and GFP-(*gfpd*) derivatives of BoNT A in *E. coli*. The problems encountered with *E. coli* based expression systems with respect to native protein folding and extensive proteolysis of the recombinant product, may be resolved by modification and optimization of the *E. coli* expression system.

#### Example 15

##### Expression of BoNT A Derivatives in Baculovirus-Based System

Bac-to-Bac® baculovirus expression system (Invitrogen, Cat. #10359-016) was used for the generation of the recom-



binant baculoviruses. A protocol for the insect cell culture was taken from the manual supplied with the kit. Recombinant donor plasmids were transformed into Max Efficiency DH10Bac™ competent cells (Invitrogen, Cat. #10361-012). Colonies containing recombinant bacmid were identified by 5 disruption of the *lacZa* gene and selected by the absence of developing blue color, while growing on the plate with the chromogenic substrate *Bluo-gal* (Invitrogen, Cat. #15519-028). High molecular weight DNA was isolated from the selected colonies on DNA plasmid purification system (Qiagen, Cat. #12245). Transposition of the DNA of interest into baculovirus genome was confirmed by PCR on high molecular weight DNA with oligonucleotides CBA14 and CBA17, resulting in amplification of 1170 b.p. DNA band in 15 samples where transposition took place. Bacmids were used to transfect serum-free medium adapted Sf9 insect cells (Invitrogen, Cat. #11496-015) to produce baculoviruses. Transfection was performed by the following protocol:  $9 \times 10^5$  cells were seeded per one 35-mm well in 2 ml of unsupplemented Grace's insect cell culture medium (Invitrogen, Cat. #11595-030). Cells from a 3 to 4 day-old suspension culture in mid-log phase with a viability of >97% were used for experiment. Cells were attached to the plastic at 27° C. for at least one hour in advance and transfected with the lipophylic complexes, formed after mixing bacmid with Cellfectin® transfection reagent (Invitrogen, Cat. #10362-010), according to the protocol supplied by the manufacturer. 72 hours after transfection, the supernatant containing recombinant baculoviruses was harvested and separated from the cells by low-speed centrifugation (Sorwall GS 3 Rotor, 2000 rpm, 20 min, 4° C.). The supernatant represents the primary baculoviral stock. Amplification of this baculoviral stock and viral plaque assay was performed according to the protocol supplied by the manufacturer. Experiments related to identification of the optimal MOI and time-course studies of recombinant protein expression were also performed according to the manufacturer recommendations.

For the purpose of protein expression, Sf9 cells were grown as a shaken culture in a SF900 II serum-free medium (Invitrogen, Cat. #10902-088) at 27° C. in humidified atmosphere. At the density of the cell culture  $\sim 1.2 \times 10^6$ /ml, baculovirus stock in the same medium was added to suspension at MOI  $\sim 0.1$ . Incubation continues for another  $\sim 50$  hours, after which medium was separated from the cells by centrifugation (Sorwall GS 3 Rotor, 2000 rpm, 20 min, 4° C.) and further processed for the protein purification by the procedure outlined below. Sf9 cells are very sensitive to growth conditions. They require a constant temperature of  $27 \pm 1$ ° C., good aeration of shaking cultures, and a sterile environment. If ambient temperatures rise above 27° C., refrigeration is required in the incubator used. An incubator sufficiently large to produce sufficient quantities of BoNT derivatives for biological testing is recommended.

To avoid poisoning of the insect cell host, the BoNT A td construct was modified by adding a signal peptide to provide for secretion of the recombinant proteins to the medium. Targeting the recombinant toxins for secretion also resulted in proper disulfide bond formation between the toxin's light and heavy chains. Improvements to this expression system were tested as described infra.

To increase the total yield of the recombinant protein, donor recombinant baculovirus plasmids and bacmids were generated with an expression cassette that allows expression of the recombinant protein to be driven by two separate and independent promoters simultaneously, p10 and PH (donor plasmid pFastBac™ Dual, Invitrogen, Cat. #10712-024).

To stabilize and increase the titer of the recombinant baculoviral stock, an approach outlined in BaculoDirect® Expression System protocol (Invitrogen (Carlsbad, Calif.), Cat. #12562-021) was used that allows negative selection to remove non-recombinant baculovirus that tend to appear in amplified stocks over the time. To improve purification of the toxins, the affinity of the recombinantly expressed proteins for Ni-NTA resin was increased by generating additional BoNT constructs with longer N-terminal His tags.

The advantages of a baculovirus expression system include proper disulfide bridge formation which has been demonstrated for numerous recombinant proteins in this system; protein purification, which is facilitated when serum-free culture medium is utilized and the expressed proteins contain a short secretory signal and affinity tag; physiological activity similar to native progenitors can be retained in the expressed products; and the absence of endotoxins endogenous to *E. coli*, which facilitates biological testing and therapeutic use of the expressed proteins (Allen et al., "Recombinant Human Nerve Growth Factor for Clinical Trials: Protein Expression, Purification, Stability and Characterisation of Binding to Infusion Pumps," *J. Biochem. Biophys. Methods.* 47:239-255 (2001); Curtis et al., "Insect Cell Production of a Secreted form of Human Alpha(1)-Proteinase Inhibitor as a Bifunctional Protein which Inhibits Neutrophil Elastase and has Growth Factor-Like Activities," *J. Biotechnol.* 93:35-44 (2002), which are hereby incorporated by reference in their entirety). The disadvantages of this system are its cost, time-consuming procedures, and generally the yield of proteins is not as high as in *E. coli*. Furthermore, because the regulated exocytosis machinery is well preserved across eukaryotic species from yeast to mammals, expression of BoNT A in this system can potentially lead to the host poisoning and cellular death. Nonetheless, since Clostridial neurotoxins are known to pass through epithelial cells by transcytosis without any toxic affects (Simpson, "Identification of the Major Steps in Botulinum Toxin Action," *Annu. Rev. Pharmacol. Toxicol.* 44:167-193 (2004); Park et al., "Inhalational Poisoning by Botulinum Toxin and Inhalation Vaccination with Its Heavy-Chain Component," *Infect. Immun.* 71:1147-1154 (2003), which are hereby incorporated by reference in their entirety), and the toxin constructs described herein are designed to remain in the single-chain propeptide form until processed to dichain mature form by enterokinase, this system is worth 45 further testing.

Plasmid constructs for expression of BoNT A derivatives in this system were subcloned into the donor vector pFastBac™1 (Invitrogen). To facilitate secretion of the recombinant proteins into the media and to allow purification of the recombinant proteins on Ni-NTA agarose, a DNA sequence coding the gp64 signal peptide and a hexahistidine affinity tag MPMLSAIVLYVLLAAAHAHSAFAAMVHHH-HHHSAS . . . (SEQ ID NO: 45), flanked with unique NheI restriction site in front of the first native methionine codon, was introduced by cloning of PCR product into all constructs. FIG. 5 provides a schematic representation of the BoNT A derivatives targeted for expression in the baculovirus system. The signal peptide shown in the illustrated recombinant proteins is removed by secretase processing during intracellular trafficking (von Heijne, "Signals for Protein Targeting Into and Across Membranes," *Subcell. Biochem.* 22:1-19 (1994), which is hereby incorporated by reference in its entirety). Expression of the genes in the vector pFastBac™1 is controlled by the *Autographa californica* multiple nuclear polyhedrosis virus (AcMNPV) polyhedrin (PH) promoter. Recombinant donor plasmids were transformed into DH10Bac® *E. coli* competent cells. Once the pFastBac™

based expression plasmid is in cellular cytoplasm, transposition occurs between the arms of mini-Tn7 element flanking the expression cassette in pFastBac™ based vector and the mini-attTn7 target site on the baculovirus shuttle vector (bacmid), already present in the cells. This event generates a recombinant bacmid. Transposition requires additional proteins supplied by helper plasmids also present in competent cells. Selection of the recombinant bacmid clones was performed visually (by size and color). The molecular nature of the isolated DNAs was confirmed by PCR with the specific oligonucleotide primers.

Recombinant bacmids and negative control bacmids (obtained as a result of transposition with empty donor plasmids) were transfected into Sf9 insect cells with the lipophilic reagent Cellfectin (Invitrogen). After 96 hours the recombinant baculoviral stock was harvested and used for infection of freshly seeded Sf9 cells.

Secondary baculoviral stock was used for multiple purposes, which include, testing the expression of recombinant proteins, amplifying recombinant baculoviruses and generating tertiary stock for future use, calculating the titer of recombinant baculoviruses, identifying the optimal ratio for multiplicity of infection (MOI), and establishing the optimal time course for protein expression. Baculovirus titer was calculated for each newly amplified baculoviral stock. For all BoNT A constructs tested, the optimal MOI was found to be ~0.1 pfu per cell and the optimal time for the protein harvest was found to be ~50 hours after infection. When recombinant proteins were allowed to accumulate in the media for 72 hours or longer, a significant portion of the recombinant protein was degraded due to virus-induced cellular lysis.

FIG. 6 illustrates the protein expression results for the toxic (td), atoxic (ad), and GFP-linked (gfpd) full-length propeptide derivatives of BoNT A (cultures harvested at 50 hrs). All recombinant proteins were soluble and secreted into the media, could be purified by binding to the affinity resin, and have the expected mobility on SDS PAGE comparable to the mobility of single chain wt BoNT A. The recombinant BoNT A derivatives expressed using these conditions were free of degradation products recognized by the polyclonal antibody.

#### Example 16

##### Enterokinase Processing and LC-HC Disulfide Bridges

FIG. 7 illustrates a dosage-titration curve for cleavage of the propeptide constructs with recombinant enterokinase (rEK), using the td derivative as an example. For the processing of the single-chain (SC) protein, different amounts have been applied to the BoNT A td. Using 0.5 U of the enzyme at 20° C. for 8 hours was found to completely digest 1 µg of the sc BoNT A td.

To evaluate whether the disulfide bridges between the light and heavy chains of the recombinant proteins were properly formed, the recombinant propeptide derivatives were compared on reducing and non-reducing gels after digestion with excess rEK. Western blots were probed with polyclonal antibodies raised against native full-length BoNT A toxoid. The results of this experiment, shown in FIGS. 8A and 8B, demonstrate that all of the recombinant propeptides were processed into a two-subunit form by rEK, and that the subunits could be readily separated after reduction of the disulfide bridges, as expected.

Expression of a GFP-linked derivative of BoNT A is demonstrated by the green fluorescence of Sf9 cells 12 hours after infection with the recombinant baculovirus expressing BoNT

A gfpd (FIGS. 8C and 8D). The significant difference in the background of FIG. 8C (recombinant baculovirus expressing BoNT A gfpd with secretion signal) versus FIG. 8D (recombinant baculovirus expressing GFP control), is believed to result from the secretion of the fluorescent recombinant protein into the media.

#### Example 17

##### Purification of the Recombinant BoNT A Derivatives

Methods to purify reasonable quantities of the full-length BoNT A derivatives were developed using BoNT A td as an example. Though the recombinant protein was found to bind to Ni-NTA resin (FIG. 5), the affinity was not sufficient to establish a one step purification scheme. The protein bound to the Ni-NTA resin in 5 mM imidazole, but was eluted from the affinity column by 40 mM imidazole. At this concentration of imidazole, there are other proteins present in the eluate, and therefore additional steps are needed to separate recombinant protein from contaminants.

Similar results were observed with the minimal essential domain of BoNT A expressed and purified in *E. coli* (Kadkhodayan et al., "Cloning, Expression, and One-Step Purification of the Minimal Essential Domain of the Light Chain of *Botulinum Neurotoxin Type A*," *Protein Expr. Purif.* 19:125-130 (2000)), which is hereby incorporated by reference in its entirety). The stable minimal essential domain of the LC expressed with two 6-His tags on the N- and C-termini of the protein was eluted from the Ni-NTA column by 90 mM imidazole, still a relatively low concentration of affinity eluant. Poor accessibility of the affinity tags may explain these difficulties. Interestingly, there were two fractions of the same protein from the affinity column, with the second fraction eluted in 250 mM imidazole. While 90 mM imidazole fraction was enzymatically active, as was shown in SNAP-25 peptide cleavage assay, the higher concentration imidazole eluate was not. Denaturation of the protein may explain its absence of activity and high affinity for the chelate matrix.

A multi-step protocol was developed for purifying BoNT A td to homogeneity. Sf9 cells (viable cells count before infection ~1.2·10<sup>6</sup>/ml) grown at 27° C. in SF900II serum-free media in humidified atmosphere at 125 rpm in shaking culture were harvested and separated from the medium. At ~50 hours after infection with recombinant baculovirus (MOI~0.1), the medium was collected, precipitated with ammonium sulfate or concentrated, dialyzed, and subjected to sequential DEAE-sepharose chromatography, MonoS chromatography, Ni-NTA affinity chromatography, and FPLC-based gel filtration chromatography. FIG. 9 illustrates the results of protein purification. The yield of the pure recombinant protein was 0.35 mg from one liter of serum-free medium. The pure protein eluted from the final gel-filtration column was competent for further processing with rEK. After the rEK cleavage, chloride ions containing buffer need to be substituted with phosphate or HEPES-based buffer to avoid instability of the recombinant toxin derivatives.

Approximately 2 ml of supernatant or cleared lysate was concentrated on Amicon Ultra-4 centrifugal filter device (Millipore, Cat. #UFC803024) to ~1 ml. The concentration procedure was done in parallel with multiple rounds of buffer substitution aimed at removing substances which could contribute to Ni<sup>2+</sup>-stripping from the affinity resin. Final buffer composition was equal to the Ni-NTA Equilibration Buffer (infra). 20 µl of Ni-NTA suspension equilibrated in the Ni-NTA Equilibration Buffer (1:1 v/v) was added to the sample, followed by the sample incubation on the rotating platform

for 1 hour. After incubation, affinity matrix was separated from the supernatant by centrifugation (3000 g, 1 min), and washed three times with Ni-NTA Equilibration Buffer, followed by centrifugation. The washing buffer was aspirated and the resin was resuspended in ~200  $\mu$ l of SDS-PAGE loading buffer. The liquid was used for the further analysis by SDS PAGE and Western blotting.

TABLE 1

BoNT A td Purification Composition of the buffers used:
DEAE Sepharose Equilibration Buffer: 20 mM NaH <sub>2</sub> PO <sub>4</sub> , 1 mM EDTA, pH 8.0
DEAE Sepharose Wash Buffer: 50 mM NaCl, 20 mM NaH <sub>2</sub> PO <sub>4</sub> , pH 8.0
DEAE Sepharose Elution Buffer: 500 mM NaCl, 20 mM NaH <sub>2</sub> PO <sub>4</sub> , pH 8.0
Mono S Equilibration Buffer: 20 mM NaH <sub>2</sub> PO <sub>4</sub> , pH 6.8
Mono S Wash Buffer: 25 mM NaCl, 20 mM NaH <sub>2</sub> PO <sub>4</sub> , pH 6.8
Mono S Elution Buffer: 300 mM NaCl, 20 mM NaH <sub>2</sub> PO <sub>4</sub> , pH 6.8
Ni-NTA Equilibration Buffer: 5 mM imidazole, 50 mM NaH <sub>2</sub> PO <sub>4</sub> , 300 mM NaCl, pH 8.0
Ni-NTA Wash Buffer I: Same as above but made up with 10 mM imidazole
Ni-NTA Wash Buffer II: Same as above but made up with 20 mM imidazole
Ni-NTA Elution Buffer: Same as above but made up with 60 mM imidazole
HiLoad 16/60 Superdex 200PG Equilibration Buffer: 50 mM NaCl, 20 mM Tris-HCl, pH 7.5

All concentration, dialysis, and chromatography steps were performed at 4° C. 300 ml of the conditioned insect medium was either concentrated on the stirred ultrafiltration cell (Millipore, Cat. #5123) with Ultracel 100-KDa cutoff membrane (Millipore, Cat. #14432) to the final volume 5 ml, or the total protein from the medium was precipitated by addition of ammonium sulfate (60 g/100 ml) with slow stirring. Pellet was separated from the supernatant by centrifugation (5000 g, 20 min, 4° C.) and dissolved in 5 ml of DEAE-Sepharose Equilibration Buffer. Recombinant protein recovered from the first procedure was less denatured, and this procedure is preferable for future work. Scale-up production of the BoNT derivatives for biological testing could be accomplished with Tangential Flow Concentration System (Pellicon 2, Millipore, Cat. #XX42PLK60) which would enable large volumes of the media to be processed. During membrane concentration or ammonium sulfate precipitation, an insoluble precipitate forms from the pluronic surfactant included in the SF900 II media (to prevent cellular aggregation and to reduce shearing forces, thereby improving the stability of the Sf9 insect cells). The insoluble pluronic pellet was removed by centrifugation of the concentrate/ammonium sulfate precipitate (5000 g, 20 min, 4° C.), and recombinant toxin in the pellet was recovered by extracting twice with DEAE-Sepharose Equilibration Buffer, followed by centrifugation.

Recovered combined supernatant was dialyzed against 100 $\times$  volumes of DEAE-Sepharose Equilibration Buffer for 16 hours, separated from the residual pellet by centrifugation, and loaded on a column (1.5 $\times$ 10 cm) packed with DEAE-Sepharose Fast Flow (Amersham Biosciences, Cat. #17-0709-01) pre-equilibrated in the same buffer at a buffer flow rate of 0.5 ml/min. The column was washed with ~15 volumes of DEAE Sepharose Wash Buffer and then a linear gradient of 100 ml DEAE Sepharose Wash Buffer: 100 ml DEAE Sepharose Elution Buffer was applied. 4-ml fractions were collected and their content was analyzed by PAGE and Western blotting. Fractions containing recombinant protein were combined and dialyzed against 100 $\times$  volumes of the Mono S

Equilibration Buffer for 16 hours. Resulting combined dialyzed was cleared by centrifugation and loaded at 1 ml/min on MonoS 5/50 GL FPLC column (Amersham Biosciences, Cat. #17-5168-01), pre-equilibrated in the same buffer. Column was washed with 100 ml of Mono S Wash Buffer and the linear gradient of 100 ml Mono S Wash Buffer: 100 ml Mono S Elution Buffer was applied. 2-ml fractions were collected and their content was analyzed by PAGE and Western blotting.

The fractions containing recombinant protein were combined, concentrated on the stirred ultrafiltration cell with Ultracel 100-KDa cutoff membrane to the final volume of 20 ml with sequential buffer change to Ni-NTA Equilibration Buffer. Combined fractions were loaded on a 1 $\times$ 4 cm column with Ni-NTA affinity resin (Novagen, Cat. #70666) pre-equilibrated in the same buffer at a buffer flow rate of 1 ml/min. The column was sequentially washed with 100 ml of Ni-NTA Wash Buffer I, followed by 100 ml of Ni-NTA Wash Buffer II, and protein was eluted from the column by 50 ml of Ni-NTA Elution Buffer. All fractions were analyzed by PAGE and Western blotting. Elution fractions enriched in recombinant protein were concentrated sequentially on the stirred ultrafiltration cell with Ultracel 100-KDa cutoff membrane followed by Amicon Ultra-4 centrifugal filter device (Millipore, Cat. #UFC803024) to a final volume of 1 ml and loaded on the FPLC HiLoad 16/60 Superdex 200PG gel filtration column (Amersham Biosciences, Cat. #17-1069-01), equilibrated with HiLoad 16/60 Superdex 200PG Equilibration Buffer. The buffer flow rate was 1 ml/min and 1-ml fractions from the column were collected and analyzed by PAGE and Western blotting.

The multi-step protocol developed for BoNT A td purification provided a yield ~0.35 mg of pure protein per liter of serum-free medium. It is believed that this procedure can be optimized to provide yields in the range of 0.7-0.9 mg/l. Several reasons may explain the relatively low yield in the purification of BoNT A td: 1) Significant amounts of the recombinant toxin may be lost due to non-specific adsorption to the brand-new separation media; 2) The delays which occurred between purification steps may have resulted in degradation of recombinant toxins. These delays were impossible to avoid during the initial purification attempts, because it was necessary to analyze the recombinant products before proceeding to the next purification step. The following modifications, aimed at simplifying and improving the current purification scheme, were tested.

#### Example 18

##### Biological Testing of the Recombinant BoNT A Derivatives

Two types of experiments were performed to assess whether the recombinant toxins retained the biological activities of native toxin. These were performed using the BoNT A td derivative, which was produced in sufficient quantities for biological testing. In the first test, recombinant BoNT A td was administered to mice by the intravenous route (~1 ng per mouse) and the time-to-death was monitored. Death was observed approximately 12 minutes after injection. Prior symptoms of muscular weakness or paralysis were not obvious. In the second test, recombinant BoNT A td was added to mouse phrenic nerve-hemidiaphragm preparations, and its ability to inhibit acetylcholine release evoked by stimulation of the nerve trunk (0.2 Hz) was evaluated by monitoring muscle twitch. At a concentration of ~1 $\times$ 10<sup>-11</sup> M, recombinant BoNT A td produced neuromuscular blockade in 167 $\pm$ 17

min (n=4). To insure that the blockade could be attributed to a *botulinum* toxin-type action, a final experiment was done, in which the polypeptide was pre-incubated (room temperature, 60 min) with rabbit antiserum raised against the carboxy terminal half of the native BoNT A heavy chain (i.e. receptor-binding domain). In these experiments (n=3), there was no neuromuscular blockade, even when the tissues were monitored for ca. 400 minutes. The pharmaceutical preparation marketed by Allergan Inc., as "BoTox" produces neuromuscular blockade in 100 minutes at a concentration of approximately  $1 \times 10^{-11}$  M (60 Units per ml). The BoNT A, B, and G recombinant products produced by Rummel (supra) require 60 to 1000 times more BoNT to effect neuromuscular blockade in a similar timeframe.

#### Example 19

##### Preparation and Modification of the BoNT Gene Constructs

DNA and protein sequences for Clostridial toxins are accessed from the Gene bank. Constructs encoding full-length toxins are available from a number of laboratories. These known sequences and constructs provide an efficient starting point for the planned genetic manipulations.

The first type of mutation introduced is designed to improve toxin stability by site-directed mutations of low specificity protease-sensitive residues within the light-heavy chain junction region, thereby reducing susceptibility to non-specific activation and poisoning of the host organism. The second type of mutation will be introduced to create a highly specific enterokinase cleavage site between the light and heavy chains, thereby enabling external control of the cleavage event leading to toxin maturation. The third type of mutation to be introduced is designed to silently inactivate DNA elements affecting RNA transcription and protein expression in the system of choice. The fourth type of modification is designed to introduce unique restriction sites that enable easy manipulation of the toxin gene, its protein products, and chimeric proteins which may be created as required.

The modified BoNT A constructs used to produce the BoNT A toxic derivative (td) described infra demonstrates the feasibility of these methods. The objective is to determine how to best adapt the methods developed for BoNT A to producing other Clostridial neurotoxins, and in the process optimize the methodology and create a library of toxin derivatives with customized biological properties. Molecular cloning techniques are generally known in the art, and other full-length neurotoxins have successfully been cloned (Ichtchenko et al., "Alpha-Latrotoxin Action Probed with Recombinant Toxin: Receptors Recruit Alpha-Latrotoxin but do not Transduce an Exocytotic Signal," *EMBO J.* 17:6188-6199 (1998), which is hereby incorporated by reference in its entirety).

#### Example 20

##### Creation and Expression of Recombinant BoNT Molecules Minimally Modified to Eliminate Toxicity

To create atoxic derivatives ("ad") that most closely resemble the native toxin with respect to their structure and physiologic activity, a single amino acid point mutation is introduced into the active site of the toxin's metalloprotease catalytic domain. Though most toxin features in this molecule remain the same as in the native toxin, it is devoid of toxicity, because it is unable to cleave its substrate in the synaptic

exocytosis machinery. The atoxic derivatives thus created are superior to other BoNT preparations being developed as vaccines, because of their structural similarity to native toxin, and their ability to generate an immune response at diverse sites along the native toxin's absorption and trafficking route. Because these derivatives are likely to compete with native toxin for the same binding sites and trafficking pathways, they may also be superior to antibody preparations as antidotes to BoNT poisoning.

The cloning and expression strategies developed can be duplicated as closely as possible in applying the methods to BoNT B and E, thereby minimizing the possibility of creating significant molecular alterations in the atoxic derivatives which might decrease their therapeutic potential. The validity of this assumption is demonstrated supra with the BoNT A atoxic derivative (ad), which has been shown to be essentially identical to native BoNT A with respect to expression level, antibody recognition, disulfide bonding, cleavage with enterokinase, and binding to Ni-NTA affinity resin.

An outline of the steps necessary to produce atoxic derivatives of BoNT B and E is as follows. Constructs encoding the atoxic derivatives (ad) of BoNT B and E are obtained by site-directed mutagenesis of BoNT B and BoNT E td constructs, using procedures established for BoNT A ad, as detailed supra. Expression constructs for BoNT B ad and BoNT E ad in the different expression systems to be tested are prepared using a protocol similar to that established for BoNT A ad, as detailed supra. The expression system, purification protocol, and rEK-cleavage protocol for BoNT B and E ad replicate the optimized procedure developed for BoNT A td and ad, as outlined supra. The expression and purification system chosen to produce the atoxic derivatives is based on the quality and yield produced by the expression systems tested.

The atoxic derivatives are tested in a substrate cleavage assay using SNAP 25 or VAMP as substrates. Though no residual proteolytic activity in the single-amino acid mutated atoxic derivatives is expected, if the rate of substrate hydrolysis for any particular atoxic derivative is significantly higher than zero, a second amino acid residue, corresponding to His<sub>227</sub> in BoNT A, is mutated at the toxin's active site before proceeding for its further biological and functional characterization.

#### Prophetic Example 21

##### Preparation of DNA Starting Material for BoNT Serotypes B and E

DNA template for all BoNT serotypes for PCR amplification can be obtained from either pure *Clostridium* cultures (serotype-specific) or soil-derived anaerobic cultures from which mixed genomic DNA as a starting material may be prepared. High fidelity Platinum®Pfx polymerase is used for all PCR reactions to minimize amplification errors. BoNT B and E serotypes are described in subsequent examples.

#### Prophetic Example 22

##### Constructs for BoNT B and E

A set of oligonucleotides similar to those used for obtaining the full-length coding sequence of BoNT A td may be designed for BoNT B and E, using sequences available from Gene bank (accession number M81186 for BoNT B and X62683 for BoNT E). Sequences are carefully evaluated for unwanted DNA regulatory elements and other features that

could affect protein expression in *E. coli*, baculovirus, and *Pichia pastoris* expression systems, and such elements eliminated by silent site-directed mutagenesis. Additional mutations targeted to remove low-specificity proteolysis sites in the toxin's LC-HC junction are introduced, and to introduce an enterokinase cleavage site in the LC-HC junction region. Based on the toxin sequence alignments and domain structure illustrated in FIGS. 1-3, gene regions which can be modified without affecting the recombinant toxin's biological properties are identified, and Nhe I, Xba I, Kpn I, and Xho I restriction sites are introduced, similar to the design scheme executed for BoNT A td. If such mutations are impossible to make through silent mutagenesis, restriction sites are introduced via neutral amino acid insertion into structurally flexible portions of the protein sequence. Any redundant restriction sites created are eliminated by silent site-directed mutagenesis. BoNT DNA sequences that can cause premature termination of gene transcription in the expression systems or interfere with the protein expression are also modified. Expression in *Pichia pastoris* (Henikoff et al., "Sequences Responsible for Transcription Termination on a Gene Segment in *Saccharomyces Cerevisiae*," *Mol. Cell Biol.* 4:1515-1520 (1984); Irniger et al., "Different Classes of Polyadenylation Sites in the Yeast *Saccharomyces Cerevisiae*," *Mol. Cell Biol.* 11:3060-3069 (1991); Scorer et al., "The Intracellular Production and Secretion of HIV-1 Envelope Protein in the Methylophilic Yeast *Pichia Pastoris*," *Gene* 136:111-119 (1993), which are hereby incorporated by reference in their entirety) requires the elimination of such sequences by designing a set of PCR oligonucleotide primers which can suppress premature termination of transcription from AT-rich templates. These procedures produce constructs containing modified coding sequences for BoNT B and BoNT E td, which are used in subsequent expression studies.

Endonuclease restriction digests are used to check all intermediate DNA products. The final full-length DNA is sequenced to prove absence of unwanted mutations.

Molecular biocomputing software, supplied by the DNAs-tar is used to analyze and compare DNA and protein sequences. This will also optimize the creation of synthetic oligonucleotides and optimal reaction conditions for all reactions of PCR amplification.

#### Prophetic Example 23

##### Expression, Purification, and Biochemical Analysis of Toxic Derivatives

Expression and purification of full-length, functionally active toxins has proven difficult in laboratories using alternative construct designs and expression systems. The ideal construct and expression system preferably do not segregate Clostridial toxins, because they contain coding sequences non-typical for the host; are not poisoned by entry of active toxin into the cytosol where it may disrupt the apparatus for regulated exocytosis, which is similar in most eukaryotes; and allow normal post-translational modification of the expressed toxins, particularly formation of disulfide bridges.

Two expression systems were tested for each toxin serotype A: baculovirus and *E. coli*. Because the baculovirus expression system was found to be most effective for expressing full-length BoNT A td, this was used as a starting point and benchmark for all the toxins. Though much concentration was centered on the baculovirus expression system, alternatives were evaluated, taking scale-up and cost into consideration, and work can be performed to optimize expression of

all serotypes in these systems, as well as in other expression systems such as *Pichia pastoris*.

Work that was performed to optimize expression and purification are described supra. The effect of these modifications on nativity of the toxin was evaluated in each case.

#### Prophetic Example 24

##### *E. coli* Expression System

Attempts to produce full-length BoNT A in *E. coli* resulted in a major C-terminally truncated propeptide which was significantly smaller than expected for the BoNT A propeptide (FIG. 4). In the future, the C-terminal composition of this product will be analyzed by microsequencing, identifying putative proteolytic cleavage sites specific to the *E. coli* system, and redesigning the pETcoco expression construct with amino acid substitutions designed to suppress this effect. It is possible the proteolysis site is similar to that recognized by trypsin, which has been demonstrated to cleave within the C-terminal BoNT A receptor-binding domain when applied in excessive amounts (Chaddock et al., "Expression and Purification of Catalytically Active, Non-Toxic Endopeptidase Derivatives of *Clostridium Botulinum* Toxin Type A," *Protein Expr. Purif.* 25:219-228 (2002), which is hereby incorporated by reference in its entirety). Expression of re-designed construct will use the advanced Rosetta-gami B (DE3) *E. coli* strain, as described infra.

#### Prophetic Example 25

##### Targeting Secretion to the Periplasm

Targeting recombinant proteins for secretion to the *E. coli* periplasm can improve stability and post-translational disulfide bond formation. The coding portion of the BoNT A td sequence will be subcloned into pET39b(+) vector (Novagen, Cat. #70090-3) which contains the signal required for export and periplasmic folding of target proteins. This system is designed for cloning and expression of peptide sequences fused with the 208 amino acids DsbA•Tag<sup>TM</sup>. DsbA is a periplasmic enzyme that catalyzes the formation and isomerization of disulfide bonds (Rietsch et al., "An In vivo Pathway for Disulfide Bond Isomerization in *Escherichia coli*," *Proc. Natl. Acad. Sci. USA* 93:13048-13053 (1996); Sone et al., "Differential In vivo Roles Played by DsbA and DsbC in the Formation of Protein Disulfide Bonds," *J. Biol. Chem.* 272:10349-10352 (1997); Missiakas et al., "The *Escherichia coli* DsbC (xprA) Gene Encodes a Periplasmic Protein Involved in Disulfide Bond Formation," *EMBO J.* 13:2013-2020 (1994); Zapun et al., "Structural and Functional Characterization of DsbC, a Protein Involved in Disulfide Bond Formation in *Escherichia coli*," *Biochemistry* 34:5075-5089 (1995); Raina et al., "Making and Breaking Disulfide Bonds," *Annu. Rev. Microbiol.* 51:179-202 (1997), which are hereby incorporated by reference in their entirety). It is possible that the degradation of BoNT A described infra occurs as a result of *E. coli* incompetence to properly form disulfide bridges for proteins which accumulate in the cytoplasm. The DsbA vector may enhance solubility and proper folding of recombinant BoNTs in the non-reducing periplasmic environment (Collins-Racie et al., "Production of Recombinant Bovine Enterokinase Catalytic Subunit in *Escherichia coli* Using the Novel Secretory Fusion Partner DsbA," *Biotechnology* 13:982-987 (1995), which is hereby incorporated by reference in its entirety). Though the yield of recombinant proteins

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targeted to the periplasm is usually low, periplasmic expression in *E. coli* is worth continued consideration.

## Prophetic Example 26

*Pichia pastoris* Expression System

Multi-copy *Pichia pastoris* expression kit (Invitrogen, Cat. #K1750-01) is used to obtain recombinant proteins. Recombinant plasmid on the backbone of the vector pPIC9K, carrying gene of interest and targeted for incorporation into *Pichia* genome is digested by restriction endonuclease Sall for linearization and transformed in the *Pichia* strains GS115 and KM71 by spheroplasting method with zymolyase, according to the supplied manufacturer's protocol. Primary and secondary rounds of the transformants selection on the histidine-deficient medium and in the presence of Geneticin is performed according to the same protocol. Protein expression is induced by the addition of methanol (0.5% final concentration) into the culture medium. Disrupted cells and medium are analyzed by SDS-PAGE and Western blotting.

Though the baculovirus expression system was found to provide satisfactory level of protein expression of BoNT A, recombinant protein expression in *Pichia pastoris* (methylotrophic yeast capable of metabolizing methanol as its sole carbon source) was evaluated because of the multiple reports describing successful expression in this system, including fragments of *botulinum* neurotoxin type A (Byrne et al., "Purification, Potency, and Efficacy of the *botulinum* Neurotoxin Type A Binding Domain from *Pichia pastoris* as a Recombinant Vaccine Candidate," *Infect. Immun.* 66:4817-4822 (1998), which is hereby incorporated by reference in its entirety). This system has the advantages of low cost, post-translational modification of the recombinant proteins typical for eukaryotes, and low amounts of naturally secreted products, which facilitate purification of the recombinant proteins. The *Pichia* expression system also can provide better yields than the baculovirus system. Disadvantages of the *Pichia* system include cumbersome procedures of cloning into the *Pichia* genome and selection of multiple-copy recombinants, the possibility of extensive glycosylation of some recombinant proteins, and the possibility of premature termination of RNA transcripts synthesized from AT-rich templates, a known characteristic of the Clostridial toxin genes. These unwanted internal DNA features in BoNT genes were eliminated at the cloning stage. The system was the first to be tested with BoNT A td to establish benchmarks for comparison to other expression systems.

## Prophetic Example 27

## Engineering of the Expression Constructs Targeted for Secretion

The coding part of the modified N-terminally 6-His tagged BoNT A td was subcloned into vector pPIC9K, which provides the alpha-factor secretion signal from *S. cerevisiae* in the expression plasmid. This should result in secretion of the expressed protein into the medium. The construct was linearized by restriction endonuclease Sal I and transfected by spheroplasting method into KM71 and GS115 strains of *Pichia pastoris*. Primary selection of the transformants was performed by testing ability of the cells to grow on histidine-deficient media, trait deficient in the wild-type cells. Second round of selection was performed in the presence of antibiotic Geneticin which allowed the identification of clones with multiple inserts of the gene of interest by the correlation

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between the number of the copies of the gene of interest and increased concentration of Geneticin in the growth medium. The ability of identified clones to express protein of interest will be tested by growing cells in the methanol-containing medium.

## Prophetic Example 28

## Lengthening of the His Affinity Tag

The length of the histidine affinity tag at the N-termini of the BoNT A td were increased, and two more constructs—8-His and 12-His tagged were tested for their ability to confer higher affinity for Ni-NTA agarose in the recombinant BoNT products. This approach has been used successfully for other recombinant proteins which showed a similar decreased affinity for Ni-NTA affinity purification media (Ichtchenko et al., "Alpha-Latrotoxin Action Probed with Recombinant Toxin: Receptors Recruit Alpha-Latrotoxin but do not Transduce an Exocytotic Signal," *EMBO J.* 17:6188-6199 (1998); Rudenko et al., "Structure of the LDL Receptor Extracellular Domain at Endosomal pH," *Science* 298:2353-2358 (2002), which are hereby incorporated by reference in their entirety). If improved purification of recombinant BoNT A td can be achieved, at least two steps of ion-exchange chromatography can be omitted from the current purification scheme.

## Example 29

## Engineering the Non-Expression Plasmid pLitBoNTAME224A Containing the Full-Length Sequence of BoNT A ad

The plasmid encoding full-length BoNT A ad cDNA with protease-inactivating mutation E224>A was created by the site-directed mutagenesis of the plasmid pLitBoNTA with phosphorylated oligonucleotides

```
CBA18: (SEQ ID NO: 46)
5' -pCCCGCGGTGACATTAGCACATGCACCTTATACATGCTGG
and
CBA19: (SEQ ID NO: 47)
5' -pCATGTGCTAATGTCCACCGGGATCTGTAGCAAAATTG
```

using GeneTailor™ Site-Directed Mutagenesis System (Invitrogen, Cat. #12397-014) and Platinum® Pfx DNA Polymerase (Invitrogen, Cat. #11708-021), according to the protocol supplied by the manufacturer. The size of pLitBoNTAME224A is 6712 b.p. with 3900 b.p. coding sequence.

## Example 30

## Engineering of the Non-Expression Plasmid pLitGFPBoNTAHC, Containing Full-Length Sequence of BoNT A gfpd

The plasmid pLitGFPBoNTAHC, encoding chimeric protein where minimal essential catalytic domain of the BoNT A light chain was substituted with the GFP was created by the following protocol: 742 b.p. PCR product, obtained on plasmid pEGFP-N3 (Clontech, Cat. #632313) with oligonucleotides



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using GeneTailor™ Site-Directed Mutagenesis System (Invitrogen, Cat. #12397-014) and Platinum® Pfx DNA Polymerase (Invitrogen, Cat. #11708-021), according to the protocol supplied by the manufacturer. The resulting plasmid pLitBoNTALCME224A is 4042 b.p. with a 1230 b.p. coding sequence.

## Chimera 1

The non-expression plasmid pLitBoNTACH1, containing the full-length sequence of BoNT A ad with three SNARE motif peptides substituting BoNT A light chain alpha-helix 1, was created by site-directed mutagenesis of the plasmid pLitBoNTAME224A with phosphorylated oligonucleotides

CBCH1: (SEQ ID NO: 52)  
5' -pGAGTTGTTGCGCTTGCTCATCCAACATCTGCAACGCGTCAGCTCGG  
TCATCCAACCTGTACTTAAATATGTTGAATCATAATATGAAACTGG  
and

CBCH2: (SEQ ID NO: 53)  
5' -pGAGCGCGAAATGGATGAAAACCTAGAGCAGGTGAGCGGCCGAGGAA  
TACCATTTTGGGGTGAAGTACAATAGATACAG

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit (Stratagene, Cat. #200502) with modification. The ExSite™ DNA polymerase blend included in the kit was substituted with a blend consisting of 75% of TaKaRa LA Taq DNA polymerase (Takara, Cat. #RR002A) and 25% of Platinum® Pfx polymerase (Invitrogen, Cat. #11708-021). The mutagenesis reaction and selection of the mutant plasmid were performed according to the protocol, included in the original Exsite™ PCR-Based Site-Directed Mutagenesis Kit. For selection purposes, two de novo endonuclease restriction sites—MluI and XhoI—were introduced into the plasmid.

## Chimera 2

The non-expression plasmid pLitBoNTACH2, containing the full-length sequence of BoNT A ad with two SNARE motif peptides substituting BoNT A light chain alpha-helix 4, was created by site-directed mutagenesis of the plasmid pLitBoNTAME224A with phosphorylated oligonucleotides

CBCH3: (SEQ ID NO: 54)  
5' -pCGCGTCTGCCCTATCGTCTAGTTTCATCTATAAACTTGCATCATGT  
CCCCC  
and

CBCH4: (SEQ ID NO: 55)  
5' -pTTACAAATGCTAGACGAACAGGGAGAGCAGCTCGAGAGGCTTAATA  
A AGCTAAATCAATAGTAGGTACTACTGC

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications, described above.

## Chimera 3

The non-expression plasmid pLitBoNTACH3, containing the full-length sequence of BoNT A ad with five SNARE motifs peptides substituting BoNT A light chain alpha-helices 1 and 4, was created by site-directed mutagenesis of the plasmid pLitBoNTACH1 with phosphorylated oligonucleotides

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CBCH3: (SEQ ID NO: 54)  
5' -pCGCGTCTGCCCTATCGTCTAGTTTCATCTATAAACTTGCATCATGT

5  
CC CCC  
and

CBCH4: (SEQ ID NO: 55)  
10 5' -pTTACAAATGCTAGACGAACAGGGAGAGCAGCTCGAGAGGC TTAAT  
AAAGCTAAATCAATAGTAGGTACTACTGC

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above.

## Chimera 4

The non-expression plasmid pLitBoNTACH4, containing the full-length sequence of BoNT A ad with three SNARE motif peptides substituting light chain alpha-helices 4 and 5, was created by site-directed mutagenesis of the plasmid pLitBoNTACH2 with phosphorylated oligonucleotides

25 CBCH5: (SEQ ID NO: 56)  
5' -pGCTTACTTGTTCCAAATTCTCGTCCATCTCTGAAGCAGTAGTACCT  
AC TATTGATTTAGC  
and

30 CBCH6: (SEQ ID NO: 57)  
5' -pGGCCGTCTCCTATCTGAAGATACATCTGG

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above. For the selection purposes, de novo endonuclease restriction site Eco52I was introduced into the plasmid.

## Chimera 5

40 The non-expression plasmid pLitBoNTACH5, containing the full-length sequence of BoNT A ad with six SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, and 5, was created by site-directed mutagenesis of the plasmid pLitBoNTACH2 with phosphorylated oligonucleotides

45 CBCH5: (SEQ ID NO: 56)  
5' -pGCTTACTTGTTCCAAATTCTCGTCCATCTCTGAAGCAGTAGTACCT  
50 AC TATTGATTTAGC  
and

CBCH6: (SEQ ID NO: 57)  
5' -pGGCCGTCTCCTATCTGAAGATACATCTGG

55 using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above. For the selection purposes, de novo endonuclease restriction site Eco52I was introduced into the plasmid.

## 60 Chimera 6

The non-expression plasmid pLitBoNTACH6, containing the full-length sequence of BoNT A ad with four SNARE motif peptides substituting BoNT A light chain alpha-helices 4, 5, and 6, was created by site-directed mutagenesis of the plasmid pLitBoNTACH4 with phosphorylated oligonucleotides



CBCH7 :  
 (SEQ ID NO: 58)  
 5' -pAATTCATCCATGAAATCTACCGAAAATTTTCC  
 and

CBCH8 :  
 (SEQ ID NO: 59)  
 5' -pCTTTGAACAGGTGGAGGAATTAACAGAGATTTACACAGAGG

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above. For the selection purposes, de novo endonuclease restriction site EcoRI was introduced into the plasmid.

#### Chimera 7

The non-expression plasmid pLitBoNTACH7, containing the full-length sequence of BoNT A ad with five SNARE motif peptides substituting BoNT A light chain alpha-helices 4, 5, 6, and 7, was created by site-directed mutagenesis of the plasmid pLitBoNTACH6 with phosphorylated oligonucleotides

CBCH9 :  
 (SEQ ID NO: 60)  
 5' -pTCGAGCTCTGTGTAATCTCTGTTAATTCC  
 and

CBCH10 :  
 (SEQ ID NO: 61)  
 5' -pGGACATGCTGGAGAGTGGGAATCTTAACAGAAAAACATATTTGAAT  
 TTTG

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above. For the selection purposes, de novo endonuclease restriction site XhoI was introduced into the plasmid.

#### Chimera 8

The non-expression plasmid pLitBoNTACH8, containing the full-length sequence of BoNT A ad with seven SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, 5, and 6, was created by site-directed mutagenesis of the plasmid pLitBoNTACH5 with phosphorylated oligonucleotides

CBCH7 :  
 (SEQ ID NO: 58)  
 5' -pAATTCATCCATGAAATCTACCGAAAATTTTCC  
 and

CBCH8 :  
 (SEQ ID NO: 59)  
 5' -pCTTTGAACAGGTGGAGGAATTAACAGAGATTTACACAGAGG

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above. For the selection purposes, de novo endonuclease restriction site EcoRI was introduced into the plasmid.

#### Chimera 9

The non-expression plasmid pLitBoNTACH9, containing the full-length sequence of BoNT A ad with eight SNARE motif peptides substituting BoNT

A light chain alpha-helices 1, 4, 5, 6, and 7, was created by site-directed mutagenesis of the plasmid pLitBoNTACH8 with phosphorylated oligonucleotides

CBCH9 :  
 (SEQ ID NO: 60)  
 5' -pTCGAGCTCTGTGTAATCTCTGTTAATTCC  
 and

-continued

CBCH10 :  
 (SEQ ID NO: 61)  
 5' -pGGACATGCTGGAGAGTGGGAATCTTAACAGAAAAACATATTTGAAT  
 TTTG

using Exsite™ PCR-Based Site-Directed Mutagenesis Kit with the modifications described above. For the selection purposes, de novo endonuclease restriction site XhoI was introduced into the plasmid.

#### Green Fluorescence Protein

The non-expression plasmid pLitEGFP, containing the full-length sequence of GFP for the fusions with BoNT A light chain, BoNT A light chain ad, and BoNT A light chain ad chimeric derivatives, was created by subcloning ~750 b.p. product obtained by PCR on plasmid pEGFP-N3 (Clontech, Cat. #6080-1) with oligonucleotides

EGFPs :  
 (SEQ ID NO: 62)  
 5' -TATTACGCGTGC GCGCTATGAATCTATAAGTTGCTAATGGTGAGCA

AGGGCGAGGAGCTGTTCACCGGG  
 and

EGFPa :  
 (SEQ ID NO: 63)  
 5' -ATTAGGGCCCTATTACTTGTACAGCTCGTCCATGCCGAGAGTGATC

CC

and digested with restriction endonucleases MluI and ApaI into vector pLitmus38i (NEB, Cat. #N3538S) pre-digested with MluI and ApaI and dephosphorylated. The size of the resulting pLitEGFP was 3479 b.p.

#### Light Chain of BoNT A td Fused with EGFP

The non-expression vector pLitBoNTALCEGFP carrying light chain of BoNT A td, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the digest of the plasmid pLitBoNTALC with restriction endonucleases BssHII and EcoRI into vector pLitEGFP pre-digested with BssHII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of BoNT A ad Fused with EGFP

The non-expression vector pLitBoNTAME224ALCEGFP carrying the sequence of the light chain of BoNT A ad, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the digest of the plasmid pLitBoNTAME224A with restriction endonucleases BssHII and EcoRI into vector pLitEGFP pre-digested with BssHII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 1 Fused with EGFP

The non-expression vector pLitBoNTACH1EGFP, carrying the sequence of the BoNT A ad light chain with three SNARE motif peptides substituting BoNT A light chain alpha-helix 1, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the digest of the plasmid pLitBoNTACH1, with restriction endonucleases BssHII and EcoRI into vector pLitEGFP pre-digested with BssHII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 2 Fused with EGFP

The non-expression vector pLitBoNTACH2EGFP, carrying the sequence of the BoNT A ad light chain with two SNARE motif peptides substituting BoNT A light chain alpha-helix 4, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the digest of the

plasmid pLitBoNTACH2 with restriction endonucleases BssHIII and EcoRI into vector pLitEGFP pre-digested with BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 3 Fused with EGFP

The non-expression vector pLitBoNTACH3EGFP, carrying the sequence of the BoNT A ad light chain with five SNARE motif peptides substituting BoNT A light chain alpha-helices 1 and 4, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the digest of the plasmid pLitBoNTACH3 with restriction endonucleases BssHIII and EcoRI into vector pLitEGFP pre-digested with BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 4 Fused with EGFP

The non-expression vector pLitBoNTACH4EGFP, carrying the sequence of the BoNT A ad light chain with three SNARE motif peptides substituting BoNT A light chain alpha-helices 4 and 5, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the digest of the plasmid pLitBoNTACH4 with restriction endonucleases BssHIII and EcoRI into vector pLitEGFP pre-digested with BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 5 Fused with EGFP

The non-expression vector pLitBoNTACH5EGFP, carrying the sequence of the BoNT A ad light chain with six SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, and 5, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment, obtained from the digest of the plasmid pLitBoNTACH5 with restriction endonucleases BssHIII and EcoRI into vector pLitEGFP pre-digested with BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 6 Fused with EGFP

The non-expression vector pLitBoNTACH6EGFP, carrying the sequence of the BoNT A ad light chain with four SNARE motif peptides substituting BoNT A light chain alpha-helices 4, 5, and 6, fused with EGFP, was created by subcloning 1296 b.p. DNA fragment, obtained from the incomplete digest with EcoRI of the 2019 b.p. DNA fragment, obtained from the plasmid pLitBoNTACH6, digested with restriction endonucleases BssHIII and AlwNI, into vector pLitEGFP pre-digested with BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 7 Fused with EGFP

The non-expression vector pLitBoNTACH7EGFP, carrying the sequence of the BoNT A ad light chain with five SNARE motif peptides substituting BoNT A light chain alpha-helices 4, 5, 6, and 7, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment, obtained from the incomplete digest with EcoRI of the 2019 b.p. DNA fragment, obtained from the plasmid pLitBoNTACH7 digested with restriction endonucleases BssHIII and AlwNI into vector pLitEGFP pre-digested with BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 8 Fused with EGFP

The non-expression vector pLitBoNTACH8EGFP, carrying the sequence of the BoNT A ad light chain with seven SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, 5, and 6, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the incomplete digest with EcoRI of the 1754 b.p. DNA fragment obtained from the plasmid pLitBoNTACH8 digested with restriction endonucleases BssHIII and HincII into vector pLitEGFP pre-digested with restriction endonucleases

BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Light Chain of Chimera 9 Fused with EGFP

The non-expression vector pLitBoNTACH9EGFP, carrying the sequence of the BoNT A ad light chain with eight SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, 5, 6, and 7, fused with EGFP, was created by subcloning the 1296 b.p. DNA fragment obtained from the incomplete digest with EcoRI of the 1754 b.p. DNA fragment obtained from the plasmid pLitBoNTACH9 digested with restriction endonucleases BssHIII and HincII, into vector pLitEGFP pre-digested with restriction endonucleases BssHIII and EcoRI and dephosphorylated. The size of the resulting plasmid was ~4800 b.p.

#### Sindbis Expression Vector—EGFP

The Sindbis expression vector pSinEGFP, carrying the EGFP sequence was created by subcloning the ~750 b.p. DNA fragment obtained from the plasmid pLitEGFP sequentially digested with restriction endonuclease EcoRI filled-in with Klenow fragment, and digested with restriction endonuclease ApaI into vector pSinRep5 (Invitrogen, Cat. #K750-1) pre-digested with restriction endonucleases StuI and ApaI and dephosphorylated. The size of the resulting plasmid was ~10,250 b.p.

#### Sindbis Expression Vector—BoNT A td Light Chain Fused with EGFP

The Sindbis expression vector pSinBoNTALCEGFP, carrying the sequence of BoNT A td light, chain fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTALCEGFP, digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

#### Sindbis Expression Vector—BoNT A ad Light Chain Fused with EGFP

The Sindbis expression vector pSinBoNTAME224ALCEGFP, carrying the sequence of BoNT A ad light, chain fused with EGFP was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTAME224ALCEGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

#### Sindbis Expression Vector—Light Chain of Chimera 1 Fused with EGFP

The Sindbis expression vector pSinBoNTACH1EGFP, carrying the sequence of BoNT A ad light chain with three SNARE motif peptides substituting BoNT A light chain alpha-helix 1, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH1EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

#### Sindbis Expression Vector—Light Chain of Chimera 2 Fused with EGFP

The Sindbis expression vector pSinBoNTACH2EGFP, carrying the sequence of BoNT A ad light chain with two SNARE motif peptides substituting BoNT A light chain alpha-helix 4, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH2EGFP, digested with restriction endonucleases NheI and ApaI into vector pSinRep5 (Invitrogen, Cat.

#K750-1) pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 3 Fused with EGFP

The Sindbis expression vector pSinBoNTACH3EGFP, carrying the sequence of BoNT A ad light chain with five SNARE motif peptides substituting BoNT A light chain alpha-helices 1 and 4, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH3EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 4 Fused with EGFP

The Sindbis expression vector pSinBoNTACH4EGFP, carrying the sequence of BoNT A ad light chain with three SNARE motif peptides substituting BoNT A light chain alpha-helices 4 and 5, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH4EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 5 Fused with EGFP

The Sindbis expression vector pSinBoNTACH5EGFP, carrying the sequence of BoNT A ad light chain with six SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, and 5, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH5EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 6 Fused with EGFP

The Sindbis expression vector pSinBoNTACH6EGFP, carrying the sequence of BoNT A ad light chain with four SNARE motif peptides substituting BoNT A light chain alpha-helices 4, 5, and 6, fused with EGFP, was created by subcloning ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH6EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 7 Fused with EGFP

The Sindbis expression vector pSinBoNTACH7EGFP, carrying the sequence of BoNT A ad light chain with five SNARE motif peptides substituting BoNT A light chain alpha-helices 4, 5, 6, and 7, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH7EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-di-

gested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 8 Fused with EGFP

The Sindbis expression vector pSinBoNTACH8EGFP, carrying the sequence of BoNT A ad light chain with seven SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, 5, and 6, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH8EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

Sindbis Expression Vector—Light Chain of Chimera 9 Fused with EGFP

The Sindbis expression vector pSinBoNTACH9EGFP, carrying the sequence of BoNT A ad light chain with eight SNARE motif peptides substituting BoNT A light chain alpha-helices 1, 4, 5, 6, and 7, fused with EGFP, was created by subcloning the ~2,050 b.p. DNA fragment obtained from the plasmid pLitBoNTACH9EGFP digested with restriction endonucleases NheI and ApaI into vector pSinRep5 pre-digested with restriction endonucleases XbaI and ApaI and dephosphorylated. The size of the resulting plasmid was ~11,550 b.p.

The Sindbis expression vectors were prepared for RNA synthesis. The plasmids pSinEGFP, pSinBoNTALCEGFP, pSinBoNTAME224ALCEGFP, pSinBoNTACH1EGFP, pSinBoNTACH2EGFP, pSinBoNTACH3EGFP, pSinBoNTACH4EGFP, pSinBoNTACH5EGFP, pSinBoNTACH6EGFP, pSinBoNTACH7EGFP, pSinBoNTACH8EGFP, and pSinBoNTACH9EGFP were linearized by the digest with restriction endonuclease NotI. The linearized plasmids were used for the RNA synthesis according to the protocol supplied with Sindbis expression system kit (Invitrogen, Cat. #K750-1).

DSGXXS Motif Library

To further mark the antagonist wild-type BoNT A light chain complex for elimination, a second library of light chain chimeras will be produced. This library will incorporate the DSGXXS (SEQ ID NO: 64) motif into the chimeras produced in the first library. The motif DSGXXS is present in a variety of cytosolic proteins and has been shown to target them for degradation via the proteasome pathway upon its phosphorylation (Cardozo et al., "The SCF Ubiquitin Ligase: Insights Into a Molecular Machine," *Nat. Rev. Mol. Cell Biol.* 5:739-751 (2004); Busino et al., "Degradation of Cdc25A by Beta-TrCP During S Phase and In Response to DNA Damage," *Nature* 426:87-91 (2003), which are hereby incorporated by reference in their entirety). The motif will be positioned to cause minimal interference with the 3D structure of the ancestral protein (i.e., wild-type BoNT A light chain).

Although the invention has been described in detail for the purposes of illustration, it is understood that such detail is solely for that purpose, and variations can be made therein by those skilled in the art without departing from the spirit and scope of the invention which is defined by the following claims.

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SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 66

<210> SEQ ID NO 1

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<400> SEQUENCE: 1

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 1                               5                               10          15
Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro
 20                               25                               30
Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg
 35                               40                               45
Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu
 50                               55                               60
Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr
 65                               70                               75          80
Asp Asn Glu Lys Asp Asn Tyr Leu Lys Gly Val Thr Lys Leu Phe Glu
 85                               90                               95
Arg Ile Tyr Ser Thr Asp Leu Gly Arg Met Leu Leu Thr Ser Ile Val
 100                              105                              110
Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys
 115                              120                              125
Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr
 130                              135                              140
Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile
 145                              150                              155          160
Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr
 165                              170                              175
Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe
 180                              185                              190
Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu
 195                              200                              205
Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Glu
 210                              215                              220
Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn
 225                              230                              235          240
Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu
 245                              250                              255
Glu Val Ser Phe Glu Glu Leu Arg Thr Phe Gly Gly His Asp Ala Lys
 260                              265                              270
Phe Ile Asp Ser Leu Gln Glu Asn Glu Phe Arg Leu Tyr Tyr Tyr Asn
 275                              280                              285
Lys Phe Lys Asp Ile Ala Ser Thr Leu Asn Lys Ala Lys Ser Ile Val
 290                              295                              300
Gly Thr Thr Ala Ser Leu Gln Tyr Met Lys Asn Val Phe Lys Glu Lys
 305                              310                              315          320
Tyr Leu Leu Ser Glu Asp Thr Ser Gly Lys Phe Ser Val Asp Lys Leu
 325                              330                              335
Lys Phe Asp Lys Leu Tyr Lys Met Leu Thr Glu Ile Thr Thr Glu Asp
 340                              345                              350
Asn Phe Val Lys Phe Phe Lys Val Leu Asn Arg Lys Thr Tyr Leu Asn
 355                              360                              365
Phe Asp Lys Ala Val Phe Lys Ile Asn Ile Val Pro Lys Val Asn Tyr
 370                              375                              380
Thr Ile Tyr Asp Gly Phe Asn Leu Arg Asn Thr Asn Leu Ala Ala Asn

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385	390	395	400
Phe Asn Gly Gln Asn Thr Glu Ile Asn Asn Met Asn Phe Thr Lys Leu	405	410	415
Lys Asn Phe Thr Gly Leu Phe Glu Phe Tyr Lys Leu Leu Cys Val Arg	420	425	430
Gly Ile Ile Thr Ser Lys Thr Lys Ser Leu Asp Lys Gly Tyr Asn Lys	435	440	445
Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn Trp Asp Leu Phe Phe	450	455	460
Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu Asn Lys Gly Glu Glu	465	470	475
Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu Glu Asn Ile Ser Leu	485	490	495
Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn Phe Asp Asn Glu Pro	500	505	510
Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp Ile Ile Gly Gln Leu	515	520	525
Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn Gly Lys Lys Tyr Glu	530	535	540
Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg Ala Gln Glu Phe Glu	545	550	555
His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser Val Asn Glu Ala Leu	565	570	575
Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser Ser Asp Tyr Val Lys	580	585	590
Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe Leu Gly Trp Val Glu	595	600	605
Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser Glu Val Ser Thr Thr	610	615	620
Asp Lys Ile Ala Asp Ile Thr Ile Ile Ile Pro Tyr Ile Gly Pro Ala	625	630	635
Leu Asn Ile Gly Asn Met Leu Tyr Lys Asp Asp Phe Val Gly Ala Leu	645	650	655
Ile Phe Ser Gly Ala Val Ile Leu Leu Glu Phe Ile Pro Glu Ile Ala	660	665	670
Ile Pro Val Leu Gly Thr Phe Ala Leu Val Ser Tyr Ile Ala Asn Lys	675	680	685
Val Leu Thr Val Gln Thr Ile Asp Asn Ala Leu Ser Lys Arg Asn Glu	690	695	700
Lys Trp Asp Glu Val Tyr Lys Tyr Ile Val Thr Asn Trp Leu Ala Lys	705	710	715
Val Asn Thr Gln Ile Asp Leu Ile Arg Lys Lys Met Lys Glu Ala Leu	725	730	735
Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile Asn Tyr Gln Tyr Asn	740	745	750
Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn Phe Asn Ile Asp Asp	755	760	765
Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys Ala Met Ile Asn Ile	770	775	780
Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr Leu Met Asn Ser Met	785	790	795
Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe Asp Ala Ser Leu Lys	805	810	815

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Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg Gly Thr Leu Ile Gly  
                   820                                  825                                  830

Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn Thr Leu Ser Thr Asp  
                   835                                  840                                  845

Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn Gln Arg Leu Leu Ser  
                   850                                  855                                  860

Thr Phe Thr Glu Tyr Ile Lys Asn Ile Ile Asn Thr Ser Ile Leu Asn  
                   865                                  870                                  875                                  880

Leu Arg Tyr Glu Ser Asn His Leu Ile Asp Leu Ser Arg Tyr Ala Ser  
                                   885                                  890                                  895

Lys Ile Asn Ile Gly Ser Lys Val Asn Phe Asp Pro Ile Asp Lys Asn  
                                   900                                  905                                  910

Gln Ile Gln Leu Phe Asn Leu Glu Ser Ser Lys Ile Glu Val Ile Leu  
                                   915                                  920                                  925

Lys Asn Ala Ile Val Tyr Asn Ser Met Tyr Glu Asn Phe Ser Thr Ser  
                                   930                                  935                                  940

Phe Trp Ile Arg Ile Pro Lys Tyr Phe Asn Ser Ile Ser Leu Asn Asn  
                                   945                                  950                                  955                                  960

Glu Tyr Thr Ile Ile Asn Cys Met Glu Asn Asn Ser Gly Trp Lys Val  
                                   965                                  970                                  975

Ser Leu Asn Tyr Gly Glu Ile Ile Trp Thr Leu Gln Asp Thr Gln Glu  
                                   980                                  985                                  990

Ile Lys Gln Arg Val Val Phe Lys Tyr Ser Gln Met Ile Asn Ile Ser  
                                   995                                  1000                                  1005

Asp Tyr Ile Asn Arg Trp Ile Phe Val Thr Ile Thr Asn Asn Arg  
                   1010                                  1015                                  1020

Leu Asn Asn Ser Lys Ile Tyr Ile Asn Gly Arg Leu Ile Asp Gln  
                   1025                                  1030                                  1035

Lys Pro Ile Ser Asn Leu Gly Asn Ile His Ala Ser Asn Asn Ile  
                   1040                                  1045                                  1050

Met Phe Lys Leu Asp Gly Cys Arg Asp Thr His Arg Tyr Ile Trp  
                   1055                                  1060                                  1065

Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu Leu Asn Glu Lys Glu  
                   1070                                  1075                                  1080

Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn Ser Gly Ile Leu Lys  
                   1085                                  1090                                  1095

Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp Lys Pro Tyr Tyr Met  
                   1100                                  1105                                  1110

Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val Asp Val Asn Asn Val  
                   1115                                  1120                                  1125

Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly Pro Arg Gly Ser Val  
                   1130                                  1135                                  1140

Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser Leu Tyr Arg Gly Thr  
                   1145                                  1150                                  1155

Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly Asn Lys Asp Asn Ile  
                   1160                                  1165                                  1170

Val Arg Asn Asn Asp Arg Val Tyr Ile Asn Val Val Val Lys Asn  
                   1175                                  1180                                  1185

Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser Gln Ala Gly Val Glu  
                   1190                                  1195                                  1200

Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp Val Gly Asn Leu Ser  
                   1205                                  1210                                  1215

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Gln Val Val Val Met Lys Ser Lys Asn Asp Gln Gly Ile Thr Asn  
1220 1225 1230

Lys Cys Lys Met Asn Leu Gln Asp Asn Asn Gly Asn Asp Ile Gly  
1235 1240 1245

Phe Ile Gly Phe His Gln Phe Asn Asn Ile Ala Lys Leu Val Ala  
1250 1255 1260

Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser Ser Arg Thr Leu  
1265 1270 1275

Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp Gly Trp Gly Glu  
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Arg Pro Leu  
1295

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20 25 30

Tyr Tyr Lys Ala Phe Lys Ile Thr Asp Arg Ile Trp Ile Ile Pro Glu  
35 40 45

Arg Tyr Thr Phe Gly Tyr Lys Pro Glu Asp Phe Asn Lys Ser Ser Gly  
50 55 60

Ile Phe Asn Arg Asp Val Cys Glu Tyr Tyr Asp Pro Asp Tyr Leu Asn  
65 70 75 80

Thr Asn Asp Lys Lys Asn Ile Phe Leu Gln Thr Met Ile Lys Leu Phe  
85 90 95

Asn Arg Ile Lys Ser Lys Pro Leu Gly Glu Lys Leu Leu Glu Met Ile  
100 105 110

Ile Asn Gly Ile Pro Tyr Leu Gly Asp Arg Arg Val Pro Leu Glu Glu  
115 120 125

Phe Asn Thr Asn Ile Ala Ser Val Thr Val Asn Lys Leu Ile Ser Asn  
130 135 140

Pro Gly Glu Val Glu Arg Lys Lys Gly Ile Phe Ala Asn Leu Ile Ile  
145 150 155 160

Phe Gly Pro Gly Pro Val Leu Asn Glu Asn Glu Thr Ile Asp Ile Gly  
165 170 175

Ile Gln Asn His Phe Ala Ser Arg Glu Gly Phe Gly Gly Ile Met Gln  
180 185 190

Met Lys Phe Cys Pro Glu Tyr Val Ser Val Phe Asn Asn Val Gln Glu  
195 200 205

Asn Lys Gly Ala Ser Ile Phe Asn Arg Arg Gly Tyr Phe Ser Asp Pro  
210 215 220

Ala Leu Ile Leu Met His Glu Leu Ile His Val Leu His Gly Leu Tyr  
225 230 235 240

Gly Ile Lys Val Asp Asp Leu Pro Ile Val Pro Asn Glu Lys Lys Phe  
245 250 255

Phe Met Gln Ser Thr Asp Ala Ile Gln Ala Glu Glu Leu Tyr Thr Phe  
260 265 270

Gly Gly Gln Asp Pro Ser Ile Ile Thr Pro Ser Thr Asp Lys Ser Ile  
275 280 285

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Tyr Asp Lys Val Leu Gln Asn Phe Arg Gly Ile Val Asp Arg Leu Asn  
 290 295 300  
 Lys Val Leu Val Cys Ile Ser Asp Pro Asn Ile Asn Ile Asn Ile Tyr  
 305 310 315 320  
 Lys Asn Lys Phe Lys Asp Lys Tyr Lys Phe Val Glu Asp Ser Glu Gly  
 325 330 335  
 Lys Tyr Ser Ile Asp Val Glu Ser Phe Asp Lys Leu Tyr Lys Ser Leu  
 340 345 350  
 Met Phe Gly Phe Thr Glu Thr Asn Ile Ala Glu Asn Tyr Lys Ile Lys  
 355 360 365  
 Thr Arg Ala Ser Tyr Phe Ser Asp Ser Leu Pro Pro Val Lys Ile Lys  
 370 375 380  
 Asn Leu Leu Asp Asn Glu Ile Tyr Thr Ile Glu Glu Gly Phe Asn Ile  
 385 390 395 400  
 Ser Asp Lys Asp Met Glu Lys Glu Tyr Arg Gly Gln Asn Lys Ala Ile  
 405 410 415  
 Asn Lys Gln Ala Tyr Glu Glu Ile Ser Lys Glu His Leu Ala Val Tyr  
 420 425 430  
 Lys Ile Gln Met Cys Lys Ser Val Lys Ala Pro Gly Ile Cys Ile Asp  
 435 440 445  
 Val Asp Asn Glu Asp Leu Phe Phe Ile Ala Asp Lys Asn Ser Phe Ser  
 450 455 460  
 Asp Asp Leu Ser Lys Asn Glu Arg Ile Glu Tyr Asn Thr Gln Ser Asn  
 465 470 475 480  
 Tyr Ile Glu Asn Asp Phe Pro Ile Asn Glu Leu Ile Leu Asp Thr Asp  
 485 490 495  
 Leu Ile Ser Lys Ile Glu Leu Pro Ser Glu Asn Thr Glu Ser Leu Thr  
 500 505 510  
 Asp Phe Asn Val Asp Val Pro Val Tyr Glu Lys Gln Pro Ala Ile Lys  
 515 520 525  
 Lys Ile Phe Thr Asp Glu Asn Thr Ile Phe Gln Tyr Leu Tyr Ser Gln  
 530 535 540  
 Thr Phe Pro Leu Asp Ile Arg Asp Ile Ser Leu Thr Ser Ser Phe Asp  
 545 550 555 560  
 Asp Ala Leu Leu Phe Ser Asn Lys Val Tyr Ser Phe Phe Ser Met Asp  
 565 570 575  
 Tyr Ile Lys Thr Ala Asn Lys Val Val Glu Ala Gly Leu Phe Ala Gly  
 580 585 590  
 Trp Val Lys Gln Ile Val Asn Asp Phe Val Ile Glu Ala Asn Lys Ser  
 595 600 605  
 Asn Thr Met Asp Lys Ile Ala Asp Ile Ser Leu Ile Val Pro Tyr Ile  
 610 615 620  
 Gly Leu Ala Leu Asn Val Gly Asn Glu Thr Ala Lys Gly Asn Phe Glu  
 625 630 635 640  
 Asn Ala Phe Glu Ile Ala Gly Ala Ser Ile Leu Leu Glu Phe Ile Pro  
 645 650 655  
 Glu Leu Leu Ile Pro Val Val Gly Ala Phe Leu Leu Glu Ser Tyr Ile  
 660 665 670  
 Asp Asn Lys Asn Lys Ile Ile Lys Thr Ile Asp Asn Ala Leu Thr Lys  
 675 680 685  
 Arg Asn Glu Lys Trp Ser Asp Met Tyr Gly Leu Ile Val Ala Gln Trp  
 690 695 700



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Leu Ser Thr Val Asn Thr Gln Phe Tyr Thr Ile Lys Glu Gly Met Tyr  
 705 710 715 720  
 Lys Ala Leu Asn Tyr Gln Ala Gln Ala Leu Lys Glu Ile Ile Lys Tyr  
 725 730 735  
 Arg Tyr Asn Ile Tyr Ser Glu Lys Glu Lys Ser Asn Ile Asn Ile Asp  
 740 745 750  
 Phe Asn Asp Ile Asn Ser Lys Leu Asn Glu Gly Ile Asn Gln Ala Ile  
 755 760 765  
 Asp Asn Ile Asn Asn Phe Ile Asn Gly Cys Ser Val Ser Tyr Leu Met  
 770 775 780  
 Lys Lys Met Ile Pro Leu Ala Val Glu Lys Leu Leu Asp Phe Asp Asn  
 785 790 795 800  
 Thr Leu Lys Lys Asn Leu Leu Asn Tyr Ile Asp Glu Asn Lys Leu Tyr  
 805 810 815  
 Leu Ile Gly Ser Ala Glu Tyr Glu Lys Ser Lys Val Asn Lys Tyr Leu  
 820 825 830  
 Lys Thr Ile Met Pro Phe Asp Leu Ser Ile Tyr Thr Asn Asp Thr Ile  
 835 840 845  
 Leu Ile Glu Met Phe Asn Lys Tyr Asn Ser Glu Ile Leu Asn Asn Ile  
 850 855 860  
 Ile Leu Asn Leu Arg Tyr Lys Asp Asn Asn Leu Ile Asp Leu Ser Gly  
 865 870 875 880  
 Tyr Gly Ala Lys Val Glu Val Tyr Asp Gly Val Glu Leu Asn Asp Lys  
 885 890 895  
 Asn Gln Phe Lys Leu Thr Ser Ser Ala Asn Ser Lys Ile Arg Val Thr  
 900 905 910  
 Gln Asn Gln Asn Ile Ile Phe Asn Ser Val Phe Leu Asp Phe Ser Val  
 915 920 925  
 Ser Phe Trp Ile Arg Ile Pro Lys Tyr Lys Asn Asp Gly Ile Gln Asn  
 930 935 940  
 Tyr Ile His Asn Glu Tyr Thr Ile Ile Asn Cys Met Lys Asn Asn Ser  
 945 950 955 960  
 Gly Trp Lys Ile Ser Ile Arg Gly Asn Arg Ile Ile Trp Thr Leu Ile  
 965 970 975  
 Asp Ile Asn Gly Lys Thr Lys Ser Val Phe Phe Glu Tyr Asn Ile Arg  
 980 985 990  
 Glu Asp Ile Ser Glu Tyr Ile Asn Arg Trp Phe Phe Val Thr Ile Thr  
 995 1000 1005  
 Asn Asn Leu Asn Asn Ala Lys Ile Tyr Ile Asn Gly Lys Leu Glu  
 1010 1015 1020  
 Ser Asn Thr Asp Ile Lys Asp Ile Arg Glu Val Ile Ala Asn Gly  
 1025 1030 1035  
 Glu Ile Ile Phe Lys Leu Asp Gly Asp Ile Asp Arg Thr Gln Phe  
 1040 1045 1050  
 Ile Trp Met Lys Tyr Phe Ser Ile Phe Asn Thr Glu Leu Ser Gln  
 1055 1060 1065  
 Ser Asn Ile Glu Glu Arg Tyr Lys Ile Gln Ser Tyr Ser Glu Tyr  
 1070 1075 1080  
 Leu Lys Asp Phe Trp Gly Asn Pro Leu Met Tyr Asn Lys Glu Tyr  
 1085 1090 1095  
 Tyr Met Phe Asn Ala Gly Asn Lys Asn Ser Tyr Ile Lys Leu Lys  
 1100 1105 1110  
 Lys Asp Ser Pro Val Gly Glu Ile Leu Thr Arg Ser Lys Tyr Asn

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1115	1120	1125
Gln Asn Ser Lys Tyr Ile	Asn Tyr Arg Asp Leu Tyr	Ile Gly Glu
1130	1135	1140
Lys Phe Ile Ile Arg Arg	Lys Ser Asn Ser Gln Ser	Ile Asn Asp
1145	1150	1155
Asp Ile Val Arg Lys Glu	Asp Tyr Ile Tyr Leu Asp	Phe Phe Asn
1160	1165	1170
Leu Asn Gln Glu Trp Arg	Val Tyr Thr Tyr Lys Tyr	Phe Lys Lys
1175	1180	1185
Glu Glu Glu Lys Leu Phe	Leu Ala Pro Ile Ser Asp	Ser Asp Glu
1190	1195	1200
Phe Tyr Asn Thr Ile Gln	Ile Lys Glu Tyr Asp Glu	Gln Pro Thr
1205	1210	1215
Tyr Ser Cys Gln Leu Leu	Phe Lys Lys Asp Glu Glu	Ser Thr Asp
1220	1225	1230
Glu Ile Gly Leu Ile Gly	Ile His Arg Phe Tyr Glu	Ser Gly Ile
1235	1240	1245
Val Phe Glu Glu Tyr Lys	Asp Tyr Phe Cys Ile Ser	Lys Trp Tyr
1250	1255	1260
Leu Lys Glu Val Lys Arg	Lys Pro Tyr Asn Leu Lys	Leu Gly Cys
1265	1270	1275
Asn Trp Gln Phe Ile Pro	Lys Asp Glu Gly Trp Thr	Glu
1280	1285	1290

&lt;210&gt; SEQ ID NO 3

&lt;211&gt; LENGTH: 1291

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Clostridium botulinum (serotype C)

&lt;400&gt; SEQUENCE: 3

Met Pro Ile Thr Ile Asn Asn Phe Asn Tyr Ser Asp Pro Val Asp Asn	1	5	10	15
Lys Asn Ile Leu Tyr Leu Asp Thr His Leu Asn Thr Leu Ala Asn Glu	20	25	30	
Pro Glu Lys Ala Phe Arg Ile Thr Gly Asn Ile Trp Val Ile Pro Asp	35	40	45	
Arg Phe Ser Arg Asn Ser Asn Pro Asn Leu Asn Lys Pro Pro Arg Val	50	55	60	
Thr Ser Pro Lys Ser Gly Tyr Tyr Asp Pro Asn Tyr Leu Ser Thr Asp	65	70	75	80
Ser Asp Lys Asp Pro Phe Leu Lys Glu Ile Ile Lys Leu Phe Lys Arg	85	90	95	
Ile Asn Ser Arg Glu Ile Gly Glu Glu Leu Ile Tyr Arg Leu Ser Thr	100	105	110	
Asp Ile Pro Phe Pro Gly Asn Asn Asn Thr Pro Ile Asn Thr Phe Asp	115	120	125	
Phe Asp Val Asp Phe Asn Ser Val Asp Val Lys Thr Arg Gln Gly Asn	130	135	140	
Asn Trp Val Lys Thr Gly Ser Ile Asn Pro Ser Val Ile Ile Thr Gly	145	150	155	160
Pro Arg Glu Asn Ile Ile Asp Pro Glu Thr Ser Thr Phe Lys Leu Thr	165	170	175	
Asn Asn Thr Phe Ala Ala Gln Glu Gly Phe Gly Ala Leu Ser Ile Ile	180	185	190	

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Ser Ile Ser Pro Arg Phe Met Leu Thr Tyr Ser Asn Ala Thr Asn Asp  
 195 200 205  
 Val Gly Glu Gly Arg Phe Ser Lys Ser Glu Phe Cys Met Asp Pro Ile  
 210 215 220  
 Leu Ile Leu Met His Glu Leu Asn His Ala Met His Asn Leu Tyr Gly  
 225 230 235 240  
 Ile Ala Ile Pro Asn Asp Gln Thr Ile Ser Ser Val Thr Ser Asn Ile  
 245 250 255  
 Phe Tyr Ser Gln Tyr Asn Val Lys Leu Glu Tyr Ala Glu Ile Tyr Ala  
 260 265 270  
 Phe Gly Gly Pro Thr Ile Asp Leu Ile Pro Lys Ser Ala Arg Lys Tyr  
 275 280 285  
 Phe Glu Glu Lys Ala Leu Asp Tyr Tyr Arg Ser Ile Ala Lys Arg Leu  
 290 295 300  
 Asn Ser Ile Thr Thr Ala Asn Pro Ser Ser Phe Asn Lys Tyr Ile Gly  
 305 310 315 320  
 Glu Tyr Lys Gln Lys Leu Ile Arg Lys Tyr Arg Phe Val Val Glu Ser  
 325 330 335  
 Ser Gly Glu Val Thr Val Asn Arg Asn Lys Phe Val Glu Leu Tyr Asn  
 340 345 350  
 Glu Leu Thr Gln Ile Phe Thr Glu Phe Asn Tyr Ala Lys Ile Tyr Asn  
 355 360 365  
 Val Gln Asn Arg Lys Ile Tyr Leu Ser Asn Val Tyr Thr Pro Val Thr  
 370 375 380  
 Ala Asn Ile Leu Asp Asp Asn Val Tyr Asp Ile Gln Asn Gly Phe Asn  
 385 390 395 400  
 Ile Pro Lys Ser Asn Leu Asn Val Leu Phe Met Gly Gln Asn Leu Ser  
 405 410 415  
 Arg Asn Pro Ala Leu Arg Lys Val Asn Pro Glu Asn Met Leu Tyr Leu  
 420 425 430  
 Phe Thr Lys Phe Cys His Lys Ala Ile Asp Gly Arg Ser Leu Tyr Asn  
 435 440 445  
 Lys Thr Leu Asp Cys Arg Glu Leu Leu Val Lys Asn Thr Asp Leu Pro  
 450 455 460  
 Phe Ile Gly Asp Ile Ser Asp Val Lys Thr Asp Ile Phe Leu Arg Lys  
 465 470 475 480  
 Asp Ile Asn Glu Glu Thr Glu Val Ile Tyr Tyr Pro Asp Asn Val Ser  
 485 490 495  
 Val Asp Gln Val Ile Leu Ser Lys Asn Thr Ser Glu His Gly Gln Leu  
 500 505 510  
 Asp Leu Leu Tyr Pro Ser Ile Asp Ser Glu Ser Glu Ile Leu Pro Gly  
 515 520 525  
 Glu Asn Gln Val Phe Tyr Asp Asn Arg Thr Gln Asn Val Asp Tyr Leu  
 530 535 540  
 Asn Ser Tyr Tyr Tyr Leu Glu Ser Gln Lys Leu Ser Asp Asn Val Glu  
 545 550 555 560  
 Asp Phe Thr Phe Thr Arg Ser Ile Glu Glu Ala Leu Asp Asn Ser Ala  
 565 570 575  
 Lys Val Tyr Thr Tyr Phe Pro Thr Leu Ala Asn Lys Val Asn Ala Gly  
 580 585 590  
 Val Gln Gly Gly Leu Phe Leu Met Trp Ala Asn Asp Val Val Glu Asp  
 595 600 605  
 Phe Thr Thr Asn Ile Leu Arg Lys Asp Thr Leu Asp Lys Ile Ser Asp

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610					615					620					
Val	Ser	Ala	Ile	Ile	Pro	Tyr	Ile	Gly	Pro	Ala	Leu	Asn	Ile	Ser	Asn
625					630					635					640
Ser	Val	Arg	Arg	Gly	Asn	Phe	Thr	Glu	Ala	Phe	Ala	Val	Thr	Gly	Val
				645					650					655	
Thr	Ile	Leu	Leu	Glu	Ala	Phe	Pro	Glu	Phe	Thr	Ile	Pro	Ala	Leu	Gly
			660					665					670		
Ala	Phe	Val	Ile	Tyr	Ser	Lys	Val	Gln	Glu	Arg	Asn	Glu	Ile	Ile	Lys
		675					680					685			
Thr	Ile	Asp	Asn	Cys	Leu	Glu	Gln	Arg	Ile	Lys	Arg	Trp	Lys	Asp	Ser
		690				695					700				
Tyr	Glu	Trp	Met	Met	Gly	Thr	Trp	Leu	Ser	Arg	Ile	Ile	Thr	Gln	Phe
705					710					715					720
Asn	Asn	Ile	Ser	Tyr	Gln	Met	Tyr	Asp	Ser	Leu	Asn	Tyr	Gln	Ala	Gly
				725					730					735	
Ala	Ile	Lys	Ala	Lys	Ile	Asp	Leu	Glu	Tyr	Lys	Lys	Tyr	Ser	Gly	Ser
			740					745						750	
Asp	Lys	Glu	Asn	Ile	Lys	Ser	Gln	Val	Glu	Asn	Leu	Lys	Asn	Ser	Leu
		755					760					765			
Asp	Val	Lys	Ile	Ser	Glu	Ala	Met	Asn	Asn	Ile	Asn	Lys	Phe	Ile	Arg
		770				775					780				
Glu	Cys	Ser	Val	Thr	Tyr	Leu	Phe	Lys	Asn	Met	Leu	Pro	Lys	Val	Ile
785					790					795					800
Asp	Glu	Leu	Asn	Glu	Phe	Asp	Arg	Asn	Thr	Lys	Ala	Lys	Leu	Ile	Asn
				805					810					815	
Leu	Ile	Asp	Ser	His	Asn	Ile	Ile	Leu	Val	Gly	Glu	Val	Asp	Lys	Leu
			820					825						830	
Lys	Ala	Lys	Val	Asn	Asn	Ser	Phe	Gln	Asn	Thr	Ile	Pro	Phe	Asn	Ile
			835				840					845			
Phe	Ser	Tyr	Thr	Asn	Asn	Ser	Leu	Leu	Lys	Asp	Ile	Ile	Asn	Glu	Tyr
		850				855					860				
Phe	Asn	Asn	Ile	Asn	Asp	Ser	Lys	Ile	Leu	Ser	Leu	Gln	Asn	Arg	Lys
865					870					875					880
Asn	Thr	Leu	Val	Asp	Thr	Ser	Gly	Tyr	Asn	Ala	Glu	Val	Ser	Glu	Glu
				885					890					895	
Gly	Asp	Val	Gln	Leu	Asn	Pro	Ile	Phe	Pro	Phe	Asp	Phe	Lys	Leu	Gly
			900					905						910	
Ser	Ser	Gly	Glu	Asp	Arg	Gly	Lys	Val	Ile	Val	Thr	Gln	Asn	Glu	Asn
		915					920					925			
Ile	Val	Tyr	Asn	Ser	Met	Tyr	Glu	Ser	Phe	Ser	Ile	Ser	Phe	Trp	Ile
		930				935					940				
Arg	Ile	Asn	Lys	Trp	Val	Ser	Asn	Leu	Pro	Gly	Tyr	Thr	Ile	Ile	Asp
945					950					955					960
Ser	Val	Lys	Asn	Asn	Ser	Gly	Trp	Ser	Ile	Gly	Ile	Ile	Ser	Asn	Phe
				965					970					975	
Leu	Val	Phe	Thr	Leu	Lys	Gln	Asn	Glu	Asp	Ser	Glu	Gln	Ser	Ile	Asn
			980					985						990	
Phe	Ser	Tyr	Asp	Ile	Ser	Asn	Asn	Ala	Pro	Gly	Tyr	Asn	Lys	Trp	Phe
			995				1000						1005		
Phe	Val	Thr	Val	Thr	Asn	Asn	Met	Met	Gly	Asn	Met	Lys	Ile	Tyr	
			1010				1015						1020		
Ile	Asn	Gly	Lys	Leu	Ile	Asp	Thr	Ile	Lys	Val	Lys	Glu	Leu	Thr	
			1025				1030						1035		

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Gly Ile Asn Phe Ser Lys Thr Ile Thr Phe Glu Ile Asn Lys Ile  
 1040 1045 1050  
 Pro Asp Thr Gly Leu Ile Thr Ser Asp Ser Asp Asn Ile Asn Met  
 1055 1060 1065  
 Trp Ile Arg Asp Phe Tyr Ile Phe Ala Lys Glu Leu Asp Gly Lys  
 1070 1075 1080  
 Asp Ile Asn Ile Leu Phe Asn Ser Leu Gln Tyr Thr Asn Val Val  
 1085 1090 1095  
 Lys Asp Tyr Trp Gly Asn Asp Leu Arg Tyr Asn Lys Glu Tyr Tyr  
 1100 1105 1110  
 Met Val Asn Ile Asp Tyr Leu Asn Arg Tyr Met Tyr Ala Asn Ser  
 1115 1120 1125  
 Arg Gln Ile Val Phe Asn Thr Arg Arg Asn Asn Asn Asp Phe Asn  
 1130 1135 1140  
 Glu Gly Tyr Lys Ile Ile Ile Lys Arg Ile Arg Gly Asn Thr Asn  
 1145 1150 1155  
 Asp Thr Arg Val Arg Gly Gly Asp Ile Leu Tyr Phe Asp Met Thr  
 1160 1165 1170  
 Ile Asn Asn Lys Ala Tyr Asn Leu Phe Met Lys Asn Glu Thr Met  
 1175 1180 1185  
 Tyr Ala Asp Asn His Ser Thr Glu Asp Ile Tyr Ala Ile Gly Leu  
 1190 1195 1200  
 Arg Glu Gln Thr Lys Asp Ile Asn Asp Asn Ile Ile Phe Gln Ile  
 1205 1210 1215  
 Gln Pro Met Asn Asn Thr Tyr Tyr Tyr Ala Ser Gln Ile Phe Lys  
 1220 1225 1230  
 Ser Asn Phe Asn Gly Glu Asn Ile Ser Gly Ile Cys Ser Ile Gly  
 1235 1240 1245  
 Thr Tyr Arg Phe Arg Leu Gly Gly Asp Trp Tyr Arg His Asn Tyr  
 1250 1255 1260  
 Leu Val Pro Thr Val Lys Gln Gly Asn Tyr Ala Ser Leu Leu Glu  
 1265 1270 1275  
 Ser Thr Ser Thr His Trp Gly Phe Val Pro Val Ser Glu  
 1280 1285 1290

&lt;210&gt; SEQ ID NO 4

&lt;211&gt; LENGTH: 1276

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Clostridium botulinum (serotype D)

&lt;400&gt; SEQUENCE: 4

Met Thr Trp Pro Val Lys Asp Phe Asn Tyr Ser Asp Pro Val Asn Asp  
 1 5 10 15  
 Asn Asp Ile Leu Tyr Leu Arg Ile Pro Gln Asn Lys Leu Ile Thr Thr  
 20 25 30  
 Pro Val Lys Ala Phe Met Ile Thr Gln Asn Ile Trp Val Ile Pro Glu  
 35 40 45  
 Arg Phe Ser Ser Asp Thr Asn Pro Ser Leu Ser Lys Pro Pro Arg Pro  
 50 55 60  
 Thr Ser Lys Tyr Gln Ser Tyr Tyr Asp Pro Ser Tyr Leu Ser Thr Asp  
 65 70 75 80  
 Glu Gln Lys Asp Thr Phe Leu Lys Gly Ile Ile Lys Leu Phe Lys Arg  
 85 90 95  
 Ile Asn Glu Arg Asp Ile Gly Lys Lys Leu Ile Asn Tyr Leu Val Val

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100					105					110					
Gly	Ser	Pro	Phe	Met	Gly	Asp	Ser	Ser	Thr	Pro	Glu	Asp	Thr	Phe	Asp
		115					120					125			
Phe	Thr	Arg	His	Thr	Thr	Asn	Ile	Ala	Val	Glu	Lys	Phe	Glu	Asn	Gly
	130					135					140				
Ser	Trp	Lys	Val	Thr	Asn	Ile	Ile	Thr	Pro	Ser	Val	Leu	Ile	Phe	Gly
	145					150					155				160
Pro	Leu	Pro	Asn	Ile	Leu	Asp	Tyr	Thr	Ala	Ser	Leu	Thr	Leu	Gln	Gly
			165						170					175	
Gln	Gln	Ser	Asn	Pro	Ser	Phe	Glu	Gly	Phe	Gly	Thr	Leu	Ser	Ile	Leu
		180						185						190	
Lys	Val	Ala	Pro	Glu	Phe	Leu	Leu	Thr	Phe	Ser	Asp	Val	Thr	Ser	Asn
		195					200					205			
Gln	Ser	Ser	Ala	Val	Leu	Gly	Lys	Ser	Ile	Phe	Cys	Met	Asp	Pro	Val
	210					215					220				
Ile	Ala	Leu	Met	His	Glu	Leu	Thr	His	Ser	Leu	His	Gln	Leu	Tyr	Gly
	225					230					235				240
Ile	Asn	Ile	Pro	Ser	Asp	Lys	Arg	Ile	Arg	Pro	Gln	Val	Ser	Glu	Gly
			245						250					255	
Phe	Phe	Ser	Gln	Asp	Gly	Pro	Asn	Val	Gln	Phe	Glu	Glu	Leu	Tyr	Thr
		260					265							270	
Phe	Gly	Gly	Leu	Asp	Val	Glu	Ile	Ile	Pro	Gln	Ile	Glu	Arg	Ser	Gln
		275					280					285			
Leu	Arg	Glu	Lys	Ala	Leu	Gly	His	Tyr	Lys	Asp	Ile	Ala	Lys	Arg	Leu
	290					295					300				
Asn	Asn	Ile	Asn	Lys	Thr	Ile	Pro	Ser	Ser	Trp	Ile	Ser	Asn	Ile	Asp
	305					310					315				320
Lys	Tyr	Lys	Lys	Ile	Phe	Ser	Glu	Lys	Tyr	Asn	Phe	Asp	Lys	Asp	Asn
			325						330					335	
Thr	Gly	Asn	Phe	Val	Val	Asn	Ile	Asp	Lys	Phe	Asn	Ser	Leu	Tyr	Ser
		340					345							350	
Asp	Leu	Thr	Asn	Val	Met	Ser	Glu	Val	Val	Tyr	Ser	Ser	Gln	Tyr	Asn
		355					360					365			
Val	Lys	Asn	Arg	Thr	His	Tyr	Phe	Ser	Arg	His	Tyr	Leu	Pro	Val	Phe
	370					375					380				
Ala	Asn	Ile	Leu	Asp	Asp	Asn	Ile	Tyr	Thr	Ile	Arg	Asp	Gly	Phe	Asn
	385					390					395				400
Leu	Thr	Asn	Lys	Gly	Phe	Asn	Ile	Glu	Asn	Ser	Gly	Gln	Asn	Ile	Glu
			405						410					415	
Arg	Asn	Pro	Ala	Leu	Gln	Lys	Leu	Ser	Ser	Glu	Ser	Val	Val	Asp	Leu
		420					425							430	
Phe	Thr	Lys	Val	Cys	Leu	Arg	Leu	Thr	Lys	Asn	Ser	Arg	Asp	Asp	Ser
		435					440					445			
Thr	Cys	Ile	Lys	Val	Lys	Asn	Asn	Arg	Leu	Pro	Tyr	Val	Ala	Asp	Lys
	450					455					460				
Asp	Ser	Ile	Ser	Gln	Glu	Ile	Phe	Glu	Asn	Lys	Ile	Ile	Thr	Asp	Glu
	465					470					475				480
Thr	Asn	Val	Gln	Asn	Tyr	Ser	Asp	Asn	Phe	Ser	Leu	Asp	Glu	Ser	Ile
			485						490					495	
Leu	Asp	Gly	Gln	Val	Pro	Ile	Asn	Pro	Glu	Ile	Val	Asp	Pro	Leu	Leu
			500				505						510		
Pro	Asn	Val	Asn	Met	Glu	Pro	Leu	Asn	Leu	Pro	Gly	Glu	Glu	Ile	Val
		515					520					525			

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Phe Tyr Asp Asp Ile Thr Lys Tyr Val Asp Tyr Leu Asn Ser Tyr Tyr  
 530 535 540  
 Tyr Leu Glu Ser Gln Lys Leu Ser Asn Asn Val Glu Asn Ile Thr Leu  
 545 550 555 560  
 Thr Thr Ser Val Glu Glu Ala Leu Gly Tyr Ser Asn Lys Ile Tyr Thr  
 565 570 575  
 Phe Leu Pro Ser Leu Ala Glu Lys Val Asn Lys Gly Val Gln Ala Gly  
 580 585 590  
 Leu Phe Leu Asn Trp Ala Asn Glu Val Val Glu Asp Phe Thr Thr Asn  
 595 600 605  
 Ile Met Lys Lys Asp Thr Leu Asp Lys Ile Ser Asp Val Ser Val Ile  
 610 615 620  
 Ile Pro Tyr Ile Gly Pro Ala Leu Asn Ile Gly Asn Ser Ala Leu Arg  
 625 630 635 640  
 Gly Asn Phe Lys Gln Ala Phe Ala Thr Ala Gly Val Ala Phe Leu Leu  
 645 650 655  
 Glu Gly Phe Pro Glu Phe Thr Ile Pro Ala Leu Gly Val Phe Thr Phe  
 660 665 670  
 Tyr Ser Ser Ile Gln Glu Arg Glu Lys Ile Ile Lys Thr Ile Glu Asn  
 675 680 685  
 Cys Leu Glu Gln Arg Val Lys Arg Trp Lys Asp Ser Tyr Gln Trp Met  
 690 695 700  
 Val Ser Asn Trp Leu Ser Arg Ile Thr Thr Gln Phe Asn His Ile Asn  
 705 710 715 720  
 Tyr Gln Met Tyr Asp Ser Leu Ser Tyr Gln Ala Asp Ala Ile Lys Ala  
 725 730 735  
 Lys Ile Asp Leu Glu Tyr Lys Lys Tyr Ser Gly Ser Asp Lys Glu Asn  
 740 745 750  
 Ile Lys Ser Gln Val Glu Asn Leu Lys Asn Ser Leu Asp Val Lys Ile  
 755 760 765  
 Ser Glu Ala Met Asn Asn Ile Asn Lys Phe Ile Arg Glu Cys Ser Val  
 770 775 780  
 Thr Tyr Leu Phe Lys Asn Met Leu Pro Lys Val Ile Asp Glu Leu Asn  
 785 790 795 800  
 Lys Phe Asp Leu Arg Thr Lys Thr Glu Leu Ile Asn Leu Ile Asp Ser  
 805 810 815  
 His Asn Ile Ile Leu Val Gly Glu Val Asp Arg Leu Lys Ala Lys Val  
 820 825 830  
 Asn Glu Ser Phe Glu Asn Thr Met Pro Phe Asn Ile Phe Ser Tyr Thr  
 835 840 845  
 Asn Asn Ser Leu Leu Lys Asp Ile Ile Asn Glu Tyr Phe Asn Ser Ile  
 850 855 860  
 Asn Asp Ser Lys Ile Leu Ser Leu Gln Asn Lys Lys Asn Ala Leu Val  
 865 870 875 880  
 Asp Thr Ser Gly Tyr Asn Ala Glu Val Arg Val Gly Asp Asn Val Gln  
 885 890 895  
 Leu Asn Thr Ile Tyr Thr Asn Asp Phe Lys Leu Ser Ser Ser Gly Asp  
 900 905 910  
 Lys Ile Ile Val Asn Leu Asn Asn Asn Ile Leu Tyr Ser Ala Ile Tyr  
 915 920 925  
 Glu Asn Ser Ser Val Ser Phe Trp Ile Lys Ile Ser Lys Asp Leu Thr  
 930 935 940

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Asn Ser His Asn Glu Tyr Thr Ile Ile Asn Ser Ile Glu Gln Asn Ser  
 945 950 955 960  
 Gly Trp Lys Leu Cys Ile Arg Asn Gly Asn Ile Glu Trp Ile Leu Gln  
 965 970 975  
 Asp Val Asn Arg Lys Tyr Lys Ser Leu Ile Phe Asp Tyr Ser Glu Ser  
 980 985 990  
 Leu Ser His Thr Gly Tyr Thr Asn Lys Trp Phe Phe Val Thr Ile Thr  
 995 1000 1005  
 Asn Asn Ile Met Gly Tyr Met Lys Leu Tyr Ile Asn Gly Glu Leu  
 1010 1015 1020  
 Lys Gln Ser Gln Lys Ile Glu Asp Leu Asp Glu Val Lys Leu Asp  
 1025 1030 1035  
 Lys Thr Ile Val Phe Gly Ile Asp Glu Asn Ile Asp Glu Asn Gln  
 1040 1045 1050  
 Met Leu Trp Ile Arg Asp Phe Asn Ile Phe Ser Lys Glu Leu Ser  
 1055 1060 1065  
 Asn Glu Asp Ile Asn Ile Val Tyr Glu Gly Gln Ile Leu Arg Asn  
 1070 1075 1080  
 Val Ile Lys Asp Tyr Trp Gly Asn Pro Leu Lys Phe Asp Thr Glu  
 1085 1090 1095  
 Tyr Tyr Ile Ile Asn Asp Asn Tyr Ile Asp Arg Tyr Ile Ala Pro  
 1100 1105 1110  
 Glu Ser Asn Val Leu Val Leu Val Arg Tyr Pro Asp Arg Ser Lys  
 1115 1120 1125  
 Leu Tyr Thr Gly Asn Pro Ile Thr Ile Lys Ser Val Ser Asp Lys  
 1130 1135 1140  
 Asn Pro Tyr Ser Arg Ile Leu Asn Gly Asp Asn Ile Ile Leu His  
 1145 1150 1155  
 Met Leu Tyr Asn Ser Arg Lys Tyr Met Ile Ile Arg Asp Thr Asp  
 1160 1165 1170  
 Thr Ile Tyr Ala Thr Gln Gly Gly Glu Cys Ser Gln Asn Cys Val  
 1175 1180 1185  
 Tyr Ala Leu Lys Leu Gln Ser Asn Leu Gly Asn Tyr Gly Ile Gly  
 1190 1195 1200  
 Ile Phe Ser Ile Lys Asn Ile Val Ser Lys Asn Lys Tyr Cys Ser  
 1205 1210 1215  
 Gln Ile Phe Ser Ser Phe Arg Glu Asn Thr Met Leu Leu Ala Asp  
 1220 1225 1230  
 Ile Tyr Lys Pro Trp Arg Phe Ser Phe Lys Asn Ala Tyr Thr Pro  
 1235 1240 1245  
 Val Ala Val Thr Asn Tyr Glu Thr Lys Leu Leu Ser Thr Ser Ser  
 1250 1255 1260  
 Phe Trp Lys Phe Ile Ser Arg Asp Pro Gly Trp Val Glu  
 1265 1270 1275

&lt;210&gt; SEQ ID NO 5

&lt;211&gt; LENGTH: 1251

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Clostridium botulinum (serotype E)

&lt;400&gt; SEQUENCE: 5

Met Pro Lys Ile Asn Ser Phe Asn Tyr Asn Asp Pro Val Asn Asp Arg  
 1 5 10 15

Thr Ile Leu Tyr Ile Lys Pro Gly Gly Cys Gln Glu Phe Tyr Lys Ser  
 20 25 30



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Phe Asn Ile Met Lys Asn Ile Trp Ile Ile Pro Glu Arg Asn Val Ile  
 35 40 45  
 Gly Thr Thr Pro Gln Asp Phe His Pro Pro Thr Ser Leu Lys Asn Gly  
 50 55 60  
 Asp Ser Ser Tyr Tyr Asp Pro Asn Tyr Leu Gln Ser Asp Glu Glu Lys  
 65 70 75 80  
 Asp Arg Phe Leu Lys Ile Val Thr Lys Ile Phe Asn Arg Ile Asn Asn  
 85 90 95  
 Asn Leu Ser Gly Gly Ile Leu Leu Glu Glu Leu Ser Lys Ala Asn Pro  
 100 105 110  
 Tyr Leu Gly Asn Asp Asn Thr Pro Asp Asn Gln Phe His Ile Gly Asp  
 115 120 125  
 Ala Ser Ala Val Glu Ile Lys Phe Ser Asn Gly Ser Gln Asp Ile Leu  
 130 135 140  
 Leu Pro Asn Val Ile Ile Met Gly Ala Glu Pro Asp Leu Phe Glu Thr  
 145 150 155 160  
 Asn Ser Ser Asn Ile Ser Leu Arg Asn Asn Tyr Met Pro Ser Asn His  
 165 170 175  
 Gly Phe Gly Ser Ile Ala Ile Val Thr Phe Ser Pro Glu Tyr Ser Phe  
 180 185 190  
 Arg Phe Asn Asp Asn Ser Met Asn Glu Phe Ile Gln Asp Pro Ala Leu  
 195 200 205  
 Thr Leu Met His Glu Leu Ile His Ser Leu His Gly Leu Tyr Gly Ala  
 210 215 220  
 Lys Gly Ile Thr Thr Lys Tyr Thr Ile Thr Gln Lys Gln Asn Pro Leu  
 225 230 235 240  
 Ile Thr Asn Ile Arg Gly Thr Asn Ile Glu Glu Phe Leu Thr Phe Gly  
 245 250 255  
 Gly Thr Asp Leu Asn Ile Ile Thr Ser Ala Gln Ser Asn Asp Ile Tyr  
 260 265 270  
 Thr Asn Leu Leu Ala Asp Tyr Lys Lys Ile Ala Ser Lys Leu Ser Lys  
 275 280 285  
 Val Gln Val Ser Asn Pro Leu Leu Asn Pro Tyr Lys Asp Val Phe Glu  
 290 295 300  
 Ala Lys Tyr Gly Leu Asp Lys Asp Ala Ser Gly Ile Tyr Ser Val Asn  
 305 310 315 320  
 Ile Asn Lys Phe Asn Asp Ile Phe Lys Lys Leu Tyr Ser Phe Thr Glu  
 325 330 335  
 Phe Asp Leu Ala Thr Lys Phe Gln Val Lys Cys Arg Gln Thr Tyr Ile  
 340 345 350  
 Gly Gln Tyr Lys Tyr Phe Lys Leu Ser Asn Leu Leu Asn Asp Ser Ile  
 355 360 365  
 Tyr Asn Ile Ser Glu Gly Tyr Asn Ile Asn Asn Leu Lys Val Asn Phe  
 370 375 380  
 Arg Gly Gln Asn Ala Asn Leu Asn Pro Arg Ile Ile Thr Pro Ile Thr  
 385 390 395 400  
 Gly Arg Gly Leu Val Lys Lys Ile Ile Arg Phe Cys Lys Asn Ile Val  
 405 410 415  
 Ser Val Lys Gly Ile Arg Lys Ser Ile Cys Ile Glu Ile Asn Asn Gly  
 420 425 430  
 Glu Leu Phe Phe Val Ala Ser Glu Asn Ser Tyr Asn Asp Asp Asn Ile  
 435 440 445

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Asn Thr Pro Lys Glu Ile Asp Asp Thr Val Thr Ser Asn Asn Asn Tyr  
 450 455 460  
 Glu Asn Asp Leu Asp Gln Val Ile Leu Asn Phe Asn Ser Glu Ser Ala  
 465 470 475 480  
 Pro Gly Leu Ser Asp Glu Lys Leu Asn Leu Thr Ile Gln Asn Asp Ala  
 485 490 495  
 Tyr Ile Pro Lys Tyr Asp Ser Asn Gly Thr Ser Asp Ile Glu Gln His  
 500 505 510  
 Asp Val Asn Glu Leu Asn Val Phe Phe Tyr Leu Asp Ala Gln Lys Val  
 515 520 525  
 Pro Glu Gly Glu Asn Asn Val Asn Leu Thr Ser Ser Ile Asp Thr Ala  
 530 535 540  
 Leu Leu Glu Gln Pro Lys Ile Tyr Thr Phe Phe Ser Ser Glu Phe Ile  
 545 550 555 560  
 Asn Asn Val Asn Lys Pro Val Gln Ala Ala Leu Phe Val Ser Trp Ile  
 565 570 575  
 Gln Gln Val Leu Val Asp Phe Thr Thr Glu Ala Asn Gln Lys Ser Thr  
 580 585 590  
 Val Asp Lys Ile Ala Asp Ile Ser Ile Val Val Pro Tyr Ile Gly Leu  
 595 600 605  
 Ala Leu Asn Ile Gly Asn Glu Ala Gln Lys Gly Asn Phe Lys Asp Ala  
 610 615 620  
 Leu Glu Leu Leu Gly Ala Gly Ile Leu Leu Glu Phe Glu Pro Glu Leu  
 625 630 635 640  
 Leu Ile Pro Thr Ile Leu Val Phe Thr Ile Lys Ser Phe Leu Gly Ser  
 645 650 655  
 Ser Asp Asn Lys Asn Lys Val Ile Lys Ala Ile Asn Asn Ala Leu Lys  
 660 665 670  
 Glu Arg Asp Glu Lys Trp Lys Glu Val Tyr Ser Phe Ile Val Ser Asn  
 675 680 685  
 Trp Met Thr Lys Ile Asn Thr Gln Phe Asn Lys Arg Lys Glu Gln Met  
 690 695 700  
 Tyr Gln Ala Leu Gln Asn Gln Val Asn Ala Ile Lys Thr Ile Ile Glu  
 705 710 715 720  
 Ser Lys Tyr Asn Ser Tyr Thr Leu Glu Glu Lys Asn Glu Leu Thr Asn  
 725 730 735  
 Lys Tyr Asp Ile Lys Gln Ile Glu Asn Glu Leu Asn Gln Lys Val Ser  
 740 745 750  
 Ile Ala Met Asn Asn Ile Asp Arg Phe Leu Thr Glu Ser Ser Ile Ser  
 755 760 765  
 Tyr Leu Met Lys Leu Ile Asn Glu Val Lys Ile Asn Lys Leu Arg Glu  
 770 775 780  
 Tyr Asp Glu Asn Val Lys Thr Tyr Leu Leu Asn Tyr Ile Ile Gln His  
 785 790 795 800  
 Gly Ser Ile Leu Gly Glu Ser Gln Gln Glu Leu Asn Ser Met Val Thr  
 805 810 815  
 Asp Thr Leu Asn Asn Ser Ile Pro Phe Lys Leu Ser Ser Tyr Thr Asp  
 820 825 830  
 Asp Lys Ile Leu Ile Ser Tyr Phe Asn Lys Phe Phe Lys Arg Ile Lys  
 835 840 845  
 Ser Ser Ser Val Leu Asn Met Arg Tyr Lys Asn Asp Lys Tyr Val Asp  
 850 855 860  
 Thr Ser Gly Tyr Asp Ser Asn Ile Asn Ile Asn Gly Asp Val Tyr Lys

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865	870	875	880
Tyr Pro Thr Asn Lys Asn Gln Phe Gly Ile Tyr Asn Asp Lys Leu Ser 885 890 895			
Glu Val Asn Ile Ser Gln Asn Asp Tyr Ile Ile Tyr Asp Asn Lys Tyr 900 905 910			
Lys Asn Phe Ser Ile Ser Phe Trp Val Arg Ile Pro Asn Tyr Asp Asn 915 920 925			
Lys Ile Val Asn Val Asn Asn Glu Tyr Thr Ile Ile Asn Cys Met Arg 930 935 940			
Asp Asn Asn Ser Gly Trp Lys Val Ser Leu Asn His Asn Glu Ile Ile 945 950 955 960			
Trp Thr Leu Gln Asp Asn Ala Gly Ile Asn Gln Lys Leu Ala Phe Asn 965 970 975			
Tyr Gly Asn Ala Asn Gly Ile Ser Asp Tyr Ile Asn Lys Trp Ile Phe 980 985 990			
Val Thr Ile Thr Asn Asp Arg Leu Gly Asp Ser Lys Leu Tyr Ile Asn 995 1000 1005			
Gly Asn Leu Ile Asp Gln Lys Ser Ile Leu Asn Leu Gly Asn Ile 1010 1015 1020			
His Val Ser Asp Asn Ile Leu Phe Lys Ile Val Asn Cys Ser Tyr 1025 1030 1035			
Thr Arg Tyr Ile Gly Ile Arg Tyr Phe Asn Ile Phe Asp Lys Glu 1040 1045 1050			
Leu Asp Glu Thr Glu Ile Gln Thr Leu Tyr Ser Asn Glu Pro Asn 1055 1060 1065			
Thr Asn Ile Leu Lys Asp Phe Trp Gly Asn Tyr Leu Leu Tyr Asp 1070 1075 1080			
Lys Glu Tyr Tyr Leu Leu Asn Val Leu Lys Pro Asn Asn Phe Ile 1085 1090 1095			
Asp Arg Arg Lys Asp Ser Thr Leu Ser Ile Asn Asn Ile Arg Ser 1100 1105 1110			
Thr Ile Leu Leu Ala Asn Arg Leu Tyr Ser Gly Ile Lys Val Lys 1115 1120 1125			
Ile Gln Arg Val Asn Asn Ser Ser Thr Asn Asp Asn Leu Val Arg 1130 1135 1140			
Lys Asn Asp Gln Val Tyr Ile Asn Phe Val Ala Ser Lys Thr His 1145 1150 1155			
Leu Phe Pro Leu Tyr Ala Asp Thr Ala Thr Thr Asn Lys Glu Lys 1160 1165 1170			
Thr Ile Lys Ile Ser Ser Ser Gly Asn Arg Phe Asn Gln Val Val 1175 1180 1185			
Val Met Asn Ser Val Gly Asn Asn Thr Met Asn Phe Lys Asn Asn 1190 1195 1200			
Asn Gly Asn Asn Ile Gly Leu Leu Gly Phe Lys Ala Asp Thr Val 1205 1210 1215			
Val Ala Ser Thr Trp Tyr Tyr Thr His Met Arg Asp His Thr Asn 1220 1225 1230			
Ser Asn Gly Cys Phe Trp Asn Phe Ile Ser Glu Glu His Gly Trp 1235 1240 1245			
Gln Glu Lys 1250			

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<211> LENGTH: 1277
<212> TYPE: PRT
<213> ORGANISM: Clostridium botulinum (serotype F)

<400> SEQUENCE: 6
Met Pro Val Val Ile Asn Ser Phe Asn Tyr Asn Asp Pro Val Asn Asp
1          5          10          15
Asp Thr Ile Leu Tyr Met Gln Ile Pro Tyr Glu Glu Lys Ser Lys Lys
20          25          30
Tyr Tyr Lys Ala Phe Glu Ile Met Arg Asn Val Trp Ile Ile Pro Glu
35          40          45
Arg Asn Thr Ile Gly Thr Asp Pro Ser Asp Phe Asp Pro Pro Ala Ser
50          55          60
Leu Glu Asn Gly Ser Ser Ala Tyr Tyr Asp Pro Asn Tyr Leu Thr Thr
65          70          75          80
Asp Ala Glu Lys Asp Arg Tyr Leu Lys Thr Thr Ile Lys Leu Phe Lys
85          90          95
Arg Ile Asn Ser Asn Pro Ala Gly Glu Val Leu Leu Gln Glu Ile Ser
100         105         110
Tyr Ala Lys Pro Tyr Leu Gly Asn Glu His Thr Pro Ile Asn Glu Phe
115         120         125
His Pro Val Thr Arg Thr Thr Ser Val Asn Ile Lys Ser Ser Thr Asn
130         135         140
Val Lys Ser Ser Ile Ile Leu Asn Leu Leu Val Leu Gly Ala Gly Pro
145         150         155         160
Asp Ile Phe Glu Asn Ser Ser Tyr Pro Val Arg Lys Leu Met Asp Ser
165         170         175
Gly Gly Val Tyr Asp Pro Ser Asn Asp Gly Phe Gly Ser Ile Asn Ile
180         185         190
Val Thr Phe Ser Pro Glu Tyr Glu Tyr Thr Phe Asn Asp Ile Ser Gly
195         200         205
Gly Tyr Asn Ser Ser Thr Glu Ser Phe Ile Ala Asp Pro Ala Ile Ser
210         215         220
Leu Ala His Glu Leu Ile His Ala Leu His Gly Leu Tyr Gly Ala Arg
225         230         235         240
Gly Val Thr Tyr Lys Glu Thr Ile Lys Val Lys Gln Ala Pro Leu Met
245         250         255
Ile Ala Ile Lys Pro Ile Arg Leu Glu Glu Phe Leu Thr Phe Gly Gly
260         265         270
Gln Asp Leu Asn Ile Ile Thr Ser Ala Met Lys Glu Lys Ile Tyr Asn
275         280         285
Asn Leu Leu Ala Asn Tyr Glu Lys Ile Ala Thr Arg Leu Ser Arg Val
290         295         300
Asn Ser Ala Pro Pro Glu Tyr Asp Ile Asn Glu Tyr Lys Asp Tyr Phe
305         310         315         320
Gln Trp Lys Tyr Gly Leu Asp Lys Asn Ala Asp Gly Ser Tyr Thr Val
325         330         335
Asn Glu Asn Lys Phe Asn Glu Ile Tyr Lys Lys Leu Tyr Ser Phe Thr
340         345         350
Glu Ile Asp Leu Ala Asn Lys Phe Lys Val Lys Cys Arg Asn Thr Tyr
355         360         365
Phe Ile Lys Tyr Gly Phe Leu Lys Val Pro Asn Leu Leu Asp Asp Asp
370         375         380
Ile Tyr Thr Val Ser Glu Gly Phe Asn Ile Gly Asn Leu Ala Val Asn

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Ile Gly Asn Asn Thr Met Asn Phe Gln Asn Asn Asn Gly Gly Asn
1220 1225 1230
Ile Gly Leu Leu Gly Phe His Ser Asn Asn Leu Val Ala Ser Ser
1235 1240 1245
Trp Tyr Tyr Asn Asn Ile Arg Lys Asn Thr Ser Ser Asn Gly Cys
1250 1255 1260
Phe Trp Ser Phe Ile Ser Lys Glu His Gly Trp Gln Glu Asn
1265 1270 1275

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<210> SEQ ID NO 7
<211> LENGTH: 1297
<212> TYPE: PRT
<213> ORGANISM: Clostridium botulinum (serotype G)

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<400> SEQUENCE: 7

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Met Pro Val Asn Ile Lys Asn Phe Asn Tyr Asn Asp Pro Ile Asn Asn
1 5 10 15
Asp Asp Ile Ile Met Met Glu Pro Phe Asn Asp Pro Gly Pro Gly Thr
20 25 30
Tyr Tyr Lys Ala Phe Arg Ile Ile Asp Arg Ile Trp Ile Val Pro Glu
35 40 45
Arg Phe Thr Tyr Gly Phe Gln Pro Asp Gln Phe Asn Ala Ser Thr Gly
50 55 60
Val Phe Ser Lys Asp Val Tyr Glu Tyr Tyr Asp Pro Thr Tyr Leu Lys
65 70 75 80
Thr Asp Ala Glu Lys Asp Lys Phe Leu Lys Thr Met Ile Lys Leu Phe
85 90 95
Asn Arg Ile Asn Ser Lys Pro Ser Gly Gln Arg Leu Leu Asp Met Ile
100 105 110
Val Asp Ala Ile Pro Tyr Leu Gly Asn Ala Ser Thr Pro Pro Asp Lys
115 120 125
Phe Ala Ala Asn Val Ala Asn Val Ser Ile Asn Lys Lys Ile Ile Gln
130 135 140
Pro Gly Ala Glu Asp Gln Ile Lys Gly Leu Met Thr Asn Leu Ile Ile
145 150 155 160
Phe Gly Pro Gly Pro Val Leu Ser Asp Asn Phe Thr Asp Ser Met Ile
165 170 175
Met Asn Gly His Ser Pro Ile Ser Glu Gly Phe Gly Ala Arg Met Met
180 185 190
Ile Arg Phe Cys Pro Ser Cys Leu Asn Val Phe Asn Asn Val Gln Glu
195 200 205
Asn Lys Asp Thr Ser Ile Phe Ser Arg Arg Ala Tyr Phe Ala Asp Pro
210 215 220
Ala Leu Thr Leu Met His Glu Leu Ile His Val Leu His Gly Leu Tyr
225 230 235 240
Gly Ile Lys Ile Ser Asn Leu Pro Ile Thr Pro Asn Thr Lys Glu Phe
245 250 255
Phe Met Gln His Ser Asp Pro Val Gln Ala Glu Glu Leu Tyr Thr Phe
260 265 270
Gly Gly His Asp Pro Ser Val Ile Ser Pro Ser Thr Asp Met Asn Ile
275 280 285
Tyr Asn Lys Ala Leu Gln Asn Phe Gln Asp Ile Ala Asn Arg Leu Asn
290 295 300
Ile Val Ser Ser Ala Gln Gly Ser Gly Ile Asp Ile Ser Leu Tyr Lys
305 310 315 320

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Lys Ile Ile Glu Asp Gln Tyr Asn Arg Tyr Ser Glu Glu Asp Lys Met  
                   740  745  750

Asn Ile Asn Ile Asp Phe Asn Asp Ile Asp Phe Lys Leu Asn Gln Ser  
                   755  760  765

Ile Asn Leu Ala Ile Asn Asn Ile Asp Asp Phe Ile Asn Gln Cys Ser  
                   770  775  780

Ile Ser Tyr Leu Met Asn Arg Met Ile Pro Leu Ala Val Lys Lys Leu  
                   785  790  795  800

Lys Asp Phe Asp Asp Asn Leu Lys Arg Asp Leu Leu Glu Tyr Ile Asp  
                                   805  810  815

Thr Asn Glu Leu Tyr Leu Leu Asp Glu Val Asn Ile Leu Lys Ser Lys  
                                   820  825  830

Val Asn Arg His Leu Lys Asp Ser Ile Pro Phe Asp Leu Ser Leu Tyr  
                                   835  840  845

Thr Lys Asp Thr Ile Leu Ile Gln Val Phe Asn Asn Tyr Ile Ser Asn  
                                   850  855  860

Ile Ser Ser Asn Ala Ile Leu Ser Leu Ser Tyr Arg Gly Gly Arg Leu  
                                   865  870  875  880

Ile Asp Ser Ser Gly Tyr Gly Ala Thr Met Asn Val Gly Ser Asp Val  
                                   885  890  895

Ile Phe Asn Asp Ile Gly Asn Gly Gln Phe Lys Leu Asn Asn Ser Glu  
                                   900  905  910

Asn Ser Asn Ile Thr Ala His Gln Ser Lys Phe Val Val Tyr Asp Ser  
                                   915  920  925

Met Phe Asp Asn Phe Ser Ile Asn Phe Trp Val Arg Thr Pro Lys Tyr  
                                   930  935  940

Asn Asn Asn Asp Ile Gln Thr Tyr Leu Gln Asn Glu Tyr Thr Ile Ile  
                                   945  950  955  960

Ser Cys Ile Lys Asn Asp Ser Gly Trp Lys Val Ser Ile Lys Gly Asn  
                                   965  970  975

Arg Ile Ile Trp Thr Leu Ile Asp Val Asn Ala Lys Ser Lys Ser Ile  
                                   980  985  990

Phe Phe Glu Tyr Ser Ile Lys Asp Asn Ile Ser Asp Tyr Ile Asn Lys  
                                   995  1000  1005

Trp Phe Ser Ile Thr Ile Thr Asn Asp Arg Leu Gly Asn Ala Asn  
                                   1010  1015  1020

Ile Tyr Ile Asn Gly Ser Leu Lys Lys Ser Glu Lys Ile Leu Asn  
                                   1025  1030  1035

Leu Asp Arg Ile Asn Ser Ser Asn Asp Ile Asp Phe Lys Leu Ile  
                                   1040  1045  1050

Asn Cys Thr Asp Thr Thr Lys Phe Val Trp Ile Lys Asp Phe Asn  
                                   1055  1060  1065

Ile Phe Gly Arg Glu Leu Asn Ala Thr Glu Val Ser Ser Leu Tyr  
                                   1070  1075  1080

Trp Ile Gln Ser Ser Thr Asn Thr Leu Lys Asp Phe Trp Gly Asn  
                                   1085  1090  1095

Pro Leu Arg Tyr Asp Thr Gln Tyr Tyr Leu Phe Asn Gln Gly Met  
                                   1100  1105  1110

Gln Asn Ile Tyr Ile Lys Tyr Phe Ser Lys Ala Ser Met Gly Glu  
                                   1115  1120  1125

Thr Ala Pro Arg Thr Asn Phe Asn Asn Ala Ala Ile Asn Tyr Gln  
                                   1130  1135  1140

Asn Leu Tyr Leu Leu Arg Phe Ile Ile Lys Lys Ala Ser Asn Ser

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1145	1150	1155
Arg Asn Ile Asn Asn Asp Asn Ile Val Arg Glu Gly Asp Tyr Ile		
1160	1165	1170
Tyr Leu Asn Ile Asp Asn Ile Ser Asp Glu Ser Tyr Arg Val Tyr		
1175	1180	1185
Val Leu Val Asn Ser Lys Glu Ile Gln Thr Gln Leu Phe Leu Ala		
1190	1195	1200
Pro Ile Asn Asp Asp Pro Thr Phe Tyr Asp Val Leu Gln Ile Gly		
1205	1210	1215
Lys Lys Tyr Tyr Glu Lys Thr Thr Tyr Asn Cys Gln Ile Leu Cys		
1220	1225	1230
Glu Lys Asp Thr Lys Thr Phe Gly Leu Phe Gly Ile Gly Lys Phe		
1235	1240	1245
Val Lys Asp Tyr Gly Tyr Val Trp Asp Thr Tyr Asp Asn Tyr Phe		
1250	1255	1260
Cys Ile Ser Gln Trp Tyr Leu Arg Arg Ile Ser Glu Asn Ile Asn		
1265	1270	1275
Lys Leu Arg Leu Gly Cys Asn Trp Gln Phe Ile Pro Val Asp Glu		
1280	1285	1290
Gly Trp Thr Glu		
1295		

<210> SEQ ID NO 8  
 <211> LENGTH: 1300  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Atoxic derivative of Clostridium botulinum  
 serotype A neurotoxin

<400> SEQUENCE: 8

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly			
1	5	10	15
Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro			
	20	25	30
Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg			
	35	40	45
Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu			
	50	55	60
Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr			
	65	70	75
Asp Asn Glu Lys Asp Asn Tyr Leu Lys Gly Val Thr Lys Leu Phe Glu			
	85	90	95
Arg Ile Tyr Ser Thr Asp Leu Gly Arg Met Leu Leu Thr Ser Ile Val			
	100	105	110
Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys			
	115	120	125
Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr			
	130	135	140
Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile			
	145	150	155
Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr			
	165	170	175
Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe			
	180	185	190

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Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu  
 195 200 205  
 Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Glu  
 210 215 220  
 Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn  
 225 230 235 240  
 Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu  
 245 250 255  
 Glu Val Ser Phe Glu Glu Leu Arg Thr Phe Gly Gly His Asp Ala Lys  
 260 265 270  
 Phe Ile Asp Ser Leu Gln Glu Asn Glu Phe Arg Leu Tyr Tyr Tyr Asn  
 275 280 285  
 Lys Phe Lys Asp Ile Ala Ser Thr Leu Asn Lys Ala Lys Ser Ile Val  
 290 295 300  
 Gly Thr Thr Ala Ser Leu Gln Tyr Met Lys Asn Val Phe Lys Glu Lys  
 305 310 315 320  
 Tyr Leu Leu Ser Glu Asp Thr Ser Gly Lys Phe Ser Val Asp Lys Leu  
 325 330 335  
 Lys Phe Asp Lys Leu Tyr Lys Met Leu Thr Glu Ile Thr Thr Glu Asp  
 340 345 350  
 Asn Phe Val Lys Phe Phe Lys Val Leu Asn Arg Lys Thr Tyr Leu Asn  
 355 360 365  
 Phe Asp Lys Ala Val Phe Lys Ile Asn Ile Val Pro Lys Val Asn Tyr  
 370 375 380  
 Thr Ile Tyr Asp Gly Phe Asn Leu Arg Asn Thr Asn Leu Ala Ala Asn  
 385 390 395 400  
 Phe Asn Gly Gln Asn Thr Glu Ile Asn Asn Met Asn Phe Thr Lys Leu  
 405 410 415  
 Lys Asn Phe Thr Gly Leu Phe Glu Phe Tyr Lys Leu Leu Cys Val Arg  
 420 425 430  
 Gly Ile Ile Thr Ser His Thr Gln Ser Leu Asp Gln Gly Tyr Asn Asp  
 435 440 445  
 Asp Asp Asp Lys Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn Trp  
 450 455 460  
 Asp Leu Phe Phe Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu Asn  
 465 470 475 480  
 Lys Gly Glu Glu Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu Glu  
 485 490 495  
 Asn Ile Ser Leu Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn Phe  
 500 505 510  
 Asp Asn Glu Pro Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp Ile  
 515 520 525  
 Ile Gly Gln Leu Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn Gly  
 530 535 540  
 Lys Lys Tyr Glu Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg Ala  
 545 550 555 560  
 Gln Glu Phe Glu His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser Val  
 565 570 575  
 Asn Glu Ala Leu Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser Ser  
 580 585 590  
 Asp Tyr Val Lys Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe Leu  
 595 600 605  
 Gly Trp Val Glu Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser Glu

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610			615			620									
Val	Ser	Thr	Thr	Asp	Lys	Ile	Ala	Asp	Ile	Thr	Ile	Ile	Ile	Pro	Tyr
625					630					635				640	
Ile	Gly	Pro	Ala	Leu	Asn	Ile	Gly	Asn	Met	Leu	Tyr	Lys	Asp	Asp	Phe
			645						650					655	
Val	Gly	Ala	Leu	Ile	Phe	Ser	Gly	Ala	Val	Ile	Leu	Leu	Glu	Phe	Ile
			660					665					670		
Pro	Glu	Ile	Ala	Ile	Pro	Val	Leu	Gly	Thr	Phe	Ala	Leu	Val	Ser	Tyr
		675					680					685			
Ile	Ala	Asn	Lys	Val	Leu	Thr	Val	Gln	Thr	Ile	Asp	Asn	Ala	Leu	Ser
	690						695				700				
Lys	Arg	Asn	Glu	Lys	Trp	Asp	Glu	Val	Tyr	Lys	Tyr	Ile	Val	Thr	Asn
705					710					715				720	
Trp	Leu	Ala	Lys	Val	Asn	Thr	Gln	Ile	Asp	Leu	Ile	Arg	Lys	Lys	Met
			725						730					735	
Lys	Glu	Ala	Leu	Glu	Asn	Gln	Ala	Glu	Ala	Thr	Lys	Ala	Ile	Ile	Asn
			740					745					750		
Tyr	Gln	Tyr	Asn	Gln	Tyr	Thr	Glu	Glu	Glu	Lys	Asn	Asn	Ile	Asn	Phe
		755					760					765			
Asn	Ile	Asp	Asp	Leu	Ser	Ser	Lys	Leu	Asn	Glu	Ser	Ile	Asn	Lys	Ala
	770						775				780				
Met	Ile	Asn	Ile	Asn	Lys	Phe	Leu	Asn	Gln	Cys	Ser	Val	Ser	Tyr	Leu
785					790					795					800
Met	Asn	Ser	Met	Ile	Pro	Tyr	Gly	Val	Lys	Arg	Leu	Glu	Asp	Phe	Asp
				805					810					815	
Ala	Ser	Leu	Lys	Asp	Ala	Leu	Leu	Lys	Tyr	Ile	Tyr	Asp	Asn	Arg	Gly
			820					825					830		
Thr	Leu	Ile	Gly	Gln	Val	Asp	Arg	Leu	Lys	Asp	Lys	Val	Asn	Asn	Thr
		835					840					845			
Leu	Ser	Thr	Asp	Ile	Pro	Phe	Gln	Leu	Ser	Lys	Tyr	Val	Asp	Asn	Gln
	850						855				860				
Arg	Leu	Leu	Ser	Thr	Phe	Thr	Glu	Tyr	Ile	Asn	Asn	Ile	Ile	Asn	Thr
865					870					875					880
Ser	Ile	Leu	Asn	Leu	Arg	Tyr	Glu	Ser	Asn	His	Leu	Ile	Asp	Leu	Ser
				885					890					895	
Arg	Tyr	Ala	Ser	Lys	Ile	Asn	Ile	Gly	Ser	Lys	Val	Asn	Phe	Asp	Pro
			900					905					910		
Ile	Asp	Lys	Asn	Gln	Ile	Gln	Leu	Phe	Asn	Leu	Glu	Ser	Ser	Lys	Ile
		915					920					925			
Glu	Val	Ile	Leu	Lys	Asn	Ala	Ile	Val	Tyr	Asn	Ser	Met	Tyr	Glu	Asn
	930				935						940				
Phe	Ser	Thr	Ser	Phe	Trp	Ile	Arg	Ile	Pro	Lys	Tyr	Phe	Asn	Ser	Ile
945					950					955					960
Ser	Leu	Asn	Asn	Glu	Tyr	Thr	Ile	Ile	Asn	Cys	Met	Glu	Asn	Asn	Ser
				965						970				975	
Gly	Trp	Lys	Val	Ser	Leu	Asn	Tyr	Gly	Glu	Ile	Ile	Trp	Thr	Leu	Gln
			980					985					990		
Asp	Thr	Gln	Glu	Ile	Lys	Gln	Arg	Val	Val	Phe	Lys	Tyr	Ser	Gln	Met
			995				1000						1005		
Ile	Asn	Ile	Ser	Asp	Tyr	Ile	Asn	Arg	Trp	Ile	Phe	Val	Thr	Ile	
	1010						1015					1020			
Thr	Asn	Asn	Arg	Leu	Asn	Asn	Ser	Lys	Ile	Tyr	Ile	Asn	Gly	Arg	
	1025						1030					1035			

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Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His Ala  
 1040 1045 1050  
 Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr His  
 1055 1060 1065  
 Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu Leu  
 1070 1075 1080  
 Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn Ser  
 1085 1090 1095  
 Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp Lys  
 1100 1105 1110  
 Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val Asp  
 1115 1120 1125  
 Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly Pro  
 1130 1135 1140  
 Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser Leu  
 1145 1150 1155  
 Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly Asn  
 1160 1165 1170  
 Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn Val  
 1175 1180 1185  
 Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser Gln  
 1190 1195 1200  
 Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp Val  
 1205 1210 1215  
 Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp Gln  
 1220 1225 1230  
 Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn Gly  
 1235 1240 1245  
 Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile Ala  
 1250 1255 1260  
 Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser  
 1265 1270 1275  
 Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp  
 1280 1285 1290  
 Gly Trp Gly Glu Arg Pro Leu  
 1295 1300

<210> SEQ ID NO 9  
 <211> LENGTH: 1295  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Atoxic derivative of Clostridium botulinum  
 serotype B neurotoxin

<400> SEQUENCE: 9

Met Pro Val Thr Ile Asn Asn Phe Asn Tyr Asn Asp Pro Ile Asp Asn  
 1 5 10 15  
 Asn Asn Ile Ile Met Met Glu Pro Pro Phe Ala Arg Gly Thr Gly Arg  
 20 25 30  
 Tyr Tyr Lys Ala Phe Lys Ile Thr Asp Arg Ile Trp Ile Ile Pro Glu  
 35 40 45  
 Arg Tyr Thr Phe Gly Tyr Lys Pro Glu Asp Phe Asn Lys Ser Ser Gly  
 50 55 60  
 Ile Phe Asn Arg Asp Val Cys Glu Tyr Tyr Asp Pro Asp Tyr Leu Asn

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65	70	75	80
Thr Asn Asp Lys Lys Asn Ile Phe Leu Gln Thr Met Ile Lys Leu Phe 85 90 95			
Asn Arg Ile Lys Ser Lys Pro Leu Gly Glu Lys Leu Leu Glu Met Ile 100 105 110			
Ile Asn Gly Ile Pro Tyr Leu Gly Asp Arg Arg Val Pro Leu Glu Glu 115 120 125			
Phe Asn Thr Asn Ile Ala Ser Val Thr Val Asn Lys Leu Ile Ser Asn 130 135 140			
Pro Gly Glu Val Glu Arg Lys Lys Gly Ile Phe Ala Asn Leu Ile Ile 145 150 155 160			
Phe Gly Pro Gly Pro Val Leu Asn Glu Asn Glu Thr Ile Asp Ile Gly 165 170 175			
Ile Gln Asn His Phe Ala Ser Arg Glu Gly Phe Gly Gly Ile Met Gln 180 185 190			
Met Lys Phe Cys Pro Glu Tyr Val Ser Val Phe Asn Asn Val Gln Glu 195 200 205			
Asn Lys Gly Ala Ser Ile Phe Asn Arg Arg Gly Tyr Phe Ser Asp Pro 210 215 220			
Ala Leu Ile Leu Met His Glu Leu Ile His Val Leu His Gly Leu Tyr 225 230 235 240			
Gly Ile Lys Val Asp Asp Leu Pro Ile Val Pro Asn Glu Lys Lys Phe 245 250 255			
Phe Met Gln Ser Thr Asp Ala Ile Gln Ala Glu Glu Leu Tyr Thr Phe 260 265 270			
Gly Gly Gln Asp Pro Ser Ile Ile Thr Pro Ser Thr Asp Lys Ser Ile 275 280 285			
Tyr Asp Lys Val Leu Gln Asn Phe Arg Gly Ile Val Asp Arg Leu Asn 290 295 300			
Lys Val Leu Val Cys Ile Ser Asp Pro Asn Ile Asn Ile Asn Ile Tyr 305 310 315 320			
Lys Asn Lys Phe Lys Asp Lys Tyr Lys Phe Val Glu Asp Ser Glu Gly 325 330 335			
Lys Tyr Ser Ile Asp Val Glu Ser Phe Asp Lys Leu Tyr Lys Ser Leu 340 345 350			
Met Phe Gly Phe Thr Glu Thr Asn Ile Ala Glu Asn Tyr Lys Ile Lys 355 360 365			
Thr Arg Ala Ser Tyr Phe Ser Asp Ser Leu Pro Pro Val Lys Ile Lys 370 375 380			
Asn Leu Leu Asp Asn Glu Ile Tyr Thr Ile Glu Glu Gly Phe Asn Ile 385 390 395 400			
Ser Asp Lys Asp Met Glu Lys Glu Tyr Arg Gly Gln Asn Lys Ala Ile 405 410 415			
Asn Lys Gln Ala Tyr Glu Glu Ile Ser Lys Glu His Leu Ala Val Tyr 420 425 430			
Lys Ile Gln Met Cys Lys Ser Val Asp Asp Asp Asp Lys Ala Pro Gly 435 440 445			
Ile Cys Ile Asp Val Asp Asn Glu Asp Leu Phe Phe Ile Ala Asp Lys 450 455 460			
Asn Ser Phe Ser Asp Asp Leu Ser Lys Asn Glu Arg Ile Glu Tyr Asn 465 470 475 480			
Thr Gln Ser Asn Tyr Ile Glu Asn Asp Phe Pro Ile Asn Glu Leu Ile 485 490 495			



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Ile Arg Val Thr Gln Asn Gln Asn Ile Ile Phe Asn Ser Val Phe Leu  
915 920 925

Asp Phe Ser Val Ser Phe Trp Ile Arg Ile Pro Lys Tyr Lys Asn Asp  
930 935 940

Gly Ile Gln Asn Tyr Ile His Asn Glu Tyr Thr Ile Ile Asn Cys Met  
945 950 955 960

Lys Asn Asn Ser Gly Trp Lys Ile Ser Ile Arg Gly Asn Arg Ile Ile  
965 970 975

Trp Thr Leu Ile Asp Ile Asn Gly Lys Thr Lys Ser Val Phe Phe Glu  
980 985 990

Tyr Asn Ile Arg Glu Asp Ile Ser Glu Tyr Ile Asn Arg Trp Phe Phe  
995 1000 1005

Val Thr Ile Thr Asn Asn Leu Asn Asn Ala Lys Ile Tyr Ile Asn  
1010 1015 1020

Gly Lys Leu Glu Ser Asn Thr Asp Ile Lys Asp Ile Arg Glu Val  
1025 1030 1035

Ile Ala Asn Gly Glu Ile Ile Phe Lys Leu Asp Gly Asp Ile Asp  
1040 1045 1050

Arg Thr Gln Phe Ile Trp Met Lys Tyr Phe Ser Ile Phe Asn Thr  
1055 1060 1065

Glu Leu Ser Gln Ser Asn Ile Glu Glu Arg Tyr Lys Ile Gln Ser  
1070 1075 1080

Tyr Ser Glu Tyr Leu Lys Asp Phe Trp Gly Asn Pro Leu Met Tyr  
1085 1090 1095

Asn Lys Glu Tyr Tyr Met Phe Asn Ala Gly Asn Lys Asn Ser Tyr  
1100 1105 1110

Ile Lys Leu Lys Lys Asp Ser Pro Val Gly Glu Ile Leu Thr Arg  
1115 1120 1125

Ser Lys Tyr Asn Gln Asn Ser Lys Tyr Ile Asn Tyr Arg Asp Leu  
1130 1135 1140

Tyr Ile Gly Glu Lys Phe Ile Ile Arg Arg Lys Ser Asn Ser Gln  
1145 1150 1155

Ser Ile Asn Asp Asp Ile Val Arg Lys Glu Asp Tyr Ile Tyr Leu  
1160 1165 1170

Asp Phe Phe Asn Leu Asn Gln Glu Trp Arg Val Tyr Thr Tyr Lys  
1175 1180 1185

Tyr Phe Lys Lys Glu Glu Glu Lys Leu Phe Leu Ala Pro Ile Ser  
1190 1195 1200

Asp Ser Asp Glu Phe Tyr Asn Thr Ile Gln Ile Lys Glu Tyr Asp  
1205 1210 1215

Glu Gln Pro Thr Tyr Ser Cys Gln Leu Leu Phe Lys Lys Asp Glu  
1220 1225 1230

Glu Ser Thr Asp Glu Ile Gly Leu Ile Gly Ile His Arg Phe Tyr  
1235 1240 1245

Glu Ser Gly Ile Val Phe Glu Glu Tyr Lys Asp Tyr Phe Cys Ile  
1250 1255 1260

Ser Lys Trp Tyr Leu Lys Glu Val Lys Arg Lys Pro Tyr Asn Leu  
1265 1270 1275

Lys Leu Gly Cys Asn Trp Gln Phe Ile Pro Lys Asp Glu Gly Trp  
1280 1285 1290

Thr Glu  
1295



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<210> SEQ ID NO 10
<211> LENGTH: 1295
<212> TYPE: PRT
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Atoxic derivative of Clostridium botulinum
        serotype C neurotoxin

<400> SEQUENCE: 10

Met Pro Ile Thr Ile Asn Asn Phe Asn Tyr Ser Asp Pro Val Asp Asn
 1           5           10           15

Lys Asn Ile Leu Tyr Leu Asp Thr His Leu Asn Thr Leu Ala Asn Glu
 20           25           30

Pro Glu Lys Ala Phe Arg Ile Thr Gly Asn Ile Trp Val Ile Pro Asp
 35           40           45

Arg Phe Ser Arg Asn Ser Asn Pro Asn Leu Asn Lys Pro Pro Arg Val
 50           55           60

Thr Ser Pro Lys Ser Gly Tyr Tyr Asp Pro Asn Tyr Leu Ser Thr Asp
 65           70           75           80

Ser Asp Lys Asp Pro Phe Leu Lys Glu Ile Ile Lys Leu Phe Lys Arg
 85           90           95

Ile Asn Ser Arg Glu Ile Gly Glu Glu Leu Ile Tyr Arg Leu Ser Thr
 100          105          110

Asp Ile Pro Phe Pro Gly Asn Asn Asn Thr Pro Ile Asn Thr Phe Asp
 115          120          125

Phe Asp Val Asp Phe Asn Ser Val Asp Val Lys Thr Arg Gln Gly Asn
 130          135          140

Asn Trp Val Lys Thr Gly Ser Ile Asn Pro Ser Val Ile Ile Thr Gly
 145          150          155          160

Pro Arg Glu Asn Ile Ile Asp Pro Glu Thr Ser Thr Phe Lys Leu Thr
 165          170          175

Asn Asn Thr Phe Ala Ala Gln Glu Gly Phe Gly Ala Leu Ser Ile Ile
 180          185          190

Ser Ile Ser Pro Arg Phe Met Leu Thr Tyr Ser Asn Ala Thr Asn Asp
 195          200          205

Val Gly Glu Gly Arg Phe Ser Lys Ser Glu Phe Cys Met Asp Pro Ile
 210          215          220

Leu Ile Leu Met His Glu Leu Asn His Ala Met His Asn Leu Tyr Gly
 225          230          235          240

Ile Ala Ile Pro Asn Asp Gln Thr Ile Ser Ser Val Thr Ser Asn Ile
 245          250          255

Phe Tyr Ser Gln Tyr Asn Val Lys Leu Glu Tyr Ala Glu Ile Tyr Ala
 260          265          270

Phe Gly Gly Pro Thr Ile Asp Leu Ile Pro Lys Ser Ala Arg Lys Tyr
 275          280          285

Phe Glu Glu Lys Ala Leu Asp Tyr Tyr Arg Ser Ile Ala Lys Arg Leu
 290          295          300

Asn Ser Ile Thr Thr Ala Asn Pro Ser Ser Phe Asn Lys Tyr Ile Gly
 305          310          315          320

Glu Tyr Lys Gln Lys Leu Ile Arg Lys Tyr Arg Phe Val Val Glu Ser
 325          330          335

Ser Gly Glu Val Thr Val Asn Arg Asn Lys Phe Val Glu Leu Tyr Asn
 340          345          350

Glu Leu Thr Gln Ile Phe Thr Glu Phe Asn Tyr Ala Lys Ile Tyr Asn
 355          360          365

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Val	Gln	Asn	Arg	Lys	Ile	Tyr	Leu	Ser	Asn	Val	Tyr	Thr	Pro	Val	Thr
370						375				380					
Ala	Asn	Ile	Leu	Asp	Asp	Asn	Val	Tyr	Asp	Ile	Gln	Asn	Gly	Phe	Asn
385				390					395						400
Ile	Pro	Lys	Ser	Asn	Leu	Asn	Val	Leu	Phe	Met	Gly	Gln	Asn	Leu	Ser
			405						410					415	
Arg	Asn	Pro	Ala	Leu	Arg	Lys	Val	Asn	Pro	Glu	Asn	Met	Leu	Tyr	Leu
			420					425					430		
Phe	Thr	Lys	Phe	Cys	His	Lys	Ala	Ile	Asp	Gly	Gln	Ser	Leu	Tyr	Asn
	435						440					445			
Asp	Asp	Asp	Asp	Lys	Thr	Leu	Asp	Cys	Arg	Glu	Leu	Leu	Val	Lys	Asn
450						455					460				
Thr	Asp	Leu	Pro	Phe	Ile	Gly	Asp	Ile	Ser	Asp	Val	Lys	Thr	Asp	Ile
465					470					475					480
Phe	Leu	Arg	Lys	Asp	Ile	Asn	Glu	Glu	Thr	Glu	Val	Ile	Tyr	Tyr	Pro
			485						490					495	
Asp	Asn	Val	Ser	Val	Asp	Gln	Val	Ile	Leu	Ser	Lys	Asn	Thr	Ser	Glu
			500					505					510		
His	Gly	Gln	Leu	Asp	Leu	Leu	Tyr	Pro	Ser	Ile	Asp	Ser	Glu	Ser	Glu
	515						520					525			
Ile	Leu	Pro	Gly	Glu	Asn	Gln	Val	Phe	Tyr	Asp	Asn	Arg	Thr	Gln	Asn
	530					535						540			
Val	Asp	Tyr	Leu	Asn	Ser	Tyr	Tyr	Tyr	Leu	Glu	Ser	Gln	Lys	Leu	Ser
545					550					555					560
Asp	Asn	Val	Glu	Asp	Phe	Thr	Phe	Thr	Arg	Ser	Ile	Glu	Glu	Ala	Leu
			565						570					575	
Asp	Asn	Ser	Ala	Lys	Val	Tyr	Thr	Tyr	Phe	Pro	Thr	Leu	Ala	Asn	Lys
			580					585					590		
Val	Asn	Ala	Gly	Val	Gln	Gly	Gly	Leu	Phe	Leu	Met	Trp	Ala	Asn	Asp
	595					600						605			
Val	Val	Glu	Asp	Phe	Thr	Thr	Asn	Ile	Leu	Arg	Lys	Asp	Thr	Leu	Asp
	610					615					620				
Lys	Ile	Ser	Asp	Val	Ser	Ala	Ile	Ile	Pro	Tyr	Ile	Gly	Pro	Ala	Leu
625					630					635					640
Asn	Ile	Ser	Asn	Ser	Val	Arg	Arg	Gly	Asn	Phe	Thr	Glu	Ala	Phe	Ala
			645						650					655	
Val	Thr	Gly	Val	Thr	Ile	Leu	Leu	Glu	Ala	Phe	Pro	Glu	Phe	Thr	Ile
			660					665					670		
Pro	Ala	Leu	Gly	Ala	Phe	Val	Ile	Tyr	Ser	Lys	Val	Gln	Glu	Arg	Asn
		675					680					685			
Glu	Ile	Ile	Lys	Thr	Ile	Asp	Asn	Cys	Leu	Glu	Gln	Arg	Ile	Lys	Arg
	690					695					700				
Trp	Lys	Asp	Ser	Tyr	Glu	Trp	Met	Met	Gly	Thr	Trp	Leu	Ser	Arg	Ile
705					710					715					720
Ile	Thr	Gln	Phe	Asn	Asn	Ile	Ser	Tyr	Gln	Met	Tyr	Asp	Ser	Leu	Asn
				725					730					735	
Tyr	Gln	Ala	Gly	Ala	Ile	Lys	Ala	Lys	Ile	Asp	Leu	Glu	Tyr	Lys	Lys
			740					745					750		
Tyr	Ser	Gly	Ser	Asp	Lys	Glu	Asn	Ile	Lys	Ser	Gln	Val	Glu	Asn	Leu
		755					760					765			
Lys	Asn	Ser	Leu	Asp	Val	Lys	Ile	Ser	Glu	Ala	Met	Asn	Asn	Ile	Asn
	770					775					780				
Lys	Phe	Ile	Arg	Glu	Cys	Ser	Val	Thr	Tyr	Leu	Phe	Lys	Asn	Met	Leu

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785	790	795	800
Pro Lys Val Ile Asp 805	Glu Leu Asn Glu Phe Asp 810	Arg Asn Thr Lys Ala 815	
Lys Leu Ile Asn Leu Ile Asp Ser His Asn Ile Ile Leu Val Gly Glu 820	825	830	
Val Asp Lys Leu Lys Ala Lys Val Asn Asn Ser Phe Gln Asn Thr Ile 835	840	845	
Pro Phe Asn Ile Phe Ser Tyr Thr Asn Asn Ser Leu Leu Lys Asp Ile 850	855	860	
Ile Asn Glu Tyr Phe Asn Asn Ile Asn Asp Ser Lys Ile Leu Ser Leu 865	870	875	880
Gln Asn Arg Lys Asn Thr Leu Val Asp Thr Ser Gly Tyr Asn Ala Glu 885	890	895	
Val Ser Glu Glu Gly Asp Val Gln Leu Asn Pro Ile Phe Pro Phe Asp 900	905	910	
Phe Lys Leu Gly Ser Ser Gly Glu Asp Arg Gly Lys Val Ile Val Thr 915	920	925	
Gln Asn Glu Asn Ile Val Tyr Asn Ser Met Tyr Glu Ser Phe Ser Ile 930	935	940	
Ser Phe Trp Ile Arg Ile Asn Lys Trp Val Ser Asn Leu Pro Gly Tyr 945	950	955	960
Thr Ile Ile Asp Ser Val Lys Asn Asn Ser Gly Trp Ser Ile Gly Ile 965	970	975	
Ile Ser Asn Phe Leu Val Phe Thr Leu Lys Gln Asn Glu Asp Ser Glu 980	985	990	
Gln Ser Ile Asn Phe Ser Tyr Asp Ile Ser Asn Asn Ala Pro Gly Tyr 995	1000	1005	
Asn Lys Trp Phe Phe Val Thr Val Thr Asn Asn Met Met Gly Asn 1010	1015	1020	
Met Lys Ile Tyr Ile Asn Gly Lys Leu Ile Asp Thr Ile Lys Val 1025	1030	1035	
Lys Glu Leu Thr Gly Ile Asn Phe Ser Lys Thr Ile Thr Phe Glu 1040	1045	1050	
Ile Asn Lys Ile Pro Asp Thr Gly Leu Ile Thr Ser Asp Ser Asp 1055	1060	1065	
Asn Ile Asn Met Trp Ile Arg Asp Phe Tyr Ile Phe Ala Lys Glu 1070	1075	1080	
Leu Asp Gly Lys Asp Ile Asn Ile Leu Phe Asn Ser Leu Gln Tyr 1085	1090	1095	
Thr Asn Val Val Lys Asp Tyr Trp Gly Asn Asp Leu Arg Tyr Asn 1100	1105	1110	
Lys Glu Tyr Tyr Met Val Asn Ile Asp Tyr Leu Asn Arg Tyr Met 1115	1120	1125	
Tyr Ala Asn Ser Arg Gln Ile Val Phe Asn Thr Arg Arg Asn Asn 1130	1135	1140	
Asn Asp Phe Asn Glu Gly Tyr Lys Ile Ile Ile Lys Arg Ile Arg 1145	1150	1155	
Gly Asn Thr Asn Asp Thr Arg Val Arg Gly Gly Asp Ile Leu Tyr 1160	1165	1170	
Phe Asp Met Thr Ile Asn Asn Lys Ala Tyr Asn Leu Phe Met Lys 1175	1180	1185	
Asn Glu Thr Met Tyr Ala Asp Asn His Ser Thr Glu Asp Ile Tyr 1190	1195	1200	

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Ala Ile Gly Leu Arg Glu Gln Thr Lys Asp Ile Asn Asp Asn Ile  
 1205 1210 1215

Ile Phe Gln Ile Gln Pro Met Asn Asn Thr Tyr Tyr Tyr Ala Ser  
 1220 1225 1230

Gln Ile Phe Lys Ser Asn Phe Asn Gly Glu Asn Ile Ser Gly Ile  
 1235 1240 1245

Cys Ser Ile Gly Thr Tyr Arg Phe Arg Leu Gly Gly Asp Trp Tyr  
 1250 1255 1260

Arg His Asn Tyr Leu Val Pro Thr Val Lys Gln Gly Asn Tyr Ala  
 1265 1270 1275

Ser Leu Leu Glu Ser Thr Ser Thr His Trp Gly Phe Val Pro Val  
 1280 1285 1290

Ser Glu  
 1295

<210> SEQ ID NO 11  
 <211> LENGTH: 1280  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Atoxic derivative of Clostridium botulinum  
 serotype D neurotoxin

<400> SEQUENCE: 11

Met Thr Trp Pro Val Lys Asp Phe Asn Tyr Ser Asp Pro Val Asn Asp  
 1 5 10 15

Asn Asp Ile Leu Tyr Leu Arg Ile Pro Gln Asn Lys Leu Ile Thr Thr  
 20 25 30

Pro Val Lys Ala Phe Met Ile Thr Gln Asn Ile Trp Val Ile Pro Glu  
 35 40 45

Arg Phe Ser Ser Asp Thr Asn Pro Ser Leu Ser Lys Pro Pro Arg Pro  
 50 55 60

Thr Ser Lys Tyr Gln Ser Tyr Tyr Asp Pro Ser Tyr Leu Ser Thr Asp  
 65 70 75 80

Glu Gln Lys Asp Thr Phe Leu Lys Gly Ile Ile Lys Leu Phe Lys Arg  
 85 90 95

Ile Asn Glu Arg Asp Ile Gly Lys Lys Leu Ile Asn Tyr Leu Val Val  
 100 105 110

Gly Ser Pro Phe Met Gly Asp Ser Ser Thr Pro Glu Asp Thr Phe Asp  
 115 120 125

Phe Thr Arg His Thr Thr Asn Ile Ala Val Glu Lys Phe Glu Asn Gly  
 130 135 140

Ser Trp Lys Val Thr Asn Ile Ile Thr Pro Ser Val Leu Ile Phe Gly  
 145 150 155 160

Pro Leu Pro Asn Ile Leu Asp Tyr Thr Ala Ser Leu Thr Leu Gln Gly  
 165 170 175

Gln Gln Ser Asn Pro Ser Phe Glu Gly Phe Gly Thr Leu Ser Ile Leu  
 180 185 190

Lys Val Ala Pro Glu Phe Leu Leu Thr Phe Ser Asp Val Thr Ser Asn  
 195 200 205

Gln Ser Ser Ala Val Leu Gly Lys Ser Ile Phe Cys Met Asp Pro Val  
 210 215 220

Ile Ala Leu Met His Glu Leu Thr His Ser Leu His Gln Leu Tyr Gly  
 225 230 235 240

Ile Asn Ile Pro Ser Asp Lys Arg Ile Arg Pro Gln Val Ser Glu Gly

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245			250			255									
Phe	Phe	Ser	Gln	Asp	Gly	Pro	Asn	Val	Gln	Phe	Glu	Glu	Leu	Tyr	Thr
			260					265						270	
Phe	Gly	Gly	Leu	Asp	Val	Glu	Ile	Ile	Pro	Gln	Ile	Glu	Arg	Ser	Gln
			275					280						285	
Leu	Arg	Glu	Lys	Ala	Leu	Gly	His	Tyr	Lys	Asp	Ile	Ala	Lys	Arg	Leu
			290					295			300				
Asn	Asn	Ile	Asn	Lys	Thr	Ile	Pro	Ser	Ser	Trp	Ile	Ser	Asn	Ile	Asp
			305					310			315				320
Lys	Tyr	Lys	Lys	Ile	Phe	Ser	Glu	Lys	Tyr	Asn	Phe	Asp	Lys	Asp	Asn
					325						330				335
Thr	Gly	Asn	Phe	Val	Val	Asn	Ile	Asp	Lys	Phe	Asn	Ser	Leu	Tyr	Ser
					340						345				350
Asp	Leu	Thr	Asn	Val	Met	Ser	Glu	Val	Val	Tyr	Ser	Ser	Gln	Tyr	Asn
					355			360							365
Val	Lys	Asn	Arg	Thr	His	Tyr	Phe	Ser	Arg	His	Tyr	Leu	Pro	Val	Phe
					370						375				380
Ala	Asn	Ile	Leu	Asp	Asp	Asn	Ile	Tyr	Thr	Ile	Arg	Asp	Gly	Phe	Asn
					385						390				395
Leu	Thr	Asn	Lys	Gly	Phe	Asn	Ile	Glu	Asn	Ser	Gly	Gln	Asn	Ile	Glu
					405						410				415
Arg	Asn	Pro	Ala	Leu	Gln	Lys	Leu	Ser	Ser	Glu	Ser	Val	Val	Asp	Leu
					420						425				430
Phe	Thr	Lys	Val	Cys	Leu	Arg	Leu	Thr	Lys	Asn	Ser	Asp	Asp	Asp	Asp
					435						440				445
Lys	Asp	Asp	Ser	Thr	Cys	Ile	Lys	Val	Lys	Asn	Asn	Arg	Leu	Pro	Tyr
					450						455				460
Val	Ala	Asp	Lys	Asp	Ser	Ile	Ser	Gln	Glu	Ile	Phe	Glu	Asn	Lys	Ile
					465						470				475
Ile	Thr	Asp	Glu	Thr	Asn	Val	Gln	Asn	Tyr	Ser	Asp	Asn	Phe	Ser	Leu
					485						490				495
Asp	Glu	Ser	Ile	Leu	Asp	Gly	Gln	Val	Pro	Ile	Asn	Pro	Glu	Ile	Val
					500						505				510
Asp	Pro	Leu	Leu	Pro	Asn	Val	Asn	Met	Glu	Pro	Leu	Asn	Leu	Pro	Gly
					515						520				525
Glu	Glu	Ile	Val	Phe	Tyr	Asp	Asp	Ile	Thr	Lys	Tyr	Val	Asp	Tyr	Leu
					530						535				540
Asn	Ser	Tyr	Tyr	Tyr	Leu	Glu	Ser	Gln	Lys	Leu	Ser	Asn	Asn	Val	Glu
					545						550				555
Asn	Ile	Thr	Leu	Thr	Thr	Ser	Val	Glu	Glu	Ala	Leu	Gly	Tyr	Ser	Asn
					565						570				575
Lys	Ile	Tyr	Thr	Phe	Leu	Pro	Ser	Leu	Ala	Glu	Lys	Val	Asn	Lys	Gly
					580						585				590
Val	Gln	Ala	Gly	Leu	Phe	Leu	Asn	Trp	Ala	Asn	Glu	Val	Val	Glu	Asp
					595						600				605
Phe	Thr	Thr	Asn	Ile	Met	Lys	Lys	Asp	Thr	Leu	Asp	Lys	Ile	Ser	Asp
					610						615				620
Val	Ser	Val	Ile	Ile	Pro	Tyr	Ile	Gly	Pro	Ala	Leu	Asn	Ile	Gly	Asn
					625						630				635
Ser	Ala	Leu	Arg	Gly	Asn	Phe	Lys	Gln	Ala	Phe	Ala	Thr	Ala	Gly	Val
					645						650				655
Ala	Phe	Leu	Leu	Glu	Gly	Phe	Pro	Glu	Phe	Thr	Ile	Pro	Ala	Leu	Gly
					660						665				670

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Val Phe Thr Phe Tyr Ser Ser Ile Gln Glu Arg Glu Lys Ile Ile Lys  
 675 680 685  
 Thr Ile Glu Asn Cys Leu Glu Gln Arg Val Lys Arg Trp Lys Asp Ser  
 690 695 700  
 Tyr Gln Trp Met Val Ser Asn Trp Leu Ser Arg Ile Thr Thr Gln Phe  
 705 710 715 720  
 Asn His Ile Asn Tyr Gln Met Tyr Asp Ser Leu Ser Tyr Gln Ala Asp  
 725 730 735  
 Ala Ile Lys Ala Lys Ile Asp Leu Glu Tyr Lys Lys Tyr Ser Gly Ser  
 740 745 750  
 Asp Lys Glu Asn Ile Lys Ser Gln Val Glu Asn Leu Lys Asn Ser Leu  
 755 760 765  
 Asp Val Lys Ile Ser Glu Ala Met Asn Asn Ile Asn Lys Phe Ile Arg  
 770 775 780  
 Glu Cys Ser Val Thr Tyr Leu Phe Lys Asn Met Leu Pro Lys Val Ile  
 785 790 795 800  
 Asp Glu Leu Asn Lys Phe Asp Leu Arg Thr Lys Thr Glu Leu Ile Asn  
 805 810 815  
 Leu Ile Asp Ser His Asn Ile Ile Leu Val Gly Glu Val Asp Arg Leu  
 820 825 830  
 Lys Ala Lys Val Asn Glu Ser Phe Glu Asn Thr Met Pro Phe Asn Ile  
 835 840 845  
 Phe Ser Tyr Thr Asn Asn Ser Leu Leu Lys Asp Ile Ile Asn Glu Tyr  
 850 855 860  
 Phe Asn Ser Ile Asn Asp Ser Lys Ile Leu Ser Leu Gln Asn Lys Lys  
 865 870 875 880  
 Asn Ala Leu Val Asp Thr Ser Gly Tyr Asn Ala Glu Val Arg Val Gly  
 885 890 895  
 Asp Asn Val Gln Leu Asn Thr Ile Tyr Thr Asn Asp Phe Lys Leu Ser  
 900 905 910  
 Ser Ser Gly Asp Lys Ile Ile Val Asn Leu Asn Asn Asn Ile Leu Tyr  
 915 920 925  
 Ser Ala Ile Tyr Glu Asn Ser Ser Val Ser Phe Trp Ile Lys Ile Ser  
 930 935 940  
 Lys Asp Leu Thr Asn Ser His Asn Glu Tyr Thr Ile Ile Asn Ser Ile  
 945 950 955 960  
 Glu Gln Asn Ser Gly Trp Lys Leu Cys Ile Arg Asn Gly Asn Ile Glu  
 965 970 975  
 Trp Ile Leu Gln Asp Val Asn Arg Lys Tyr Lys Ser Leu Ile Phe Asp  
 980 985 990  
 Tyr Ser Glu Ser Leu Ser His Thr Gly Tyr Thr Asn Lys Trp Phe Phe  
 995 1000 1005  
 Val Thr Ile Thr Asn Asn Ile Met Gly Tyr Met Lys Leu Tyr Ile  
 1010 1015 1020  
 Asn Gly Glu Leu Lys Gln Ser Gln Lys Ile Glu Asp Leu Asp Glu  
 1025 1030 1035  
 Val Lys Leu Asp Lys Thr Ile Val Phe Gly Ile Asp Glu Asn Ile  
 1040 1045 1050  
 Asp Glu Asn Gln Met Leu Trp Ile Arg Asp Phe Asn Ile Phe Ser  
 1055 1060 1065  
 Lys Glu Leu Ser Asn Glu Asp Ile Asn Ile Val Tyr Glu Gly Gln  
 1070 1075 1080

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Ile Leu Arg Asn Val Ile Lys Asp Tyr Trp Gly Asn Pro Leu Lys
 1085                1090                1095

Phe Asp Thr Glu Tyr Tyr Ile Ile Asn Asp Asn Tyr Ile Asp Arg
 1100                1105                1110

Tyr Ile Ala Pro Glu Ser Asn Val Leu Val Leu Val Arg Tyr Pro
 1115                1120                1125

Asp Arg Ser Lys Leu Tyr Thr Gly Asn Pro Ile Thr Ile Lys Ser
 1130                1135                1140

Val Ser Asp Lys Asn Pro Tyr Ser Arg Ile Leu Asn Gly Asp Asn
 1145                1150                1155

Ile Ile Leu His Met Leu Tyr Asn Ser Arg Lys Tyr Met Ile Ile
 1160                1165                1170

Arg Asp Thr Asp Thr Ile Tyr Ala Thr Gln Gly Gly Glu Cys Ser
 1175                1180                1185

Gln Asn Cys Val Tyr Ala Leu Lys Leu Gln Ser Asn Leu Gly Asn
 1190                1195                1200

Tyr Gly Ile Gly Ile Phe Ser Ile Lys Asn Ile Val Ser Lys Asn
 1205                1210                1215

Lys Tyr Cys Ser Gln Ile Phe Ser Ser Phe Arg Glu Asn Thr Met
 1220                1225                1230

Leu Leu Ala Asp Ile Tyr Lys Pro Trp Arg Phe Ser Phe Lys Asn
 1235                1240                1245

Ala Tyr Thr Pro Val Ala Val Thr Asn Tyr Glu Thr Lys Leu Leu
 1250                1255                1260

Ser Thr Ser Ser Phe Trp Lys Phe Ile Ser Arg Asp Pro Gly Trp
 1265                1270                1275

Val Glu
 1280

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<210> SEQ ID NO 12
<211> LENGTH: 1255
<212> TYPE: PRT
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Atoxic derivative of Clostridium botulinum
        serotype E neurotoxin

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<400> SEQUENCE: 12

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Met Pro Lys Ile Asn Ser Phe Asn Tyr Asn Asp Pro Val Asn Asp Arg
 1          5          10          15

Thr Ile Leu Tyr Ile Lys Pro Gly Gly Cys Gln Glu Phe Tyr Lys Ser
 20        25        30

Phe Asn Ile Met Lys Asn Ile Trp Ile Ile Pro Glu Arg Asn Val Ile
 35        40        45

Gly Thr Thr Pro Gln Asp Phe His Pro Pro Thr Ser Leu Lys Asn Gly
 50        55        60

Asp Ser Ser Tyr Tyr Asp Pro Asn Tyr Leu Gln Ser Asp Glu Glu Lys
 65        70        75        80

Asp Arg Phe Leu Lys Ile Val Thr Lys Ile Phe Asn Arg Ile Asn Asn
 85        90        95

Asn Leu Ser Gly Gly Ile Leu Leu Glu Glu Leu Ser Lys Ala Asn Pro
 100       105       110

Tyr Leu Gly Asn Asp Asn Thr Pro Asp Asn Gln Phe His Ile Gly Asp
 115       120       125

Ala Ser Ala Val Glu Ile Lys Phe Ser Asn Gly Ser Gln Asp Ile Leu
 130       135       140

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Leu Pro Asn Val Ile Ile Met Gly Ala Glu Pro Asp Leu Phe Glu Thr  
 145 150 155 160  
 Asn Ser Ser Asn Ile Ser Leu Arg Asn Asn Tyr Met Pro Ser Asn His  
 165 170 175  
 Gly Phe Gly Ser Ile Ala Ile Val Thr Phe Ser Pro Glu Tyr Ser Phe  
 180 185 190  
 Arg Phe Asn Asp Asn Ser Met Asn Glu Phe Ile Gln Asp Pro Ala Leu  
 195 200 205  
 Thr Leu Met His Glu Leu Ile His Ser Leu His Gly Leu Tyr Gly Ala  
 210 215 220  
 Lys Gly Ile Thr Thr Lys Tyr Thr Ile Thr Gln Lys Gln Asn Pro Leu  
 225 230 235 240  
 Ile Thr Asn Ile Arg Gly Thr Asn Ile Glu Glu Phe Leu Thr Phe Gly  
 245 250 255  
 Gly Thr Asp Leu Asn Ile Ile Thr Ser Ala Gln Ser Asn Asp Ile Tyr  
 260 265 270  
 Thr Asn Leu Leu Ala Asp Tyr Lys Lys Ile Ala Ser Lys Leu Ser Lys  
 275 280 285  
 Val Gln Val Ser Asn Pro Leu Leu Asn Pro Tyr Lys Asp Val Phe Glu  
 290 295 300  
 Ala Lys Tyr Gly Leu Asp Lys Asp Ala Ser Gly Ile Tyr Ser Val Asn  
 305 310 315 320  
 Ile Asn Lys Phe Asn Asp Ile Phe Lys Lys Leu Tyr Ser Phe Thr Glu  
 325 330 335  
 Phe Asp Leu Ala Thr Lys Phe Gln Val Lys Cys Arg Gln Thr Tyr Ile  
 340 345 350  
 Gly Gln Tyr Lys Tyr Phe Lys Leu Ser Asn Leu Leu Asn Asp Ser Ile  
 355 360 365  
 Tyr Asn Ile Ser Glu Gly Tyr Asn Ile Asn Asn Leu Lys Val Asn Phe  
 370 375 380  
 Arg Gly Gln Asn Ala Asn Leu Asn Pro Arg Ile Ile Thr Pro Ile Thr  
 385 390 395 400  
 Gly Arg Gly Leu Val Lys Lys Ile Ile Arg Phe Cys Lys Asn Ile Val  
 405 410 415  
 Ser Val Lys Gly Ile Asp Asp Asp Asp Lys Lys Ser Ile Cys Ile Glu  
 420 425 430  
 Ile Asn Asn Gly Glu Leu Phe Phe Val Ala Ser Glu Asn Ser Tyr Asn  
 435 440 445  
 Asp Asp Asn Ile Asn Thr Pro Lys Glu Ile Asp Asp Thr Val Thr Ser  
 450 455 460  
 Asn Asn Asn Tyr Glu Asn Asp Leu Asp Gln Val Ile Leu Asn Phe Asn  
 465 470 475 480  
 Ser Glu Ser Ala Pro Gly Leu Ser Asp Glu Lys Leu Asn Leu Thr Ile  
 485 490 495  
 Gln Asn Asp Ala Tyr Ile Pro Lys Tyr Asp Ser Asn Gly Thr Ser Asp  
 500 505 510  
 Ile Glu Gln His Asp Val Asn Glu Leu Asn Val Phe Phe Tyr Leu Asp  
 515 520 525  
 Ala Gln Lys Val Pro Glu Gly Glu Asn Asn Val Asn Leu Thr Ser Ser  
 530 535 540  
 Ile Asp Thr Ala Leu Leu Glu Gln Pro Lys Ile Tyr Thr Phe Phe Ser  
 545 550 555 560



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Ser Glu Phe Ile Asn Asn Val Asn Lys Pro Val Gln Ala Ala Leu Phe  
 565 570 575  
 Val Ser Trp Ile Gln Gln Val Leu Val Asp Phe Thr Thr Glu Ala Asn  
 580 585 590  
 Gln Lys Ser Thr Val Asp Lys Ile Ala Asp Ile Ser Ile Val Val Pro  
 595 600 605  
 Tyr Ile Gly Leu Ala Leu Asn Ile Gly Asn Glu Ala Gln Lys Gly Asn  
 610 615 620  
 Phe Lys Asp Ala Leu Glu Leu Leu Gly Ala Gly Ile Leu Leu Glu Phe  
 625 630 635 640  
 Glu Pro Glu Leu Leu Ile Pro Thr Ile Leu Val Phe Thr Ile Lys Ser  
 645 650 655  
 Phe Leu Gly Ser Ser Asp Asn Lys Asn Lys Val Ile Lys Ala Ile Asn  
 660 665 670  
 Asn Ala Leu Lys Glu Arg Asp Glu Lys Trp Lys Glu Val Tyr Ser Phe  
 675 680 685  
 Ile Val Ser Asn Trp Met Thr Lys Ile Asn Thr Gln Phe Asn Lys Arg  
 690 695 700  
 Lys Glu Gln Met Tyr Gln Ala Leu Gln Asn Gln Val Asn Ala Ile Lys  
 705 710 715 720  
 Thr Ile Ile Glu Ser Lys Tyr Asn Ser Tyr Thr Leu Glu Glu Lys Asn  
 725 730 735  
 Glu Leu Thr Asn Lys Tyr Asp Ile Lys Gln Ile Glu Asn Glu Leu Asn  
 740 745 750  
 Gln Lys Val Ser Ile Ala Met Asn Asn Ile Asp Arg Phe Leu Thr Glu  
 755 760 765  
 Ser Ser Ile Ser Tyr Leu Met Lys Leu Ile Asn Glu Val Lys Ile Asn  
 770 775 780  
 Lys Leu Arg Glu Tyr Asp Glu Asn Val Lys Thr Tyr Leu Leu Asn Tyr  
 785 790 795 800  
 Ile Ile Gln His Gly Ser Ile Leu Gly Glu Ser Gln Gln Glu Leu Asn  
 805 810 815  
 Ser Met Val Thr Asp Thr Leu Asn Asn Ser Ile Pro Phe Lys Leu Ser  
 820 825 830  
 Ser Tyr Thr Asp Asp Lys Ile Leu Ile Ser Tyr Phe Asn Lys Phe Phe  
 835 840 845  
 Asn Asn Ile Lys Ser Ser Ser Val Leu Asn Met Arg Tyr Lys Asn Asp  
 850 855 860  
 Lys Tyr Val Asp Thr Ser Gly Tyr Asp Ser Asn Ile Asn Ile Asn Gly  
 865 870 875 880  
 Asp Val Tyr Lys Tyr Pro Thr Asn Lys Asn Gln Phe Gly Ile Tyr Asn  
 885 890 895  
 Asp Lys Leu Ser Glu Val Asn Ile Ser Gln Asn Asp Tyr Ile Ile Tyr  
 900 905 910  
 Asp Asn Lys Tyr Lys Asn Phe Ser Ile Ser Phe Trp Val Arg Ile Pro  
 915 920 925  
 Asn Tyr Asp Asn Lys Ile Val Asn Val Asn Asn Glu Tyr Thr Ile Ile  
 930 935 940  
 Asn Cys Met Arg Asp Asn Asn Ser Gly Trp Lys Val Ser Leu Asn His  
 945 950 955 960  
 Asn Glu Ile Ile Trp Thr Leu Gln Asp Asn Ala Gly Ile Asn Gln Lys  
 965 970 975  
 Leu Ala Phe Asn Tyr Gly Asn Ala Asn Gly Ile Ser Asp Tyr Ile Asn

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980				985				990							
Lys	Trp	Ile	Phe	Val	Thr	Ile	Thr	Asn	Asp	Arg	Leu	Gly	Asp	Ser	Lys
		995					1000					1005			
Leu	Tyr	Ile	Asn	Gly	Asn	Leu	Ile	Asp	Gln	Lys	Ser	Ile	Leu	Asn	
	1010					1015					1020				
Leu	Gly	Asn	Ile	His	Val	Ser	Asp	Asn	Ile	Leu	Phe	Lys	Ile	Val	
	1025					1030					1035				
Asn	Cys	Ser	Tyr	Thr	Arg	Tyr	Ile	Gly	Ile	Arg	Tyr	Phe	Asn	Ile	
	1040					1045					1050				
Phe	Asp	Lys	Glu	Leu	Asp	Glu	Thr	Glu	Ile	Gln	Thr	Leu	Tyr	Ser	
	1055					1060					1065				
Asn	Glu	Pro	Asn	Thr	Asn	Ile	Leu	Lys	Asp	Phe	Trp	Gly	Asn	Tyr	
	1070					1075					1080				
Leu	Leu	Tyr	Asp	Lys	Glu	Tyr	Tyr	Leu	Leu	Asn	Val	Leu	Lys	Pro	
	1085					1090					1095				
Asn	Asn	Phe	Ile	Asp	Arg	Arg	Lys	Asp	Ser	Thr	Leu	Ser	Ile	Asn	
	1100					1105					1110				
Asn	Ile	Arg	Ser	Thr	Ile	Leu	Leu	Ala	Asn	Arg	Leu	Tyr	Ser	Gly	
	1115					1120					1125				
Ile	Lys	Val	Lys	Ile	Gln	Arg	Val	Asn	Asn	Ser	Ser	Thr	Asn	Asp	
	1130					1135					1140				
Asn	Leu	Val	Arg	Lys	Asn	Asp	Gln	Val	Tyr	Ile	Asn	Phe	Val	Ala	
	1145					1150					1155				
Ser	Lys	Thr	His	Leu	Phe	Pro	Leu	Tyr	Ala	Asp	Thr	Ala	Thr	Thr	
	1160					1165					1170				
Asn	Lys	Glu	Lys	Thr	Ile	Lys	Ile	Ser	Ser	Ser	Gly	Asn	Arg	Phe	
	1175					1180					1185				
Asn	Gln	Val	Val	Val	Met	Asn	Ser	Val	Gly	Asn	Asn	Thr	Met	Asn	
	1190					1195					1200				
Phe	Lys	Asn	Asn	Asn	Gly	Asn	Asn	Ile	Gly	Leu	Leu	Gly	Phe	Lys	
	1205					1210					1215				
Ala	Asp	Thr	Val	Val	Ala	Ser	Thr	Trp	Tyr	Tyr	Thr	His	Met	Arg	
	1220					1225					1230				
Asp	His	Thr	Asn	Ser	Asn	Gly	Cys	Phe	Trp	Asn	Phe	Ile	Ser	Glu	
	1235					1240					1245				
Glu	His	Gly	Trp	Gln	Glu	Lys									
	1250					1255									

&lt;210&gt; SEQ ID NO 13

&lt;211&gt; LENGTH: 1281

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Atoxic derivative of Clostridium botulinum serotype F neurotoxin

&lt;400&gt; SEQUENCE: 13

Met Pro Val Val Ile Asn Ser Phe Asn Tyr Asn Asp Pro Val Asn Asp  
 1 5 10 15

Asp Thr Ile Leu Tyr Met Gln Ile Pro Tyr Glu Glu Lys Ser Lys Lys  
 20 25 30

Tyr Tyr Lys Ala Phe Glu Ile Met Arg Asn Val Trp Ile Ile Pro Glu  
 35 40 45

Arg Asn Thr Ile Gly Thr Asp Pro Ser Asp Phe Asp Pro Pro Ala Ser  
 50 55 60

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Leu Glu Asn Gly Ser Ser Ala Tyr Tyr Asp Pro Asn Tyr Leu Thr Thr  
 65 70 75 80  
 Asp Ala Glu Lys Asp Arg Tyr Leu Lys Thr Thr Ile Lys Leu Phe Lys  
 85 90 95  
 Arg Ile Asn Ser Asn Pro Ala Gly Glu Val Leu Leu Gln Glu Ile Ser  
 100 105 110  
 Tyr Ala Lys Pro Tyr Leu Gly Asn Glu His Thr Pro Ile Asn Glu Phe  
 115 120 125  
 His Pro Val Thr Arg Thr Thr Ser Val Asn Ile Lys Ser Ser Thr Asn  
 130 135 140  
 Val Lys Ser Ser Ile Ile Leu Asn Leu Leu Val Leu Gly Ala Gly Pro  
 145 150 155 160  
 Asp Ile Phe Glu Asn Ser Ser Tyr Pro Val Arg Lys Leu Met Asp Ser  
 165 170 175  
 Gly Gly Val Tyr Asp Pro Ser Asn Asp Gly Phe Gly Ser Ile Asn Ile  
 180 185 190  
 Val Thr Phe Ser Pro Glu Tyr Glu Tyr Thr Phe Asn Asp Ile Ser Gly  
 195 200 205  
 Gly Tyr Asn Ser Ser Thr Glu Ser Phe Ile Ala Asp Pro Ala Ile Ser  
 210 215 220  
 Leu Ala His Glu Leu Ile His Ala Leu His Gly Leu Tyr Gly Ala Arg  
 225 230 235 240  
 Gly Val Thr Tyr Lys Glu Thr Ile Lys Val Lys Gln Ala Pro Leu Met  
 245 250 255  
 Ile Ala Ile Lys Pro Ile Arg Leu Glu Glu Phe Leu Thr Phe Gly Gly  
 260 265 270  
 Gln Asp Leu Asn Ile Ile Thr Ser Ala Met Lys Glu Lys Ile Tyr Asn  
 275 280 285  
 Asn Leu Leu Ala Asn Tyr Glu Lys Ile Ala Thr Arg Leu Ser Arg Val  
 290 295 300  
 Asn Ser Ala Pro Pro Glu Tyr Asp Ile Asn Glu Tyr Lys Asp Tyr Phe  
 305 310 315 320  
 Gln Trp Lys Tyr Gly Leu Asp Lys Asn Ala Asp Gly Ser Tyr Thr Val  
 325 330 335  
 Asn Glu Asn Lys Phe Asn Glu Ile Tyr Lys Lys Leu Tyr Ser Phe Thr  
 340 345 350  
 Glu Ile Asp Leu Ala Asn Lys Phe Lys Val Lys Cys Arg Asn Thr Tyr  
 355 360 365  
 Phe Ile Lys Tyr Gly Phe Leu Lys Val Pro Asn Leu Leu Asp Asp Asp  
 370 375 380  
 Ile Tyr Thr Val Ser Glu Gly Phe Asn Ile Gly Asn Leu Ala Val Asn  
 385 390 395 400  
 Asn Arg Gly Gln Asn Ile Lys Leu Asn Pro Lys Ile Ile Asp Ser Ile  
 405 410 415  
 Pro Asp Lys Gly Leu Val Glu Lys Ile Val Lys Phe Cys Lys Ser Val  
 420 425 430  
 Ile Pro Arg Lys Gly Thr Asp Asp Asp Lys Ala Pro Pro Arg Leu  
 435 440 445  
 Cys Ile Arg Val Asn Asn Arg Glu Leu Phe Phe Val Ala Ser Glu Ser  
 450 455 460  
 Ser Tyr Asn Glu Asn Asp Ile Asn Thr Pro Lys Glu Ile Asp Asp Thr  
 465 470 475 480  
 Thr Asn Leu Asn Asn Asn Tyr Arg Asn Asn Leu Asp Glu Val Ile Leu

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485					490					495					
Asp	Tyr	Asn	Ser	Glu	Thr	Ile	Pro	Gln	Ile	Ser	Asn	Gln	Thr	Leu	Asn
			500					505					510		
Thr	Leu	Val	Gln	Asp	Asp	Ser	Tyr	Val	Pro	Arg	Tyr	Asp	Ser	Asn	Gly
		515					520					525			
Thr	Ser	Glu	Ile	Glu	Glu	His	Asn	Val	Val	Asp	Leu	Asn	Val	Phe	Phe
	530					535					540				
Tyr	Leu	His	Ala	Gln	Lys	Val	Pro	Glu	Gly	Glu	Thr	Asn	Ile	Ser	Leu
545					550					555					560
Thr	Ser	Ser	Ile	Asp	Thr	Ala	Leu	Ser	Glu	Glu	Ser	Gln	Val	Tyr	Thr
			565						570					575	
Phe	Phe	Ser	Ser	Glu	Phe	Ile	Asn	Thr	Ile	Asn	Lys	Pro	Val	His	Ala
		580						585					590		
Ala	Leu	Phe	Ile	Ser	Trp	Ile	Asn	Gln	Val	Ile	Arg	Asp	Phe	Thr	Thr
		595					600					605			
Glu	Ala	Thr	Gln	Lys	Ser	Thr	Phe	Asp	Lys	Ile	Ala	Asp	Ile	Ser	Leu
610						615					620				
Val	Val	Pro	Tyr	Val	Gly	Leu	Ala	Leu	Asn	Ile	Gly	Asn	Glu	Val	Gln
625					630					635					640
Lys	Glu	Asn	Phe	Lys	Glu	Ala	Phe	Glu	Leu	Leu	Gly	Ala	Gly	Ile	Leu
			645						650					655	
Leu	Glu	Phe	Val	Pro	Glu	Leu	Leu	Ile	Pro	Thr	Ile	Leu	Val	Phe	Thr
		660						665					670		
Ile	Lys	Ser	Phe	Ile	Gly	Ser	Ser	Glu	Asn	Lys	Asn	Lys	Ile	Ile	Lys
	675						680					685			
Ala	Ile	Asn	Asn	Ser	Leu	Met	Glu	Arg	Glu	Thr	Lys	Trp	Lys	Glu	Ile
	690					695						700			
Tyr	Ser	Trp	Ile	Val	Ser	Asn	Trp	Leu	Thr	Arg	Ile	Asn	Thr	Gln	Phe
705					710					715					720
Asn	Lys	Arg	Lys	Glu	Gln	Met	Tyr	Gln	Ala	Leu	Gln	Asn	Gln	Val	Asp
			725						730					735	
Ala	Ile	Lys	Thr	Val	Ile	Glu	Tyr	Lys	Tyr	Asn	Asn	Tyr	Thr	Ser	Asp
		740						745					750		
Glu	Arg	Asn	Arg	Leu	Glu	Ser	Glu	Tyr	Asn	Ile	Asn	Asn	Ile	Arg	Glu
	755						760					765			
Glu	Leu	Asn	Lys	Lys	Val	Ser	Leu	Ala	Met	Glu	Asn	Ile	Glu	Arg	Phe
770						775						780			
Ile	Thr	Glu	Ser	Ser	Ile	Phe	Tyr	Leu	Met	Lys	Leu	Ile	Asn	Glu	Ala
785						790					795				800
Lys	Val	Ser	Lys	Leu	Arg	Glu	Tyr	Asp	Glu	Gly	Val	Lys	Glu	Tyr	Leu
			805						810					815	
Leu	Asp	Tyr	Ile	Ser	Glu	His	Arg	Ser	Ile	Leu	Gly	Asn	Ser	Val	Gln
		820						825					830		
Glu	Leu	Asn	Asp	Leu	Val	Thr	Ser	Thr	Leu	Asn	Asn	Ser	Ile	Pro	Phe
		835						840				845			
Glu	Leu	Ser	Ser	Tyr	Thr	Asn	Asp	Lys	Ile	Leu	Ile	Leu	Tyr	Phe	Asn
	850					855					860				
Lys	Leu	Tyr	Asn	Asn	Ile	Lys	Asp	Asn	Ser	Ile	Leu	Asp	Met	Arg	Tyr
865						870					875				880
Glu	Asn	Asn	Lys	Phe	Ile	Asp	Ile	Ser	Gly	Tyr	Gly	Ser	Asn	Ile	Ser
			885						890					895	
Ile	Asn	Gly	Asp	Val	Tyr	Ile	Tyr	Ser	Thr	Asn	Arg	Asn	Gln	Phe	Gly
		900						905					910		

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Ile Tyr Ser Ser Lys Pro Ser Glu Val Asn Ile Ala Gln Asn Asn Asp
    915                      920                      925

Ile Ile Tyr Asn Gly Arg Tyr Gln Asn Phe Ser Ile Ser Phe Trp Val
    930                      935                      940

Arg Ile Pro Lys Tyr Phe Asn Lys Val Asn Leu Asn Asn Glu Tyr Thr
    945                      950                      955                      960

Ile Ile Asp Cys Ile Arg Asn Asn Asn Ser Gly Trp Lys Ile Ser Leu
    965                      970                      975

Asn Tyr Asn Lys Ile Ile Trp Thr Leu Gln Asp Thr Ala Gly Asn Asn
    980                      985                      990

Gln Lys Leu Val Phe Asn Tyr Thr Gln Met Ile Ser Ile Ser Asp Tyr
    995                      1000                     1005

Ile Asn Lys Trp Ile Phe Val Thr Ile Thr Asn Asn Arg Leu Gly
    1010                      1015                     1020

Asn Ser Arg Ile Tyr Ile Asn Gly Asn Leu Ile Asp Glu Lys Ser
    1025                      1030                     1035

Ile Ser Asn Leu Gly Asp Ile His Val Ser Asp Asn Ile Leu Phe
    1040                      1045                     1050

Lys Ile Val Gly Cys Asn Asp Thr Arg Tyr Val Gly Ile Arg Tyr
    1055                      1060                     1065

Phe Lys Val Phe Asp Thr Glu Leu Gly Lys Thr Glu Ile Glu Thr
    1070                      1075                     1080

Leu Tyr Ser Asp Glu Pro Asp Pro Ser Ile Leu Lys Asp Phe Trp
    1085                      1090                     1095

Gly Asn Tyr Leu Leu Tyr Asn Lys Arg Tyr Tyr Leu Leu Asn Leu
    1100                      1105                     1110

Leu Arg Thr Asp Lys Ser Ile Thr Gln Asn Ser Asn Phe Leu Asn
    1115                      1120                     1125

Ile Asn Gln Gln Arg Gly Val Tyr Gln Lys Pro Asn Ile Phe Ser
    1130                      1135                     1140

Asn Thr Arg Leu Tyr Thr Gly Val Glu Val Ile Ile Arg Lys Asn
    1145                      1150                     1155

Gly Ser Thr Asp Ile Ser Asn Thr Asp Asn Phe Val Arg Lys Asn
    1160                      1165                     1170

Asp Leu Ala Tyr Ile Asn Val Val Asp Arg Asp Val Glu Tyr Arg
    1175                      1180                     1185

Leu Tyr Ala Asp Ile Ser Ile Ala Lys Pro Glu Lys Ile Ile Lys
    1190                      1195                     1200

Leu Ile Arg Thr Ser Asn Ser Asn Asn Ser Leu Gly Gln Ile Ile
    1205                      1210                     1215

Val Met Asp Ser Ile Gly Asn Asn Thr Met Asn Phe Gln Asn Asn
    1220                      1225                     1230

Asn Gly Gly Asn Ile Gly Leu Leu Gly Phe His Ser Asn Asn Leu
    1235                      1240                     1245

Val Ala Ser Ser Trp Tyr Tyr Asn Asn Ile Arg Lys Asn Thr Ser
    1250                      1255                     1260

Ser Asn Gly Cys Phe Trp Ser Phe Ile Ser Lys Glu His Gly Trp
    1265                      1270                     1275

Gln Glu Asn
    1280

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&lt;210&gt; SEQ ID NO 14

&lt;211&gt; LENGTH: 1301

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<212> TYPE: PRT  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Atoxic derivative of Clostridium botulinum  
 serotype G neurotoxin

<400> SEQUENCE: 14

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Met Pro Val Asn Ile Lys Asn Phe Asn Tyr Asn Asp Pro Ile Asn Asn
1           5           10           15
Asp Asp Ile Ile Met Met Glu Pro Phe Asn Asp Pro Gly Pro Gly Thr
20           25           30
Tyr Tyr Lys Ala Phe Arg Ile Ile Asp Arg Ile Trp Ile Val Pro Glu
35           40           45
Arg Phe Thr Tyr Gly Phe Gln Pro Asp Gln Phe Asn Ala Ser Thr Gly
50           55           60
Val Phe Ser Lys Asp Val Tyr Glu Tyr Tyr Asp Pro Thr Tyr Leu Lys
65           70           75           80
Thr Asp Ala Glu Lys Asp Lys Phe Leu Lys Thr Met Ile Lys Leu Phe
85           90           95
Asn Arg Ile Asn Ser Lys Pro Ser Gly Gln Arg Leu Leu Asp Met Ile
100          105          110
Val Asp Ala Ile Pro Tyr Leu Gly Asn Ala Ser Thr Pro Pro Asp Lys
115          120          125
Phe Ala Ala Asn Val Ala Asn Val Ser Ile Asn Lys Lys Ile Ile Gln
130          135          140
Pro Gly Ala Glu Asp Gln Ile Lys Gly Leu Met Thr Asn Leu Ile Ile
145          150          155          160
Phe Gly Pro Gly Pro Val Leu Ser Asp Asn Phe Thr Asp Ser Met Ile
165          170          175
Met Asn Gly His Ser Pro Ile Ser Glu Gly Phe Gly Ala Arg Met Met
180          185          190
Ile Arg Phe Cys Pro Ser Cys Leu Asn Val Phe Asn Asn Val Gln Glu
195          200          205
Asn Lys Asp Thr Ser Ile Phe Ser Arg Arg Ala Tyr Phe Ala Asp Pro
210          215          220
Ala Leu Thr Leu Met His Glu Leu Ile His Val Leu His Gly Leu Tyr
225          230          235          240
Gly Ile Lys Ile Ser Asn Leu Pro Ile Thr Pro Asn Thr Lys Glu Phe
245          250          255
Phe Met Gln His Ser Asp Pro Val Gln Ala Glu Glu Leu Tyr Thr Phe
260          265          270
Gly Gly His Asp Pro Ser Val Ile Ser Pro Ser Thr Asp Met Asn Ile
275          280          285
Tyr Asn Lys Ala Leu Gln Asn Phe Gln Asp Ile Ala Asn Arg Leu Asn
290          295          300
Ile Val Ser Ser Ala Gln Gly Ser Gly Ile Asp Ile Ser Leu Tyr Lys
305          310          315          320
Gln Ile Tyr Lys Asn Lys Tyr Asp Phe Val Glu Asp Pro Asn Gly Lys
325          330          335
Tyr Ser Val Asp Lys Asp Lys Phe Asp Lys Leu Tyr Lys Ala Leu Met
340          345          350
Phe Gly Phe Thr Glu Thr Asn Leu Ala Gly Glu Tyr Gly Ile Lys Thr
355          360          365
Arg Tyr Ser Tyr Phe Ser Glu Tyr Leu Pro Pro Ile Lys Thr Glu Lys
370          375          380

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Leu Leu Asp Asn Thr Ile Tyr Thr Gln Asn Glu Gly Phe Asn Ile Ala  
 385 390 395 400  
 Ser Lys Asn Leu Lys Thr Glu Phe Asn Gly Gln Asn Lys Ala Val Asn  
 405 410 415  
 Lys Glu Ala Tyr Glu Glu Ile Ser Leu Glu His Leu Val Ile Tyr Arg  
 420 425 430  
 Ile Ala Met Cys Lys Pro Val Met Tyr Lys Asn Thr Gly Asp Asp Asp  
 435 440 445  
 Asp Lys Ser Glu Gln Cys Ile Ile Val Asn Asn Glu Asp Leu Phe Phe  
 450 455 460  
 Ile Ala Asn Lys Asp Ser Phe Ser Lys Asp Leu Ala Lys Ala Glu Thr  
 465 470 475 480  
 Ile Ala Tyr Asn Thr Gln Asn Asn Thr Ile Glu Asn Asn Phe Ser Ile  
 485 490 495  
 Asp Gln Leu Ile Leu Asp Asn Asp Leu Ser Ser Gly Ile Asp Leu Pro  
 500 505 510  
 Asn Glu Asn Thr Glu Pro Phe Thr Asn Phe Asp Asp Ile Asp Ile Pro  
 515 520 525  
 Val Tyr Ile Lys Gln Ser Ala Leu Lys Lys Ile Phe Val Asp Gly Asp  
 530 535 540  
 Ser Leu Phe Glu Tyr Leu His Ala Gln Thr Phe Pro Ser Asn Ile Glu  
 545 550 555 560  
 Asn Leu Gln Leu Thr Asn Ser Leu Asn Asp Ala Leu Arg Asn Asn Asn  
 565 570 575  
 Lys Val Tyr Thr Phe Phe Ser Thr Asn Leu Val Glu Lys Ala Asn Thr  
 580 585 590  
 Val Val Gly Ala Ser Leu Phe Val Asn Trp Val Lys Gly Val Ile Asp  
 595 600 605  
 Asp Phe Thr Ser Glu Ser Thr Gln Lys Ser Thr Ile Asp Lys Val Ser  
 610 615 620  
 Asp Val Ser Ile Ile Ile Pro Tyr Ile Gly Pro Ala Leu Asn Val Gly  
 625 630 635 640  
 Asn Glu Thr Ala Lys Glu Asn Phe Lys Asn Ala Phe Glu Ile Gly Gly  
 645 650 655  
 Ala Ala Ile Leu Met Glu Phe Ile Pro Glu Leu Ile Val Pro Ile Val  
 660 665 670  
 Gly Phe Phe Thr Leu Glu Ser Tyr Val Gly Asn Lys Gly His Ile Ile  
 675 680 685  
 Met Thr Ile Ser Asn Ala Leu Lys Lys Arg Asp Gln Lys Trp Thr Asp  
 690 695 700  
 Met Tyr Gly Leu Ile Val Ser Gln Trp Leu Ser Thr Val Asn Thr Gln  
 705 710 715 720  
 Phe Tyr Thr Ile Lys Glu Arg Met Tyr Asn Ala Leu Asn Asn Gln Ser  
 725 730 735  
 Gln Ala Ile Glu Lys Ile Ile Glu Asp Gln Tyr Asn Arg Tyr Ser Glu  
 740 745 750  
 Glu Asp Lys Met Asn Ile Asn Ile Asp Phe Asn Asp Ile Asp Phe Lys  
 755 760 765  
 Leu Asn Gln Ser Ile Asn Leu Ala Ile Asn Asn Ile Asp Asp Phe Ile  
 770 775 780  
 Asn Gln Cys Ser Ile Ser Tyr Leu Met Asn Arg Met Ile Pro Leu Ala  
 785 790 795 800

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Val Lys Lys Leu Lys Asp Phe Asp Asp Asn Leu Lys Arg Asp Leu Leu  
 805 810 815  
 Glu Tyr Ile Asp Thr Asn Glu Leu Tyr Leu Leu Asp Glu Val Asn Ile  
 820 825 830  
 Leu Lys Ser Lys Val Asn Arg His Leu Lys Asp Ser Ile Pro Phe Asp  
 835 840 845  
 Leu Ser Leu Tyr Thr Lys Asp Thr Ile Leu Ile Gln Val Phe Asn Asn  
 850 855 860  
 Tyr Ile Ser Asn Ile Ser Ser Asn Ala Ile Leu Ser Leu Ser Tyr Arg  
 865 870 875 880  
 Gly Gly Arg Leu Ile Asp Ser Ser Gly Tyr Gly Ala Thr Met Asn Val  
 885 890 895  
 Gly Ser Asp Val Ile Phe Asn Asp Ile Gly Asn Gly Gln Phe Lys Leu  
 900 905 910  
 Asn Asn Ser Glu Asn Ser Asn Ile Thr Ala His Gln Ser Lys Phe Val  
 915 920 925  
 Val Tyr Asp Ser Met Phe Asp Asn Phe Ser Ile Asn Phe Trp Val Arg  
 930 935 940  
 Thr Pro Lys Tyr Asn Asn Asn Asp Ile Gln Thr Tyr Leu Gln Asn Glu  
 945 950 955 960  
 Tyr Thr Ile Ile Ser Cys Ile Lys Asn Asp Ser Gly Trp Lys Val Ser  
 965 970 975  
 Ile Lys Gly Asn Arg Ile Ile Trp Thr Leu Ile Asp Val Asn Ala Lys  
 980 985 990  
 Ser Lys Ser Ile Phe Phe Glu Tyr Ser Ile Lys Asp Asn Ile Ser Asp  
 995 1000 1005  
 Tyr Ile Asn Lys Trp Phe Ser Ile Thr Ile Thr Asn Asp Arg Leu  
 1010 1015 1020  
 Gly Asn Ala Asn Ile Tyr Ile Asn Gly Ser Leu Lys Lys Ser Glu  
 1025 1030 1035  
 Lys Ile Leu Asn Leu Asp Arg Ile Asn Ser Ser Asn Asp Ile Asp  
 1040 1045 1050  
 Phe Lys Leu Ile Asn Cys Thr Asp Thr Thr Lys Phe Val Trp Ile  
 1055 1060 1065  
 Lys Asp Phe Asn Ile Phe Gly Arg Glu Leu Asn Ala Thr Glu Val  
 1070 1075 1080  
 Ser Ser Leu Tyr Trp Ile Gln Ser Ser Thr Asn Thr Leu Lys Asp  
 1085 1090 1095  
 Phe Trp Gly Asn Pro Leu Arg Tyr Asp Thr Gln Tyr Tyr Leu Phe  
 1100 1105 1110  
 Asn Gln Gly Met Gln Asn Ile Tyr Ile Lys Tyr Phe Ser Lys Ala  
 1115 1120 1125  
 Ser Met Gly Glu Thr Ala Pro Arg Thr Asn Phe Asn Asn Ala Ala  
 1130 1135 1140  
 Ile Asn Tyr Gln Asn Leu Tyr Leu Gly Leu Arg Phe Ile Ile Lys  
 1145 1150 1155  
 Lys Ala Ser Asn Ser Arg Asn Ile Asn Asn Asp Asn Ile Val Arg  
 1160 1165 1170  
 Glu Gly Asp Tyr Ile Tyr Leu Asn Ile Asp Asn Ile Ser Asp Glu  
 1175 1180 1185  
 Ser Tyr Arg Val Tyr Val Leu Val Asn Ser Lys Glu Ile Gln Thr  
 1190 1195 1200  
 Gln Leu Phe Leu Ala Pro Ile Asn Asp Asp Pro Thr Phe Tyr Asp



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1205	1210	1215
Val Leu Gln Ile Lys Lys Tyr Tyr Glu Lys Thr Thr Tyr Asn Cys 1220 1225 1230		
Gln Ile Leu Cys Glu Lys Asp Thr Lys Thr Phe Gly Leu Phe Gly 1235 1240 1245		
Ile Gly Lys Phe Val Lys Asp Tyr Gly Tyr Val Trp Asp Thr Tyr 1250 1255 1260		
Asp Asn Tyr Phe Cys Ile Ser Gln Trp Tyr Leu Arg Arg Ile Ser 1265 1270 1275		
Glu Asn Ile Asn Lys Leu Arg Leu Gly Cys Asn Trp Gln Phe Ile 1280 1285 1290		
Pro Val Asp Glu Gly Trp Thr Glu 1295 1300		

<210> SEQ ID NO 15  
 <211> LENGTH: 1300  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
 chimera 1

<400> SEQUENCE: 15

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly 1 5 10 15
Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro 20 25 30
Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg 35 40 45
Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu 50 55 60
Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr 65 70 75 80
Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp Glu Gln Gly 85 90 95
Glu Gln Leu Glu Arg Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly 100 105 110
Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys 115 120 125
Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr 130 135 140
Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile 145 150 155 160
Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr 165 170 175
Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe 180 185 190
Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu 195 200 205
Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Ala 210 215 220
Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn 225 230 235 240
Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu 245 250 255



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675					680					685					
Ile	Ala	Asn	Lys	Val	Leu	Thr	Val	Gln	Thr	Ile	Asp	Asn	Ala	Leu	Ser
690					695					700					
Lys	Arg	Asn	Glu	Lys	Trp	Asp	Glu	Val	Tyr	Lys	Tyr	Ile	Val	Thr	Asn
705					710					715					720
Trp	Leu	Ala	Lys	Val	Asn	Thr	Gln	Ile	Asp	Leu	Ile	Arg	Lys	Lys	Met
				725					730					735	
Lys	Glu	Ala	Leu	Glu	Asn	Gln	Ala	Glu	Ala	Thr	Lys	Ala	Ile	Ile	Asn
			740					745						750	
Tyr	Gln	Tyr	Asn	Gln	Tyr	Thr	Glu	Glu	Glu	Lys	Asn	Asn	Ile	Asn	Phe
		755					760					765			
Asn	Ile	Asp	Asp	Leu	Ser	Ser	Lys	Leu	Asn	Glu	Ser	Ile	Asn	Lys	Ala
		770					775					780			
Met	Ile	Asn	Ile	Asn	Lys	Phe	Leu	Asn	Gln	Cys	Ser	Val	Ser	Tyr	Leu
785					790					795					800
Met	Asn	Ser	Met	Ile	Pro	Tyr	Gly	Val	Lys	Arg	Leu	Glu	Asp	Phe	Asp
				805					810					815	
Ala	Ser	Leu	Lys	Asp	Ala	Leu	Leu	Lys	Tyr	Ile	Tyr	Asp	Asn	Arg	Gly
			820					825						830	
Thr	Leu	Ile	Gly	Gln	Val	Asp	Arg	Leu	Lys	Asp	Lys	Val	Asn	Asn	Thr
		835					840					845			
Leu	Ser	Thr	Asp	Ile	Pro	Phe	Gln	Leu	Ser	Lys	Tyr	Val	Asp	Asn	Gln
		850					855					860			
Arg	Leu	Leu	Ser	Thr	Phe	Thr	Glu	Tyr	Ile	Lys	Asn	Ile	Ile	Asn	Thr
865					870					875					880
Ser	Ile	Leu	Asn	Leu	Arg	Tyr	Glu	Ser	Asn	His	Leu	Ile	Asp	Leu	Ser
				885					890					895	
Arg	Tyr	Ala	Ser	Lys	Ile	Asn	Ile	Gly	Ser	Lys	Val	Asn	Phe	Asp	Pro
			900					905						910	
Ile	Asp	Lys	Asn	Gln	Ile	Gln	Leu	Phe	Asn	Leu	Glu	Ser	Ser	Lys	Ile
		915					920					925			
Glu	Val	Ile	Leu	Lys	Asn	Ala	Ile	Val	Tyr	Asn	Ser	Met	Tyr	Glu	Asn
		930					935					940			
Phe	Ser	Thr	Ser	Phe	Trp	Ile	Arg	Ile	Pro	Lys	Tyr	Phe	Asn	Ser	Ile
945					950					955					960
Ser	Leu	Asn	Asn	Glu	Tyr	Thr	Ile	Ile	Asn	Cys	Met	Glu	Asn	Asn	Ser
				965					970					975	
Gly	Trp	Lys	Val	Ser	Leu	Asn	Tyr	Gly	Glu	Ile	Ile	Trp	Thr	Leu	Gln
			980					985						990	
Asp	Thr	Gln	Glu	Ile	Lys	Gln	Arg	Val	Val	Phe	Lys	Tyr	Ser	Gln	Met
		995					1000							1005	
Ile	Asn	Ile	Ser	Asp	Tyr	Ile	Asn	Arg	Trp	Ile	Phe	Val	Thr	Ile	
		1010					1015							1020	
Thr	Asn	Asn	Arg	Leu	Asn	Asn	Ser	Lys	Ile	Tyr	Ile	Asn	Gly	Arg	
				1025			1030							1035	
Leu	Ile	Asp	Gln	Lys	Pro	Ile	Ser	Asn	Leu	Gly	Asn	Ile	His	Ala	
				1040			1045							1050	
Ser	Asn	Asn	Ile	Met	Phe	Lys	Leu	Asp	Gly	Cys	Arg	Asp	Thr	His	
				1055			1060							1065	
Arg	Tyr	Ile	Trp	Ile	Lys	Tyr	Phe	Asn	Leu	Phe	Asp	Lys	Glu	Leu	
				1070			1075							1080	
Asn	Glu	Lys	Glu	Ile	Lys	Asp	Leu	Tyr	Asp	Asn	Gln	Ser	Asn	Ser	
				1085			1090							1095	

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Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp Lys  
 1100 1105 1110  
 Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val Asp  
 1115 1120 1125  
 Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly Pro  
 1130 1135 1140  
 Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser Leu  
 1145 1150 1155  
 Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly Asn  
 1160 1165 1170  
 Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn Val  
 1175 1180 1185  
 Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser Gln  
 1190 1195 1200  
 Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp Val  
 1205 1210 1215  
 Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp Gln  
 1220 1225 1230  
 Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn Gly  
 1235 1240 1245  
 Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile Ala  
 1250 1255 1260  
 Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser  
 1265 1270 1275  
 Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp  
 1280 1285 1290  
 Gly Trp Gly Glu Arg Pro Leu  
 1295 1300

&lt;210&gt; SEQ ID NO 16

&lt;211&gt; LENGTH: 1300

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
chimera 2

&lt;400&gt; SEQUENCE: 16

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly  
 1 5 10 15  
 Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro  
 20 25 30  
 Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg  
 35 40 45  
 Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu  
 50 55 60  
 Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr  
 65 70 75 80  
 Asp Asn Glu Lys Asp Asn Tyr Leu Lys Gly Val Thr Lys Leu Phe Glu  
 85 90 95  
 Arg Ile Tyr Ser Thr Asp Leu Gly Arg Met Leu Leu Thr Ser Ile Val  
 100 105 110  
 Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys  
 115 120 125  
 Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr





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Gly Trp Lys Val Ser Leu Asn Tyr Gly Glu Ile Ile Trp Thr Leu Gln  
980 985 990

Asp Thr Gln Glu Ile Lys Gln Arg Val Val Phe Lys Tyr Ser Gln Met  
995 1000 1005

Ile Asn Ile Ser Asp Tyr Ile Asn Arg Trp Ile Phe Val Thr Ile  
1010 1015 1020

Thr Asn Asn Arg Leu Asn Asn Ser Lys Ile Tyr Ile Asn Gly Arg  
1025 1030 1035

Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His Ala  
1040 1045 1050

Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr His  
1055 1060 1065

Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu Leu  
1070 1075 1080

Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn Ser  
1085 1090 1095

Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp Lys  
1100 1105 1110

Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val Asp  
1115 1120 1125

Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly Pro  
1130 1135 1140

Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser Leu  
1145 1150 1155

Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly Asn  
1160 1165 1170

Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn Val  
1175 1180 1185

Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser Gln  
1190 1195 1200

Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp Val  
1205 1210 1215

Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp Gln  
1220 1225 1230

Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn Gly  
1235 1240 1245

Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile Ala  
1250 1255 1260

Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser  
1265 1270 1275

Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp  
1280 1285 1290

Gly Trp Gly Glu Arg Pro Leu  
1295 1300

&lt;210&gt; SEQ ID NO 17

&lt;211&gt; LENGTH: 1300

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
chimera 3

&lt;400&gt; SEQUENCE: 17

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly  
1 5 10 15





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Gly Ile Ile Thr Ser His Thr Gln Ser Leu Asp Gln Gly Tyr Asn Asp  
 435 440 445  
 Asp Asp Asp Lys Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn Trp  
 450 455 460  
 Asp Leu Phe Phe Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu Asn  
 465 470 475 480  
 Lys Gly Glu Glu Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu Glu  
 485 490 495  
 Asn Ile Ser Leu Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn Phe  
 500 505 510  
 Asp Asn Glu Pro Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp Ile  
 515 520 525  
 Ile Gly Gln Leu Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn Gly  
 530 535 540  
 Lys Lys Tyr Glu Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg Ala  
 545 550 555 560  
 Gln Glu Phe Glu His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser Val  
 565 570 575  
 Asn Glu Ala Leu Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser Ser  
 580 585 590  
 Asp Tyr Val Lys Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe Leu  
 595 600 605  
 Gly Trp Val Glu Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser Glu  
 610 615 620  
 Val Ser Thr Thr Asp Lys Ile Ala Asp Ile Thr Ile Ile Ile Pro Tyr  
 625 630 635 640  
 Ile Gly Pro Ala Leu Asn Ile Gly Asn Met Leu Tyr Lys Asp Asp Phe  
 645 650 655  
 Val Gly Ala Leu Ile Phe Ser Gly Ala Val Ile Leu Leu Glu Phe Ile  
 660 665 670  
 Pro Glu Ile Ala Ile Pro Val Leu Gly Thr Phe Ala Leu Val Ser Tyr  
 675 680 685  
 Ile Ala Asn Lys Val Leu Thr Val Gln Thr Ile Asp Asn Ala Leu Ser  
 690 695 700  
 Lys Arg Asn Glu Lys Trp Asp Glu Val Tyr Lys Tyr Ile Val Thr Asn  
 705 710 715 720  
 Trp Leu Ala Lys Val Asn Thr Gln Ile Asp Leu Ile Arg Lys Lys Met  
 725 730 735  
 Lys Glu Ala Leu Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile Asn  
 740 745 750  
 Tyr Gln Tyr Asn Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn Phe  
 755 760 765  
 Asn Ile Asp Asp Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys Ala  
 770 775 780  
 Met Ile Asn Ile Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr Leu  
 785 790 795 800  
 Met Asn Ser Met Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe Asp  
 805 810 815  
 Ala Ser Leu Lys Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg Gly  
 820 825 830  
 Thr Leu Ile Gly Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn Thr  
 835 840 845  
 Leu Ser Thr Asp Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn Gln

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850		855		860											
Arg	Leu	Leu	Ser	Thr	Phe	Thr	Glu	Tyr	Ile	Lys	Asn	Ile	Ile	Asn	Thr
865					870					875					880
Ser	Ile	Leu	Asn	Leu	Arg	Tyr	Glu	Ser	Asn	His	Leu	Ile	Asp	Leu	Ser
			885						890						895
Arg	Tyr	Ala	Ser	Lys	Ile	Asn	Ile	Gly	Ser	Lys	Val	Asn	Phe	Asp	Pro
			900					905						910	
Ile	Asp	Lys	Asn	Gln	Ile	Gln	Leu	Phe	Asn	Leu	Glu	Ser	Ser	Lys	Ile
		915					920							925	
Glu	Val	Ile	Leu	Lys	Asn	Ala	Ile	Val	Tyr	Asn	Ser	Met	Tyr	Glu	Asn
		930				935								940	
Phe	Ser	Thr	Ser	Phe	Trp	Ile	Arg	Ile	Pro	Lys	Tyr	Phe	Asn	Ser	Ile
945					950					955					960
Ser	Leu	Asn	Asn	Glu	Tyr	Thr	Ile	Ile	Asn	Cys	Met	Glu	Asn	Asn	Ser
				965						970					975
Gly	Trp	Lys	Val	Ser	Leu	Asn	Tyr	Gly	Glu	Ile	Ile	Trp	Thr	Leu	Gln
			980						985						990
Asp	Thr	Gln	Glu	Ile	Lys	Gln	Arg	Val	Val	Phe	Lys	Tyr	Ser	Gln	Met
		995					1000							1005	
Ile	Asn	Ile	Ser	Asp	Tyr	Ile	Asn	Arg	Trp	Ile	Phe	Val	Thr	Ile	
	1010						1015						1020		
Thr	Asn	Asn	Arg	Leu	Asn	Asn	Ser	Lys	Ile	Tyr	Ile	Asn	Gly	Arg	
	1025						1030						1035		
Leu	Ile	Asp	Gln	Lys	Pro	Ile	Ser	Asn	Leu	Gly	Asn	Ile	His	Ala	
	1040						1045						1050		
Ser	Asn	Asn	Ile	Met	Phe	Lys	Leu	Asp	Gly	Cys	Arg	Asp	Thr	His	
	1055					1060							1065		
Arg	Tyr	Ile	Trp	Ile	Lys	Tyr	Phe	Asn	Leu	Phe	Asp	Lys	Glu	Leu	
	1070					1075							1080		
Asn	Glu	Lys	Glu	Ile	Lys	Asp	Leu	Tyr	Asp	Asn	Gln	Ser	Asn	Ser	
	1085					1090							1095		
Gly	Ile	Leu	Lys	Asp	Phe	Trp	Gly	Asp	Tyr	Leu	Gln	Tyr	Asp	Lys	
	1100					1105							1110		
Pro	Tyr	Tyr	Met	Leu	Asn	Leu	Tyr	Asp	Pro	Asn	Lys	Tyr	Val	Asp	
	1115					1120							1125		
Val	Asn	Asn	Val	Gly	Ile	Arg	Gly	Tyr	Met	Tyr	Leu	Lys	Gly	Pro	
	1130					1135							1140		
Arg	Gly	Ser	Val	Met	Thr	Thr	Asn	Ile	Tyr	Leu	Asn	Ser	Ser	Leu	
	1145					1150							1155		
Tyr	Arg	Gly	Thr	Lys	Phe	Ile	Ile	Lys	Lys	Tyr	Ala	Ser	Gly	Asn	
	1160					1165							1170		
Lys	Asp	Asn	Ile	Val	Arg	Asn	Asn	Asp	Arg	Val	Tyr	Ile	Asn	Val	
	1175					1180							1185		
Val	Val	Lys	Asn	Lys	Glu	Tyr	Arg	Leu	Ala	Thr	Asn	Ala	Ser	Gln	
	1190					1195							1200		
Ala	Gly	Val	Glu	Lys	Ile	Leu	Ser	Ala	Leu	Glu	Ile	Pro	Asp	Val	
	1205					1210							1215		
Gly	Asn	Leu	Ser	Gln	Val	Val	Val	Met	Lys	Ser	Lys	Asn	Asp	Gln	
	1220					1225							1230		
Gly	Ile	Thr	Asn	Lys	Cys	Lys	Met	Asn	Leu	Gln	Asp	Asn	Asn	Gly	
	1235					1240							1245		
Asn	Asp	Ile	Gly	Phe	Ile	Gly	Phe	His	Gln	Phe	Asn	Asn	Ile	Ala	
	1250					1255							1260		

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Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser  
1265 1270 1275

Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp  
1280 1285 1290

Gly Trp Gly Glu Arg Pro Leu  
1295 1300

<210> SEQ ID NO 18

<211> LENGTH: 1300

<212> TYPE: PRT

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A, chimera 4

<400> SEQUENCE: 18

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly  
1 5 10 15

Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro  
20 25 30

Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg  
35 40 45

Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu  
50 55 60

Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr  
65 70 75 80

Asp Asn Glu Lys Asp Asn Tyr Leu Lys Gly Val Thr Lys Leu Phe Glu  
85 90 95

Arg Ile Tyr Ser Thr Asp Leu Gly Arg Met Leu Leu Thr Ser Ile Val  
100 105 110

Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys  
115 120 125

Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr  
130 135 140

Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile  
145 150 155 160

Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr  
165 170 175

Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe  
180 185 190

Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu  
195 200 205

Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Ala  
210 215 220

Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn  
225 230 235 240

Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu  
245 250 255

Glu Val Ser Phe Glu Glu Leu Arg Thr Phe Gly Gly His Asp Ala Lys  
260 265 270

Phe Ile Asp Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp  
275 280 285

Glu Gln Gly Glu Gln Leu Glu Arg Leu Asn Lys Ala Lys Ser Ile Val  
290 295 300

Gly Thr Thr Ala Ser Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly  
305 310 315 320



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Lys Glu Ala Leu Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile Asn  
 740 745 750  
 Tyr Gln Tyr Asn Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn Phe  
 755 760 765  
 Asn Ile Asp Asp Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys Ala  
 770 775 780  
 Met Ile Asn Ile Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr Leu  
 785 790 795 800  
 Met Asn Ser Met Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe Asp  
 805 810 815  
 Ala Ser Leu Lys Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg Gly  
 820 825 830  
 Thr Leu Ile Gly Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn Thr  
 835 840 845  
 Leu Ser Thr Asp Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn Gln  
 850 855 860  
 Arg Leu Leu Ser Thr Phe Thr Glu Tyr Ile Lys Asn Ile Ile Asn Thr  
 865 870 875 880  
 Ser Ile Leu Asn Leu Arg Tyr Glu Ser Asn His Leu Ile Asp Leu Ser  
 885 890 895  
 Arg Tyr Ala Ser Lys Ile Asn Ile Gly Ser Lys Val Asn Phe Asp Pro  
 900 905 910  
 Ile Asp Lys Asn Gln Ile Gln Leu Phe Asn Leu Glu Ser Ser Lys Ile  
 915 920 925  
 Glu Val Ile Leu Lys Asn Ala Ile Val Tyr Asn Ser Met Tyr Glu Asn  
 930 935 940  
 Phe Ser Thr Ser Phe Trp Ile Arg Ile Pro Lys Tyr Phe Asn Ser Ile  
 945 950 955 960  
 Ser Leu Asn Asn Glu Tyr Thr Ile Ile Asn Cys Met Glu Asn Asn Ser  
 965 970 975  
 Gly Trp Lys Val Ser Leu Asn Tyr Gly Glu Ile Ile Trp Thr Leu Gln  
 980 985 990  
 Asp Thr Gln Glu Ile Lys Gln Arg Val Val Phe Lys Tyr Ser Gln Met  
 995 1000 1005  
 Ile Asn Ile Ser Asp Tyr Ile Asn Arg Trp Ile Phe Val Thr Ile  
 1010 1015 1020  
 Thr Asn Asn Arg Leu Asn Asn Ser Lys Ile Tyr Ile Asn Gly Arg  
 1025 1030 1035  
 Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His Ala  
 1040 1045 1050  
 Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr His  
 1055 1060 1065  
 Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu Leu  
 1070 1075 1080  
 Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn Ser  
 1085 1090 1095  
 Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp Lys  
 1100 1105 1110  
 Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val Asp  
 1115 1120 1125  
 Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly Pro  
 1130 1135 1140  
 Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser Leu

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1145	1150	1155
Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly Asn		
1160	1165	1170
Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn Val		
1175	1180	1185
Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser Gln		
1190	1195	1200
Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp Val		
1205	1210	1215
Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp Gln		
1220	1225	1230
Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn Gly		
1235	1240	1245
Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile Ala		
1250	1255	1260
Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser		
1265	1270	1275
Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp		
1280	1285	1290
Gly Trp Gly Glu Arg Pro Leu		
1295	1300	

&lt;210&gt; SEQ ID NO 19

&lt;211&gt; LENGTH: 1300

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
chimera 5

&lt;400&gt; SEQUENCE: 19

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly			
1	5	10	15
Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro			
	20	25	30
Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg			
	35	40	45
Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu			
	50	55	60
Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr			
	65	70	75
Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp Glu Gln Gly			
	85	90	95
Glu Gln Leu Glu Arg Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly			
	100	105	110
Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys			
	115	120	125
Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr			
	130	135	140
Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile			
	145	150	155
Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr			
	165	170	175
Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe			
	180	185	190

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Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu  
 195 200 205  
 Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Ala  
 210 215 220  
 Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn  
 225 230 235 240  
 Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu  
 245 250 255  
 Glu Val Ser Phe Glu Glu Leu Arg Thr Phe Gly Gly His Asp Ala Lys  
 260 265 270  
 Phe Ile Asp Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp  
 275 280 285  
 Glu Gln Gly Glu Gln Leu Glu Arg Leu Asn Lys Ala Lys Ser Ile Val  
 290 295 300  
 Gly Thr Thr Ala Ser Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly  
 305 310 315 320  
 Arg Leu Leu Ser Glu Asp Thr Ser Gly Lys Phe Ser Val Asp Lys Leu  
 325 330 335  
 Lys Phe Asp Lys Leu Tyr Lys Met Leu Thr Glu Ile Tyr Thr Glu Asp  
 340 345 350  
 Asn Phe Val Lys Phe Phe Lys Val Leu Asn Arg Lys Thr Tyr Leu Asn  
 355 360 365  
 Phe Asp Lys Ala Val Phe Lys Ile Asn Ile Val Pro Lys Val Asn Tyr  
 370 375 380  
 Thr Ile Tyr Asp Gly Phe Asn Leu Arg Asn Thr Asn Leu Ala Ala Asn  
 385 390 395 400  
 Phe Asn Gly Gln Asn Thr Glu Ile Asn Asn Met Asn Phe Thr Lys Leu  
 405 410 415  
 Lys Asn Phe Thr Gly Leu Phe Glu Phe Tyr Lys Leu Leu Cys Val Arg  
 420 425 430  
 Gly Ile Ile Thr Ser His Thr Gln Ser Leu Asp Gln Gly Tyr Asn Asp  
 435 440 445  
 Asp Asp Asp Lys Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn Trp  
 450 455 460  
 Asp Leu Phe Phe Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu Asn  
 465 470 475 480  
 Lys Gly Glu Glu Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu Glu  
 485 490 495  
 Asn Ile Ser Leu Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn Phe  
 500 505 510  
 Asp Asn Glu Pro Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp Ile  
 515 520 525  
 Ile Gly Gln Leu Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn Gly  
 530 535 540  
 Lys Lys Tyr Glu Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg Ala  
 545 550 555 560  
 Gln Glu Phe Glu His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser Val  
 565 570 575  
 Asn Glu Ala Leu Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser Ser  
 580 585 590  
 Asp Tyr Val Lys Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe Leu  
 595 600 605  
 Gly Trp Val Glu Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser Glu

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610				615				620							
Val	Ser	Thr	Thr	Asp	Lys	Ile	Ala	Asp	Ile	Thr	Ile	Ile	Ile	Pro	Tyr
625					630					635				640	
Ile	Gly	Pro	Ala	Leu	Asn	Ile	Gly	Asn	Met	Leu	Tyr	Lys	Asp	Asp	Phe
				645					650					655	
Val	Gly	Ala	Leu	Ile	Phe	Ser	Gly	Ala	Val	Ile	Leu	Leu	Glu	Phe	Ile
			660					665					670		
Pro	Glu	Ile	Ala	Ile	Pro	Val	Leu	Gly	Thr	Phe	Ala	Leu	Val	Ser	Tyr
		675					680					685			
Ile	Ala	Asn	Lys	Val	Leu	Thr	Val	Gln	Thr	Ile	Asp	Asn	Ala	Leu	Ser
	690					695					700				
Lys	Arg	Asn	Glu	Lys	Trp	Asp	Glu	Val	Tyr	Lys	Tyr	Ile	Val	Thr	Asn
705					710					715					720
Trp	Leu	Ala	Lys	Val	Asn	Thr	Gln	Ile	Asp	Leu	Ile	Arg	Lys	Lys	Met
			725						730					735	
Lys	Glu	Ala	Leu	Glu	Asn	Gln	Ala	Glu	Ala	Thr	Lys	Ala	Ile	Ile	Asn
			740					745					750		
Tyr	Gln	Tyr	Asn	Gln	Tyr	Thr	Glu	Glu	Glu	Lys	Asn	Asn	Ile	Asn	Phe
		755					760					765			
Asn	Ile	Asp	Asp	Leu	Ser	Ser	Lys	Leu	Asn	Glu	Ser	Ile	Asn	Lys	Ala
	770					775					780				
Met	Ile	Asn	Ile	Asn	Lys	Phe	Leu	Asn	Gln	Cys	Ser	Val	Ser	Tyr	Leu
785					790					795					800
Met	Asn	Ser	Met	Ile	Pro	Tyr	Gly	Val	Lys	Arg	Leu	Glu	Asp	Phe	Asp
				805					810					815	
Ala	Ser	Leu	Lys	Asp	Ala	Leu	Leu	Lys	Tyr	Ile	Tyr	Asp	Asn	Arg	Gly
			820					825					830		
Thr	Leu	Ile	Gly	Gln	Val	Asp	Arg	Leu	Lys	Asp	Lys	Val	Asn	Asn	Thr
		835					840						845		
Leu	Ser	Thr	Asp	Ile	Pro	Phe	Gln	Leu	Ser	Lys	Tyr	Val	Asp	Asn	Gln
	850					855					860				
Arg	Leu	Leu	Ser	Thr	Phe	Thr	Glu	Tyr	Ile	Lys	Asn	Ile	Ile	Asn	Thr
865					870					875					880
Ser	Ile	Leu	Asn	Leu	Arg	Tyr	Glu	Ser	Asn	His	Leu	Ile	Asp	Leu	Ser
				885					890					895	
Arg	Tyr	Ala	Ser	Lys	Ile	Asn	Ile	Gly	Ser	Lys	Val	Asn	Phe	Asp	Pro
		900						905					910		
Ile	Asp	Lys	Asn	Gln	Ile	Gln	Leu	Phe	Asn	Leu	Glu	Ser	Ser	Lys	Ile
		915					920						925		
Glu	Val	Ile	Leu	Lys	Asn	Ala	Ile	Val	Tyr	Asn	Ser	Met	Tyr	Glu	Asn
	930					935						940			
Phe	Ser	Thr	Ser	Phe	Trp	Ile	Arg	Ile	Pro	Lys	Tyr	Phe	Asn	Ser	Ile
945					950					955					960
Ser	Leu	Asn	Asn	Glu	Tyr	Thr	Ile	Ile	Asn	Cys	Met	Glu	Asn	Asn	Ser
				965					970					975	
Gly	Trp	Lys	Val	Ser	Leu	Asn	Tyr	Gly	Glu	Ile	Ile	Trp	Thr	Leu	Gln
			980					985					990		
Asp	Thr	Gln	Glu	Ile	Lys	Gln	Arg	Val	Val	Phe	Lys	Tyr	Ser	Gln	Met
		995					1000						1005		
Ile	Asn	Ile	Ser	Asp	Tyr	Ile	Asn	Arg	Trp	Ile	Phe	Val	Thr	Ile	
	1010						1015					1020			
Thr	Asn	Asn	Arg	Leu	Asn	Asn	Ser	Lys	Ile	Tyr	Ile	Asn	Gly	Arg	
	1025						1030					1035			



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Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His Ala  
 1040 1045 1050  
 Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr His  
 1055 1060 1065  
 Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu Leu  
 1070 1075 1080  
 Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn Ser  
 1085 1090 1095  
 Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp Lys  
 1100 1105 1110  
 Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val Asp  
 1115 1120 1125  
 Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly Pro  
 1130 1135 1140  
 Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser Leu  
 1145 1150 1155  
 Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly Asn  
 1160 1165 1170  
 Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn Val  
 1175 1180 1185  
 Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser Gln  
 1190 1195 1200  
 Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp Val  
 1205 1210 1215  
 Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp Gln  
 1220 1225 1230  
 Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn Gly  
 1235 1240 1245  
 Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile Ala  
 1250 1255 1260  
 Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg Ser  
 1265 1270 1275  
 Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp Asp  
 1280 1285 1290  
 Gly Trp Gly Glu Arg Pro Leu  
 1295 1300

&lt;210&gt; SEQ ID NO 20

&lt;211&gt; LENGTH: 1301

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
chimera 6

&lt;400&gt; SEQUENCE: 20

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly  
 1 5 10 15  
 Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro  
 20 25 30  
 Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg  
 35 40 45  
 Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu  
 50 55 60  
 Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr

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65	70	75	80
Asp Asn Glu Lys Asp Asn Tyr Leu Lys Gly Val Thr Lys Leu Phe Glu 85 90 95			
Arg Ile Tyr Ser Thr Asp Leu Gly Arg Met Leu Leu Thr Ser Ile Val 100 105 110			
Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys 115 120 125			
Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr 130 135 140			
Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile 145 150 155 160			
Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr 165 170 175			
Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe 180 185 190			
Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu 195 200 205			
Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Ala 210 215 220			
Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn 225 230 235 240			
Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu 245 250 255			
Glu Val Ser Phe Glu Glu Leu Arg Thr Phe Gly Gly His Asp Ala Lys 260 265 270			
Phe Ile Asp Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp 275 280 285			
Glu Gln Gly Glu Gln Leu Glu Arg Leu Asn Lys Ala Lys Ser Ile Val 290 295 300			
Gly Thr Thr Ala Ser Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly 305 310 315 320			
Arg Leu Leu Ser Glu Asp Thr Ser Gly Lys Phe Ser Val Asp Phe Met 325 330 335			
Asp Glu Phe Phe Glu Gln Val Glu Glu Leu Thr Glu Ile Tyr Thr Glu 340 345 350			
Asp Asn Phe Val Lys Phe Phe Lys Val Leu Asn Arg Lys Thr Tyr Leu 355 360 365			
Asn Phe Asp Lys Ala Val Phe Lys Ile Asn Ile Val Pro Lys Val Asn 370 375 380			
Tyr Thr Ile Tyr Asp Gly Phe Asn Leu Arg Asn Thr Asn Leu Ala Ala 385 390 395 400			
Asn Phe Asn Gly Gln Asn Thr Glu Ile Asn Asn Met Asn Phe Thr Lys 405 410 415			
Leu Lys Asn Phe Thr Gly Leu Phe Glu Phe Tyr Lys Leu Leu Cys Val 420 425 430			
Arg Gly Ile Ile Thr Ser His Thr Gln Ser Leu Asp Gln Gly Tyr Asn 435 440 445			
Asp Asp Asp Asp Lys Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn 450 455 460			
Trp Asp Leu Phe Phe Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu 465 470 475 480			
Asn Lys Gly Glu Glu Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu 485 490 495			

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Glu Asn Ile Ser Leu Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn  
 500 505 510  
 Phe Asp Asn Glu Pro Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp  
 515 520 525  
 Ile Ile Gly Gln Leu Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn  
 530 535 540  
 Gly Lys Lys Tyr Glu Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg  
 545 550 555 560  
 Ala Gln Glu Phe Glu His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser  
 565 570 575  
 Val Asn Glu Ala Leu Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser  
 580 585 590  
 Ser Asp Tyr Val Lys Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe  
 595 600 605  
 Leu Gly Trp Val Glu Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser  
 610 615 620  
 Glu Val Ser Thr Thr Asp Lys Ile Ala Asp Ile Thr Ile Ile Ile Pro  
 625 630 635 640  
 Tyr Ile Gly Pro Ala Leu Asn Ile Gly Asn Met Leu Tyr Lys Asp Asp  
 645 650 655  
 Phe Val Gly Ala Leu Ile Phe Ser Gly Ala Val Ile Leu Leu Glu Phe  
 660 665 670  
 Ile Pro Glu Ile Ala Ile Pro Val Leu Gly Thr Phe Ala Leu Val Ser  
 675 680 685  
 Tyr Ile Ala Asn Lys Val Leu Thr Val Gln Thr Ile Asp Asn Ala Leu  
 690 695 700  
 Ser Lys Arg Asn Glu Lys Trp Asp Glu Val Tyr Lys Tyr Ile Val Thr  
 705 710 715 720  
 Asn Trp Leu Ala Lys Val Asn Thr Gln Ile Asp Leu Ile Arg Lys Lys  
 725 730 735  
 Met Lys Glu Ala Leu Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile  
 740 745 750  
 Asn Tyr Gln Tyr Asn Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn  
 755 760 765  
 Phe Asn Ile Asp Asp Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys  
 770 775 780  
 Ala Met Ile Asn Ile Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr  
 785 790 795 800  
 Leu Met Asn Ser Met Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe  
 805 810 815  
 Asp Ala Ser Leu Lys Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg  
 820 825 830  
 Gly Thr Leu Ile Gly Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn  
 835 840 845  
 Thr Leu Ser Thr Asp Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn  
 850 855 860  
 Gln Arg Leu Leu Ser Thr Phe Thr Glu Tyr Ile Lys Asn Ile Ile Asn  
 865 870 875 880  
 Thr Ser Ile Leu Asn Leu Arg Tyr Glu Ser Asn His Leu Ile Asp Leu  
 885 890 895  
 Ser Arg Tyr Ala Ser Lys Ile Asn Ile Gly Ser Lys Val Asn Phe Asp  
 900 905 910

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Pro Ile Asp Lys Asn Gln Ile Gln Leu Phe Asn Leu Glu Ser Ser Lys  
 915 920 925  
 Ile Glu Val Ile Leu Lys Asn Ala Ile Val Tyr Asn Ser Met Tyr Glu  
 930 935 940  
 Asn Phe Ser Thr Ser Phe Trp Ile Arg Ile Pro Lys Tyr Phe Asn Ser  
 945 950 955 960  
 Ile Ser Leu Asn Asn Glu Tyr Thr Ile Ile Asn Cys Met Glu Asn Asn  
 965 970 975  
 Ser Gly Trp Lys Val Ser Leu Asn Tyr Gly Glu Ile Ile Trp Thr Leu  
 980 985 990  
 Gln Asp Thr Gln Glu Ile Lys Gln Arg Val Val Phe Lys Tyr Ser Gln  
 995 1000 1005  
 Met Ile Asn Ile Ser Asp Tyr Ile Asn Arg Trp Ile Phe Val Thr  
 1010 1015 1020  
 Ile Thr Asn Asn Arg Leu Asn Asn Ser Lys Ile Tyr Ile Asn Gly  
 1025 1030 1035  
 Arg Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His  
 1040 1045 1050  
 Ala Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr  
 1055 1060 1065  
 His Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu  
 1070 1075 1080  
 Leu Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn  
 1085 1090 1095  
 Ser Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp  
 1100 1105 1110  
 Lys Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val  
 1115 1120 1125  
 Asp Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly  
 1130 1135 1140  
 Pro Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser  
 1145 1150 1155  
 Leu Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly  
 1160 1165 1170  
 Asn Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn  
 1175 1180 1185  
 Val Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser  
 1190 1195 1200  
 Gln Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp  
 1205 1210 1215  
 Val Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp  
 1220 1225 1230  
 Gln Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn  
 1235 1240 1245  
 Gly Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile  
 1250 1255 1260  
 Ala Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg  
 1265 1270 1275  
 Ser Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp  
 1280 1285 1290  
 Asp Gly Trp Gly Glu Arg Pro Leu  
 1295 1300



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Asn Phe Asp Lys Ala Val Phe Lys Ile Asn Ile Val Pro Lys Val Asn  
 370 375 380

Tyr Thr Ile Tyr Asp Gly Phe Asn Leu Arg Asn Thr Asn Leu Ala Ala  
 385 390 395 400

Asn Phe Asn Gly Gln Asn Thr Glu Ile Asn Asn Met Asn Phe Thr Lys  
 405 410 415

Leu Lys Asn Phe Thr Gly Leu Phe Glu Phe Tyr Lys Leu Leu Cys Val  
 420 425 430

Arg Gly Ile Ile Thr Ser His Thr Gln Ser Leu Asp Gln Gly Tyr Asn  
 435 440 445

Asp Asp Asp Asp Lys Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn  
 450 455 460

Trp Asp Leu Phe Phe Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu  
 465 470 475 480

Asn Lys Gly Glu Glu Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu  
 485 490 495

Glu Asn Ile Ser Leu Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn  
 500 505 510

Phe Asp Asn Glu Pro Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp  
 515 520 525

Ile Ile Gly Gln Leu Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn  
 530 535 540

Gly Lys Lys Tyr Glu Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg  
 545 550 555 560

Ala Gln Glu Phe Glu His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser  
 565 570 575

Val Asn Glu Ala Leu Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser  
 580 585 590

Ser Asp Tyr Val Lys Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe  
 595 600 605

Leu Gly Trp Val Glu Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser  
 610 615 620

Glu Val Ser Thr Thr Asp Lys Ile Ala Asp Ile Thr Ile Ile Ile Pro  
 625 630 635 640

Tyr Ile Gly Pro Ala Leu Asn Ile Gly Asn Met Leu Tyr Lys Asp Asp  
 645 650 655

Phe Val Gly Ala Leu Ile Phe Ser Gly Ala Val Ile Leu Leu Glu Phe  
 660 665 670

Ile Pro Glu Ile Ala Ile Pro Val Leu Gly Thr Phe Ala Leu Val Ser  
 675 680 685

Tyr Ile Ala Asn Lys Val Leu Thr Val Gln Thr Ile Asp Asn Ala Leu  
 690 695 700

Ser Lys Arg Asn Glu Lys Trp Asp Glu Val Tyr Lys Tyr Ile Val Thr  
 705 710 715 720

Asn Trp Leu Ala Lys Val Asn Thr Gln Ile Asp Leu Ile Arg Lys Lys  
 725 730 735

Met Lys Glu Ala Leu Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile  
 740 745 750

Asn Tyr Gln Tyr Asn Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn  
 755 760 765

Phe Asn Ile Asp Asp Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys  
 770 775 780

Ala Met Ile Asn Ile Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr

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785	790	795	800
Leu Met Asn Ser Met Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe	805	810	815
Asp Ala Ser Leu Lys Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg	820	825	830
Gly Thr Leu Ile Gly Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn	835	840	845
Thr Leu Ser Thr Asp Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn	850	855	860
Gln Arg Leu Leu Ser Thr Phe Thr Glu Tyr Ile Lys Asn Ile Ile Asn	865	870	875
Thr Ser Ile Leu Asn Leu Arg Tyr Glu Ser Asn His Leu Ile Asp Leu	885	890	895
Ser Arg Tyr Ala Ser Lys Ile Asn Ile Gly Ser Lys Val Asn Phe Asp	900	905	910
Pro Ile Asp Lys Asn Gln Ile Gln Leu Phe Asn Leu Glu Ser Ser Lys	915	920	925
Ile Glu Val Ile Leu Lys Asn Ala Ile Val Tyr Asn Ser Met Tyr Glu	930	935	940
Asn Phe Ser Thr Ser Phe Trp Ile Arg Ile Pro Lys Tyr Phe Asn Ser	945	950	955
Ile Ser Leu Asn Asn Glu Tyr Thr Ile Ile Asn Cys Met Glu Asn Asn	965	970	975
Ser Gly Trp Lys Val Ser Leu Asn Tyr Gly Glu Ile Ile Trp Thr Leu	980	985	990
Gln Asp Thr Gln Glu Ile Lys Gln Arg Val Val Phe Lys Tyr Ser Gln	995	1000	1005
Met Ile Asn Ile Ser Asp Tyr Ile Asn Arg Trp Ile Phe Val Thr	1010	1015	1020
Ile Thr Asn Asn Arg Leu Asn Asn Ser Lys Ile Tyr Ile Asn Gly	1025	1030	1035
Arg Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His	1040	1045	1050
Ala Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr	1055	1060	1065
His Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu	1070	1075	1080
Leu Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn	1085	1090	1095
Ser Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp	1100	1105	1110
Lys Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val	1115	1120	1125
Asp Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly	1130	1135	1140
Pro Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser	1145	1150	1155
Leu Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly	1160	1165	1170
Asn Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn	1175	1180	1185
Val Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser	1190	1195	1200

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Gln Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp  
 1205 1210 1215

Val Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp  
 1220 1225 1230

Gln Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn  
 1235 1240 1245

Gly Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile  
 1250 1255 1260

Ala Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg  
 1265 1270 1275

Ser Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp  
 1280 1285 1290

Asp Gly Trp Gly Glu Arg Pro Leu  
 1295 1300

&lt;210&gt; SEQ ID NO 22

&lt;211&gt; LENGTH: 1301

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
 chimera 8

&lt;400&gt; SEQUENCE: 22

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly  
 1 5 10 15

Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro  
 20 25 30

Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg  
 35 40 45

Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu  
 50 55 60

Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr  
 65 70 75 80

Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp Glu Gln Gly  
 85 90 95

Glu Gln Leu Glu Arg Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly  
 100 105 110

Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys  
 115 120 125

Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr  
 130 135 140

Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile  
 145 150 155 160

Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr  
 165 170 175

Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe  
 180 185 190

Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu  
 195 200 205

Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Ala  
 210 215 220

Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn  
 225 230 235 240

Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu



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245				250				255							
Glu	Val	Ser	Phe	Glu	Glu	Leu	Arg	Thr	Phe	Gly	Gly	His	Asp	Ala	Lys
			260												270
Phe	Ile	Asp	Glu	Leu	Asp	Asp	Arg	Ala	Asp	Ala	Leu	Gln	Met	Leu	Asp
			275												285
Glu	Gln	Gly	Glu	Gln	Leu	Glu	Arg	Leu	Asn	Lys	Ala	Lys	Ser	Ile	Val
			290												300
Gly	Thr	Thr	Ala	Ser	Glu	Met	Asp	Glu	Asn	Leu	Glu	Gln	Val	Ser	Gly
															320
Arg	Leu	Leu	Ser	Glu	Asp	Thr	Ser	Gly	Lys	Phe	Ser	Val	Asp	Phe	Met
															335
Asp	Glu	Phe	Phe	Glu	Gln	Val	Glu	Glu	Leu	Thr	Glu	Ile	Tyr	Thr	Glu
															350
Asp	Asn	Phe	Val	Lys	Phe	Phe	Lys	Val	Leu	Asn	Arg	Lys	Thr	Tyr	Leu
															365
Asn	Phe	Asp	Lys	Ala	Val	Phe	Lys	Ile	Asn	Ile	Val	Pro	Lys	Val	Asn
															380
Tyr	Thr	Ile	Tyr	Asp	Gly	Phe	Asn	Leu	Arg	Asn	Thr	Asn	Leu	Ala	Ala
															400
Asn	Phe	Asn	Gly	Gln	Asn	Thr	Glu	Ile	Asn	Asn	Met	Asn	Phe	Thr	Lys
															415
Leu	Lys	Asn	Phe	Thr	Gly	Leu	Phe	Glu	Phe	Tyr	Lys	Leu	Leu	Cys	Val
															430
Arg	Gly	Ile	Ile	Thr	Ser	His	Thr	Gln	Ser	Leu	Asp	Gln	Gly	Tyr	Asn
															445
Asp	Asp	Asp	Asp	Lys	Ala	Leu	Asn	Asp	Leu	Cys	Ile	Lys	Val	Asn	Asn
															460
Trp	Asp	Leu	Phe	Phe	Ser	Pro	Ser	Glu	Asp	Asn	Phe	Thr	Asn	Asp	Leu
															480
Asn	Lys	Gly	Glu	Glu	Ile	Thr	Ser	Asp	Thr	Asn	Ile	Glu	Ala	Ala	Glu
															495
Glu	Asn	Ile	Ser	Leu	Asp	Leu	Ile	Gln	Gln	Tyr	Tyr	Leu	Thr	Phe	Asn
															510
Phe	Asp	Asn	Glu	Pro	Glu	Asn	Ile	Ser	Ile	Glu	Asn	Leu	Ser	Ser	Asp
															525
Ile	Ile	Gly	Gln	Leu	Glu	Leu	Met	Pro	Asn	Ile	Glu	Arg	Phe	Pro	Asn
															540
Gly	Lys	Lys	Tyr	Glu	Leu	Asp	Lys	Tyr	Thr	Met	Phe	His	Tyr	Leu	Arg
															560
Ala	Gln	Glu	Phe	Glu	His	Gly	Lys	Ser	Arg	Ile	Ala	Leu	Thr	Asn	Ser
															575
Val	Asn	Glu	Ala	Leu	Leu	Asn	Pro	Ser	Arg	Val	Tyr	Thr	Phe	Phe	Ser
															590
Ser	Asp	Tyr	Val	Lys	Lys	Val	Asn	Lys	Ala	Thr	Glu	Ala	Ala	Met	Phe
															605
Leu	Gly	Trp	Val	Glu	Gln	Leu	Val	Tyr	Asp	Phe	Thr	Asp	Glu	Thr	Ser
															620
Glu	Val	Ser	Thr	Thr	Asp	Lys	Ile	Ala	Asp	Ile	Thr	Ile	Ile	Ile	Pro
															640
Tyr	Ile	Gly	Pro	Ala	Leu	Asn	Ile	Gly	Asn	Met	Leu	Tyr	Lys	Asp	Asp
															655
Phe	Val	Gly	Ala	Leu	Ile	Phe	Ser	Gly	Ala	Val	Ile	Leu	Leu	Glu	Phe
															670

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Ile Pro Glu Ile Ala Ile Pro Val Leu Gly Thr Phe Ala Leu Val Ser  
 675 680 685  
 Tyr Ile Ala Asn Lys Val Leu Thr Val Gln Thr Ile Asp Asn Ala Leu  
 690 695 700  
 Ser Lys Arg Asn Glu Lys Trp Asp Glu Val Tyr Lys Tyr Ile Val Thr  
 705 710 715 720  
 Asn Trp Leu Ala Lys Val Asn Thr Gln Ile Asp Leu Ile Arg Lys Lys  
 725 730 735  
 Met Lys Glu Ala Leu Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile  
 740 745 750  
 Asn Tyr Gln Tyr Asn Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn  
 755 760 765  
 Phe Asn Ile Asp Asp Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys  
 770 775 780  
 Ala Met Ile Asn Ile Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr  
 785 790 795 800  
 Leu Met Asn Ser Met Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe  
 805 810 815  
 Asp Ala Ser Leu Lys Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg  
 820 825 830  
 Gly Thr Leu Ile Gly Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn  
 835 840 845  
 Thr Leu Ser Thr Asp Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn  
 850 855 860  
 Gln Arg Leu Leu Ser Thr Phe Thr Glu Tyr Ile Lys Asn Ile Ile Asn  
 865 870 875 880  
 Thr Ser Ile Leu Asn Leu Arg Tyr Glu Ser Asn His Leu Ile Asp Leu  
 885 890 895  
 Ser Arg Tyr Ala Ser Lys Ile Asn Ile Gly Ser Lys Val Asn Phe Asp  
 900 905 910  
 Pro Ile Asp Lys Asn Gln Ile Gln Leu Phe Asn Leu Glu Ser Ser Lys  
 915 920 925  
 Ile Glu Val Ile Leu Lys Asn Ala Ile Val Tyr Asn Ser Met Tyr Glu  
 930 935 940  
 Asn Phe Ser Thr Ser Phe Trp Ile Arg Ile Pro Lys Tyr Phe Asn Ser  
 945 950 955 960  
 Ile Ser Leu Asn Asn Glu Tyr Thr Ile Ile Asn Cys Met Glu Asn Asn  
 965 970 975  
 Ser Gly Trp Lys Val Ser Leu Asn Tyr Gly Glu Ile Ile Trp Thr Leu  
 980 985 990  
 Gln Asp Thr Gln Glu Ile Lys Gln Arg Val Val Phe Lys Tyr Ser Gln  
 995 1000 1005  
 Met Ile Asn Ile Ser Asp Tyr Ile Asn Arg Trp Ile Phe Val Thr  
 1010 1015 1020  
 Ile Thr Asn Asn Arg Leu Asn Asn Ser Lys Ile Tyr Ile Asn Gly  
 1025 1030 1035  
 Arg Leu Ile Asp Gln Lys Pro Ile Ser Asn Leu Gly Asn Ile His  
 1040 1045 1050  
 Ala Ser Asn Asn Ile Met Phe Lys Leu Asp Gly Cys Arg Asp Thr  
 1055 1060 1065  
 His Arg Tyr Ile Trp Ile Lys Tyr Phe Asn Leu Phe Asp Lys Glu  
 1070 1075 1080

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Leu Asn Glu Lys Glu Ile Lys Asp Leu Tyr Asp Asn Gln Ser Asn  
 1085 1090  
 Ser Gly Ile Leu Lys Asp Phe Trp Gly Asp Tyr Leu Gln Tyr Asp  
 1100 1105 1110  
 Lys Pro Tyr Tyr Met Leu Asn Leu Tyr Asp Pro Asn Lys Tyr Val  
 1115 1120 1125  
 Asp Val Asn Asn Val Gly Ile Arg Gly Tyr Met Tyr Leu Lys Gly  
 1130 1135 1140  
 Pro Arg Gly Ser Val Met Thr Thr Asn Ile Tyr Leu Asn Ser Ser  
 1145 1150 1155  
 Leu Tyr Arg Gly Thr Lys Phe Ile Ile Lys Lys Tyr Ala Ser Gly  
 1160 1165 1170  
 Asn Lys Asp Asn Ile Val Arg Asn Asn Asp Arg Val Tyr Ile Asn  
 1175 1180 1185  
 Val Val Val Lys Asn Lys Glu Tyr Arg Leu Ala Thr Asn Ala Ser  
 1190 1195 1200  
 Gln Ala Gly Val Glu Lys Ile Leu Ser Ala Leu Glu Ile Pro Asp  
 1205 1210 1215  
 Val Gly Asn Leu Ser Gln Val Val Val Met Lys Ser Lys Asn Asp  
 1220 1225 1230  
 Gln Gly Ile Thr Asn Lys Cys Lys Met Asn Leu Gln Asp Asn Asn  
 1235 1240 1245  
 Gly Asn Asp Ile Gly Phe Ile Gly Phe His Gln Phe Asn Asn Ile  
 1250 1255 1260  
 Ala Lys Leu Val Ala Ser Asn Trp Tyr Asn Arg Gln Ile Glu Arg  
 1265 1270 1275  
 Ser Ser Arg Thr Leu Gly Cys Ser Trp Glu Phe Ile Pro Val Asp  
 1280 1285 1290  
 Asp Gly Trp Gly Glu Arg Pro Leu  
 1295 1300

&lt;210&gt; SEQ ID NO 23

&lt;211&gt; LENGTH: 1301

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Clostridium botulinum serotype A neurotoxin,  
chimera 9

&lt;400&gt; SEQUENCE: 23

Met Pro Phe Val Asn Lys Gln Phe Asn Tyr Lys Asp Pro Val Asn Gly  
 1 5 10 15  
 Val Asp Ile Ala Tyr Ile Lys Ile Pro Asn Ala Gly Gln Met Gln Pro  
 20 25 30  
 Val Lys Ala Phe Lys Ile His Asn Lys Ile Trp Val Ile Pro Glu Arg  
 35 40 45  
 Asp Thr Phe Thr Asn Pro Glu Glu Gly Asp Leu Asn Pro Pro Pro Glu  
 50 55 60  
 Ala Lys Gln Val Pro Val Ser Tyr Tyr Asp Ser Thr Tyr Leu Ser Thr  
 65 70 75 80  
 Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp Glu Gln Gly  
 85 90 95  
 Glu Gln Leu Glu Arg Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly  
 100 105 110  
 Arg Gly Ile Pro Phe Trp Gly Gly Ser Thr Ile Asp Thr Glu Leu Lys  
 115 120 125

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Val Ile Asp Thr Asn Cys Ile Asn Val Ile Gln Pro Asp Gly Ser Tyr  
 130 135 140

Arg Ser Glu Glu Leu Asn Leu Val Ile Ile Gly Pro Ser Ala Asp Ile  
 145 150 155 160

Ile Gln Phe Glu Cys Lys Ser Phe Gly His Glu Val Leu Asn Leu Thr  
 165 170 175

Arg Asn Gly Tyr Gly Ser Thr Gln Tyr Ile Arg Phe Ser Pro Asp Phe  
 180 185 190

Thr Phe Gly Phe Glu Glu Ser Leu Glu Val Asp Thr Asn Pro Leu Leu  
 195 200 205

Gly Ala Gly Lys Phe Ala Thr Asp Pro Ala Val Thr Leu Ala His Ala  
 210 215 220

Leu Ile His Ala Gly His Arg Leu Tyr Gly Ile Ala Ile Asn Pro Asn  
 225 230 235 240

Arg Val Phe Lys Val Asn Thr Asn Ala Tyr Tyr Glu Met Ser Gly Leu  
 245 250 255

Glu Val Ser Phe Glu Glu Leu Arg Thr Phe Gly Gly His Asp Ala Lys  
 260 265 270

Phe Ile Asp Glu Leu Asp Asp Arg Ala Asp Ala Leu Gln Met Leu Asp  
 275 280 285

Glu Gln Gly Glu Gln Leu Glu Arg Leu Asn Lys Ala Lys Ser Ile Val  
 290 295 300

Gly Thr Thr Ala Ser Glu Met Asp Glu Asn Leu Glu Gln Val Ser Gly  
 305 310 315 320

Arg Leu Leu Ser Glu Asp Thr Ser Gly Lys Phe Ser Val Asp Phe Met  
 325 330 335

Asp Glu Phe Phe Glu Gln Val Glu Glu Leu Thr Glu Ile Tyr Thr Glu  
 340 345 350

Leu Glu Asp Met Leu Glu Ser Gly Asn Leu Asn Arg Lys Thr Tyr Leu  
 355 360 365

Asn Phe Asp Lys Ala Val Phe Lys Ile Asn Ile Val Pro Lys Val Asn  
 370 375 380

Tyr Thr Ile Tyr Asp Gly Phe Asn Leu Arg Asn Thr Asn Leu Ala Ala  
 385 390 395 400

Asn Phe Asn Gly Gln Asn Thr Glu Ile Asn Asn Met Asn Phe Thr Lys  
 405 410 415

Leu Lys Asn Phe Thr Gly Leu Phe Glu Phe Tyr Lys Leu Leu Cys Val  
 420 425 430

Arg Gly Ile Ile Thr Ser His Thr Gln Ser Leu Asp Gln Gly Tyr Asn  
 435 440 445

Asp Asp Asp Asp Lys Ala Leu Asn Asp Leu Cys Ile Lys Val Asn Asn  
 450 455 460

Trp Asp Leu Phe Phe Ser Pro Ser Glu Asp Asn Phe Thr Asn Asp Leu  
 465 470 475 480

Asn Lys Gly Glu Glu Ile Thr Ser Asp Thr Asn Ile Glu Ala Ala Glu  
 485 490 495

Glu Asn Ile Ser Leu Asp Leu Ile Gln Gln Tyr Tyr Leu Thr Phe Asn  
 500 505 510

Phe Asp Asn Glu Pro Glu Asn Ile Ser Ile Glu Asn Leu Ser Ser Asp  
 515 520 525

Ile Ile Gly Gln Leu Glu Leu Met Pro Asn Ile Glu Arg Phe Pro Asn  
 530 535 540

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Gly Lys Lys Tyr Glu Leu Asp Lys Tyr Thr Met Phe His Tyr Leu Arg  
 545 550 555 560  
 Ala Gln Glu Phe Glu His Gly Lys Ser Arg Ile Ala Leu Thr Asn Ser  
 565 570 575  
 Val Asn Glu Ala Leu Leu Asn Pro Ser Arg Val Tyr Thr Phe Phe Ser  
 580 585 590  
 Ser Asp Tyr Val Lys Lys Val Asn Lys Ala Thr Glu Ala Ala Met Phe  
 595 600 605  
 Leu Gly Trp Val Glu Gln Leu Val Tyr Asp Phe Thr Asp Glu Thr Ser  
 610 615 620  
 Glu Val Ser Thr Thr Asp Lys Ile Ala Asp Ile Thr Ile Ile Ile Pro  
 625 630 635 640  
 Tyr Ile Gly Pro Ala Leu Asn Ile Gly Asn Met Leu Tyr Lys Asp Asp  
 645 650 655  
 Phe Val Gly Ala Leu Ile Phe Ser Gly Ala Val Ile Leu Leu Glu Phe  
 660 665 670  
 Ile Pro Glu Ile Ala Ile Pro Val Leu Gly Thr Phe Ala Leu Val Ser  
 675 680 685  
 Tyr Ile Ala Asn Lys Val Leu Thr Val Gln Thr Ile Asp Asn Ala Leu  
 690 695 700  
 Ser Lys Arg Asn Glu Lys Trp Asp Glu Val Tyr Lys Tyr Ile Val Thr  
 705 710 715 720  
 Asn Trp Leu Ala Lys Val Asn Thr Gln Ile Asp Leu Ile Arg Lys Lys  
 725 730 735  
 Met Lys Glu Ala Leu Glu Asn Gln Ala Glu Ala Thr Lys Ala Ile Ile  
 740 745 750  
 Asn Tyr Gln Tyr Asn Gln Tyr Thr Glu Glu Glu Lys Asn Asn Ile Asn  
 755 760 765  
 Phe Asn Ile Asp Asp Leu Ser Ser Lys Leu Asn Glu Ser Ile Asn Lys  
 770 775 780  
 Ala Met Ile Asn Ile Asn Lys Phe Leu Asn Gln Cys Ser Val Ser Tyr  
 785 790 795 800  
 Leu Met Asn Ser Met Ile Pro Tyr Gly Val Lys Arg Leu Glu Asp Phe  
 805 810 815  
 Asp Ala Ser Leu Lys Asp Ala Leu Leu Lys Tyr Ile Tyr Asp Asn Arg  
 820 825 830  
 Gly Thr Leu Ile Gly Gln Val Asp Arg Leu Lys Asp Lys Val Asn Asn  
 835 840 845  
 Thr Leu Ser Thr Asp Ile Pro Phe Gln Leu Ser Lys Tyr Val Asp Asn  
 850 855 860  
 Gln Arg Leu Leu Ser Thr Phe Thr Glu Tyr Ile Lys Asn Ile Ile Asn  
 865 870 875 880  
 Thr Ser Ile Leu Asn Leu Arg Tyr Glu Ser Asn His Leu Ile Asp Leu  
 885 890 895  
 Ser Arg Tyr Ala Ser Lys Ile Asn Ile Gly Ser Lys Val Asn Phe Asp  
 900 905 910  
 Pro Ile Asp Lys Asn Gln Ile Gln Leu Phe Asn Leu Glu Ser Ser Lys  
 915 920 925  
 Ile Glu Val Ile Leu Lys Asn Ala Ile Val Tyr Asn Ser Met Tyr Glu  
 930 935 940  
 Asn Phe Ser Thr Ser Phe Trp Ile Arg Ile Pro Lys Tyr Phe Asn Ser  
 945 950 955 960  
 Ile Ser Leu Asn Asn Glu Tyr Thr Ile Ile Asn Cys Met Glu Asn Asn



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 <223> OTHER INFORMATION: Metalloprotease motif  
 <220> FEATURE:  
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 <223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
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 <223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

<400> SEQUENCE: 25

His Glu Xaa Xaa His Xaa Xaa His  
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<210> SEQ ID NO 26  
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 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
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<400> SEQUENCE: 26

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<210> SEQ ID NO 27  
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 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: annealed phosphorylated linker CBA02

<400> SEQUENCE: 27

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<210> SEQ ID NO 28  
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 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA03

<400> SEQUENCE: 28

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<210> SEQ ID NO 29  
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 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA04

<400> SEQUENCE: 29

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<210> SEQ ID NO 30  
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 <212> TYPE: DNA  
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 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA05

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<400> SEQUENCE: 30  
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<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: oligonucleotide CBA06

<400> SEQUENCE: 31  
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ctgtatddd acc 73

<210> SEQ ID NO 32  
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<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: annealed phosphorylated linker CBA08

<400> SEQUENCE: 32  
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<220> FEATURE:  
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<213> ORGANISM: Artificial  
<220> FEATURE:  
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<400> SEQUENCE: 35  
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<210> SEQ ID NO 36  
<211> LENGTH: 89  
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<213> ORGANISM: Artificial  
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<223> OTHER INFORMATION: oligonucleotide CBA12

<400> SEQUENCE: 36  
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gttaataata cacttagtac agatatacc 89



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 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA13  
  
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 tcatctacag gaataaatc 80

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 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA14  
  
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 tatcaaagtt aataattggg 80

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 <223> OTHER INFORMATION: oligonucleotide CBA16  
  
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 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA17  
  
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 gac 63

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 <220> FEATURE:  
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<400> SEQUENCE: 44

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<210> SEQ ID NO 45  
 <211> LENGTH: 34  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: gp64 signal peptide and hexahistadine affinity tag

<400> SEQUENCE: 45

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Ala His Ser Ala Phe Ala Ala Met Val His His His His His His Ser  
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Ala Ser

<210> SEQ ID NO 46  
 <211> LENGTH: 38  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
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<400> SEQUENCE: 46

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<210> SEQ ID NO 47  
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 <220> FEATURE:  
 <223> OTHER INFORMATION: phosphorylated oligonucleotide CBA19

<400> SEQUENCE: 47

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<210> SEQ ID NO 48  
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 <223> OTHER INFORMATION: oligonucleotide CBA20

<400> SEQUENCE: 48

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 <223> OTHER INFORMATION: oligonucleotide CBA21  
  
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<210> SEQ ID NO 50  
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 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: oligonucleotide CBA22  
  
 <400> SEQUENCE: 50  
  
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<210> SEQ ID NO 51  
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 <212> TYPE: DNA  
 <213> ORGANISM: Artificial  
 <220> FEATURE:  
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<210> SEQ ID NO 52  
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 <220> FEATURE:  
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 acttaaatat gttgaatcat aatatgaaac tgg 93

<210> SEQ ID NO 53  
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 <213> ORGANISM: Artificial  
 <220> FEATURE:  
 <223> OTHER INFORMATION: phosphorylated oligonucleotide CBCH2  
  
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 ggaagtacaa tagatacag 79

<210> SEQ ID NO 54  
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 <220> FEATURE:  
 <223> OTHER INFORMATION: phosphorylated oligonucleotide CBCH3  
  
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<210> SEQ ID NO 55

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 gtaggtacta ctgc 74  
  
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 <212> TYPE: DNA  
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 <220> FEATURE:  
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 <220> FEATURE:  
 <223> OTHER INFORMATION: phosphorylated oligonucleotide CBCH6  
  
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 <211> LENGTH: 32  
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 <213> ORGANISM: Artificial  
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 <212> TYPE: DNA  
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 <212> TYPE: DNA  
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 <220> FEATURE:  
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 <220> FEATURE:  
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&lt;400&gt; SEQUENCE: 61

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&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: oligonucleotide EGFPs

&lt;400&gt; SEQUENCE: 62

tattacgcgt gcgcgctatg aattctataa gttgctaag gtgagcaagg gcgaggagct 60

gttcaccggg 70

&lt;210&gt; SEQ ID NO 63

&lt;211&gt; LENGTH: 49

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: oligonucleotide EGFPa

&lt;400&gt; SEQUENCE: 63

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&lt;210&gt; SEQ ID NO 64

&lt;211&gt; LENGTH: 6

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: cytosolic protein motif

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (4)..(5)

&lt;223&gt; OTHER INFORMATION: Xaa can be any naturally occurring amino acid

&lt;400&gt; SEQUENCE: 64

Asp Ser Gly Xaa Xaa Ser

1 5

&lt;210&gt; SEQ ID NO 65

&lt;211&gt; LENGTH: 3888

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Clostridium botulinum (serotype A) wt fragment

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (4)..(30)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (40)..(648)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

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&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (658)..(1269)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

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&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1348)..(1359)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1363)..(1785)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

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&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1807)..(2484)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1807)..(2484)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

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&lt;222&gt; LOCATION: (2491)..(3873)

&lt;223&gt; OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 1

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<221> NAME/KEY: misc_feature
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<223> OTHER INFORMATION: n is nucleotide encoding SEQ ID NO: 8
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What is claimed:

1. An isolated, physiologically active Clostridial neuro-<sup>60</sup>  
toxin comprising:  
a light chain terminating at its C-terminus with a lysine  
residue of a highly specific protease cleavage site comprising three or more specific adjacent amino acid resi-<sup>65</sup>  
dues that are recognized by a highly specific protease to  
enable cleavage and

a heavy chain, wherein the light chain and the heavy chain  
are linked by a disulfide bond.  
2. The isolated Clostridial neurotoxin according to claim 1,  
wherein the neurotoxin is from *Clostridium botulinum*.  
3. The isolated Clostridial neurotoxin according to claim 2,  
wherein the *Clostridium botulinum* has a serotype selected  
from the group consisting of *Clostridium botulinum* serotype  
A, *Clostridium botulinum* serotype B, *Clostridium botulinum*  
serotype C, *Clostridium botulinum* serotype D, *Clostridium*

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*botulinum* serotype E, *Clostridium botulinum* serotype F, and *Clostridium botulinum* serotype G.

4. The isolated Clostridial neurotoxin according to claim 1, wherein said physiological activity is selected from the group consisting of toxin immunogenicity, trans- and intra-cellular trafficking, and cell recognition.

5 5. The isolated Clostridial neurotoxin according to claim 1, wherein the neurotoxin is toxic.

6. The isolated Clostridial neurotoxin according to claim 1, wherein the neurotoxin is atoxic.

10 7. The isolated Clostridial neurotoxin according to claim 6, wherein the neurotoxin has a disabling mutation in an active metalloprotease site.

8. The isolated Clostridial neurotoxin according to claim 6, wherein the light chain comprises a non-native motif capable of inactivating light chain metalloprotease activity in a toxic Clostridial neurotoxin.

15 9. The isolated Clostridial neurotoxin according to claim 1, wherein the neurotoxin further comprises:

a signal peptide coupled to the light chain, wherein the signal peptide is suitable to permit secretion of the neurotoxin from a eukaryotic cell to a medium and an affinity tag located between the signal peptide and the light chain.

20 10. The isolated Clostridial neurotoxin according to claim 9, wherein the affinity tag is a hexahistidine affinity tag.

11. The isolated Clostridial neurotoxin according to claim 10, wherein the affinity tag comprises SEQ ID NO: 45.

12. The isolated Clostridial neurotoxin according to claim 1, wherein the heavy chain comprises a modification at a Lys amino acid residue adjacent to a receptor binding domain.

13. The isolated Clostridial neurotoxin according to claim 12, wherein the modification is a Lys to Asn substitution.

14. The isolated Clostridial neurotoxin according to claim 1, wherein the highly specific protease cleavage site is an enterokinase cleavage site of SEQ ID NO:24.

15. A vaccine or antidote comprising the isolated Clostridial neurotoxin according to claim 6.

16. The vaccine or antidote according to claim 15, wherein the neurotoxin is from *Clostridium botulinum*.

17. The vaccine or antidote according to claim 16, wherein the *Clostridium botulinum* has a serotype selected from the group consisting of *Clostridium botulinum* serotype A, *Clostridium botulinum* serotype B, *Clostridium botulinum* serotype C, *Clostridium botulinum* serotype D, *Clostridium botulinum* serotype E, *Clostridium botulinum* serotype F, and *Clostridium botulinum* serotype G.

18. The vaccine or antidote according to claim 15, wherein the neurotoxin has a disabling mutation in an active metalloprotease site.

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19. The vaccine or antidote according to claim 15, wherein said physiological activity is selected from the group consisting of toxin immunogenicity, trans- and intra-cellular trafficking, and cell recognition.

20. A method of immunizing a subject against toxic effects of a Clostridial neurotoxin, said method comprising:

administering the vaccine according to claim 15 to the subject under conditions effective to immunize the subject against toxic effects of Clostridial neurotoxin.

10 21. The method according to claim 20, wherein the neurotoxin is from *Clostridium botulinum*.

22. The method according to claim 21, wherein the *Clostridium botulinum* has a serotype selected from the group consisting of *Clostridium botulinum* serotype A, *Clostridium botulinum* serotype B, *Clostridium botulinum* serotype C, *Clostridium botulinum* serotype D, *Clostridium botulinum* serotype E, *Clostridium botulinum* serotype F, and *Clostridium botulinum* serotype G.

15 23. The method according to claim 20, wherein the neurotoxin has a disabling mutation in an active metalloprotease site.

24. The method according to claim 20 further comprising: administering a booster of the vaccine to the subject under conditions effective to enhance immunization of the subject.

25. A method of treating a subject for toxic effects of a Clostridial neurotoxin, said method comprising:

administering the antidote according to claim 15 to the subject under conditions effective to treat the subject for toxic effects of Clostridial neurotoxin.

26. The method according to claim 25, wherein the neurotoxin is from *Clostridium botulinum*.

27. The method according to claim 26, wherein the *Clostridium botulinum* has a serotype selected from the group consisting of *Clostridium botulinum* serotype A, *Clostridium botulinum* serotype B, *Clostridium botulinum* serotype C, *Clostridium botulinum* serotype D, *Clostridium botulinum* serotype E, *Clostridium botulinum* serotype F, and *Clostridium botulinum* serotype G.

28. The method according to claim 25, wherein the neurotoxin has a disabling mutation in an active metalloprotease site.

29. A treatment method comprising:

contacting a patient with an isolated Clostridial neurotoxin according to claim 5 under conditions effective to treat the patient.

\* \* \* \* \*